

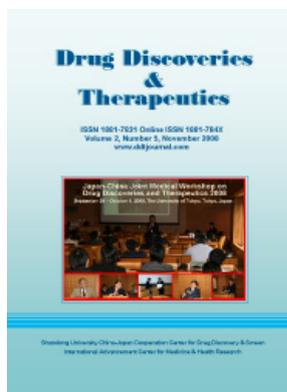
Drug Discoveries & Therapeutics

ISSN 1881-7831 Online ISSN 1881-784X
Volume 2, Number 5, November 2008
www.ddtjournal.com



Shandong University China-Japan Cooperation Center for Drug Discovery & Screen
International Advancement Center for Medicine & Health Research

Drug Discoveries & Therapeutics



Editor-in-Chief:

Kazuhisa SEKIMIZU
(The University of Tokyo, Tokyo, Japan)

Associate Editor:

Norihiro KOKUDO
(The University of Tokyo, Tokyo, Japan)

Drug Discoveries & Therapeutics is a peer-reviewed international journal published bimonthly by *Shandong University China-Japan Cooperation Center for Drug Discovery & Screen (SDU-DDSC)* and *International Advancement Center for Medicine & Health Research Co., Ltd. (IACMHR Co., Ltd.)*.

Drug Discoveries & Therapeutics mainly publishes articles related to basic and clinical pharmaceutical research such as pharmaceutical and therapeutical chemistry, pharmacology, pharmacy, pharmacokinetics, industrial pharmacy, pharmaceutical manufacturing, pharmaceutical technology, drug delivery, toxicology, and traditional herb medicine. Studies on drug-related fields such as biology, biochemistry, physiology, microbiology, and immunology are also within the scope of this journal.

Subject Coverage: Basic and clinical pharmaceutical research including Pharmaceutical and therapeutical chemistry, Pharmacology, Pharmacy, Pharmacokinetics, Industrial pharmacy, Pharmaceutical manufacturing, Pharmaceutical technology, Drug delivery, Toxicology, and Traditional herb medicine.

Language: English

Issues/Year: 6

Published by: IACMHR and SDU-DDSC

ISSN: 1881-7831 (Online ISSN 1881-784X)

Editorial and Head Office

Wei TANG, MD PhD
Secretary-in-General

TSUIN-IKIZAKA 410
2-17-5 Hongo, Bunkyo-ku
Tokyo 113-0033, Japan
Tel: 03-5840-9697
Fax: 03-5840-9698
E-mail: office@ddtjournal.com
URL: www.ddtjournal.com



Drug Discoveries & Therapeutics

Editorial Board

Editor-in-Chief:

Kazuhisa SEKIMIZU (*The University of Tokyo, Tokyo, Japan*)

Associate Editor:

Norihiro KOKUDO (*The University of Tokyo, Tokyo, Japan*)

Secretary-in-General:

Wei TANG (*The University of Tokyo, Tokyo, Japan*)

Office Manager:

Munehiro NAKATA (*Tokai University, Kanagawa, Japan*)

Web Editor:

Yu CHEN (*The University of Tokyo, Tokyo, Japan*)

English Editor:

Curtis BENTLEY (*Roswell, GA, USA*)

China Office:

Wenfang XU (*Shandong University, Shandong, China*)

Editors:

Yoshihiro ARAKAWA (*Tokyo, Japan*)
Santad CHANPRAPAPH (*Bangkok, Thailand*)
Fen Er CHEN (*Shanghai, China*)
Zilin CHEN (*Wuhan, China*)
Guanhua DU (*Beijing, China*)
Chandradhar DWIVEDI (*Brookings, SD, USA*)
Mohamed F. EL-MILIGI (*Cairo, Egypt*)
Harald HAMACHER (*Tuebingen, Germany*)
Hiroshi HAMAMOTO (*Tokyo, Japan*)
Xiao-Jiang HAO (*Kunming, China*)
Langchong HE (*Xi'an, China*)
David A. HORNE (*Duarte, CA, USA*)
Yongzhou HU (*Hangzhou, China*)
Wei HUANG (*Shanghai, China*)
Yu HUANG (*Hong Kong, China*)
Hans E. JUNGINGER (*Phitsanulok, Thailand*)
Toshiaki KATADA (*Tokyo, Japan*)
Ibrahim S. KHATTAB (*Safat, Kuwait*)
Hiromichi KIMURA (*Tokyo, Japan*)
Shiroh KISHIOKA (*Wakayama, Japan*)
Kam Ming KO (*Hong Kong, China*)
Nobuyuki KOBAYASHI (*Nagasaki, Japan*)
Toshiro KONISHI (*Tokyo, Japan*)
Masahiro KUROYANAGI (*Hiroshima, Japan*)
Chun Guang LI (*Victoria, Australia*)
Hongmin LIU (*Zhengzhou, China*)
Ji-Kai LIU (*Kunming, China*)

Hongxiang LOU (*Jinan, China*)
Ken-ichi MAFUNE (*Tokyo, Japan*)
Norio MATSUKI (*Tokyo, Japan*)
Tohru MIZUSHIMA (*Kumamoto, Japan*)
Abdulla M. MOLOKHIA (*Alexandria, Egypt*)
Masahiro MURAKAMI (*Osaka, Japan*)
Yoshinobu NAKANISHI (*Ishikawa, Japan*)
Yutaka ORIHARA (*Tokyo, Japan*)
Xiao-Ming OU (*Jackson, MS, USA*)
Wei-San PAN (*Shenyang, China*)
Shafiqur RAHMAN (*Brookings, SD, USA*)
Adel SAKR (*Cincinnati, OH, USA*)
Abdel Aziz M. SALEH (*Cairo, Egypt*)
Tomofumi SANTA (*Tokyo, Japan*)
Yasufumi SAWADA (*Tokyo, Japan*)
Brahma N. SINGH (*Commack, NY, USA*)
Hongbin SUN (*Nanjing, China*)
Benny K. H. TAN (*Singapore, Singapore*)
Ren-Xiang TAN (*Nanjing, China*)
Murat TURKOGLU (*Istanbul, Turkey*)
Stephen G. WARD (*Bath, UK*)
Takako YOKOZAWA (*Toyama, Japan*)
Liangren ZHANG (*Beijing, China*)
Jian-Ping ZUO (*Shanghai, China*)

(as of September 29, 2008)

News

- 262 - 263 **Japan-China Joint Medical Workshop on Drug Discoveries and Therapeutics 2008: The need of Asian pharmaceutical researchers' cooperation.**

Munehiro Nakata, Wei Tang

Review

- 264 - 276 **Pulmonary drug delivery: Implication for new strategy for pharmacotherapy for neurodegenerative disorders.**

Nazrul Islam, Shafiqur Rahman

Brief Reports

- 277 - 281 **Effect of drug-polymer binary mixtures on the *in-vitro* release of ibuprofen from transdermal drug-in-adhesive layers.**

Kwong Yat Ho, Michael Ord, Kalliopi Dodou

- 282 - 285 **Effect of benzyl-*N*-acetyl- α -galactosaminide on KL-6 mucin expression and invasive properties of a human pancreatic carcinoma cell line.**

Huanli Xu, Yoshinori Inagaki, Fengshan Wang, Norihiro Kokudo, Munehiro Nakata, Wei Tang

- 286 - 288 **Antioxidant activity of wild plants collected in Beni-Sueif governorate, Upper Egypt.**

Sameh AbouZid, Abdelaaty Elshahaat, Sajjad Ali, Mohamed I. Choudhary

Original Articles

- 289 - 295 **The neuroprotective effect of antidepressant drug *via* inhibition of TIEG2-MAO B mediated cell death.**

Deyin Lu, Chandra Johnson, Shakevia Johnson, Shawna Tazik, Xiao-Ming Ou

- 296 - 304** **Regulation of the nitric oxide synthesis pathway and cytokine balance contributes to the healing action of *Myristica malabarica* against indomethacin-induced gastric ulceration in mice.**

*Biswanath Maity, Debashish Banerjee, Sandip K. Bandyopadhyay,
Subrata Chattopadhyay*

Guide for Authors

Copyright

Japan-China Joint Medical Workshop on Drug Discoveries and Therapeutics 2008: The need of Asian pharmaceutical researchers' cooperation

Munehiro Nakata¹, Wei Tang²

Keywords: Drug discovery, Therapeutic, Influenza

The Japan-China Joint Medical Workshop on Drug Discoveries and Therapeutics 2008 (JCMWDDT 2008) was held from September 29 to October 1, 2008 at The University of Tokyo, Tokyo, Japan. JCMWDDT is an international workshop that is mainly organized by Asian editorial members of Drug Discoveries & Therapeutics (<http://www.ddtjournal.com/home>) for the purpose of promoting research exchanges in the field of drug discovery and therapeutic. This year's JCMWDDT is the second workshop and focused particularly on novel development and technological innovation of anti-influenza agents. The workshop began with an announcement by the Japanese Co-chairperson, Dr. Sekimizu (Department of Microbiology, Graduate School of Pharmaceutical Sciences, The University of Tokyo, Japan; Editor-in-Chief of Drug Discoveries & Therapeutics, DDT) followed by a speech by the Chinese Co-chairperson, Dr. Wenfang Xu (School of Pharmaceutical Sciences, Shandong University, Shandong, China; Editor in China Office of DDT), with additional speeches by Dr. Norio Matsuki (The University of Tokyo, Japan; Editor of DDT) and Dr. Guanhua Du (Chinese Academy of Medical Science, China; Editor of DDT). Fifty-nine titles were presented in 6 specialized sessions (Research Advances in Drug Discoveries and Therapeutics, Drug Synthesis/Clinical Therapeutics, Medicinal Chemistry/Natural Products, Anti-influenza Drugs, Anti-infection/antiviral Drugs, Biochemistry/Molecular Biology /Pharmacology) and a poster session (*Drug Discov Ther 2008; 2, Suppl; available at <http://www.ddtjournal.com/Announce/index.htm>*). An annual outbreak of avian influenza in Asian countries including China and Japan has sparked fears that the virus will mutate and then cause an epidemic in humans. Therefore, Asian researchers need to work together to control this infection. This year's JCMWDDT helped provide an opportunity to reiterate the crucial role of medicinal chemistry in conquering influenza and created an environment for cooperative research in Asian countries. (reported on October 1st, with grateful thanks to all participants)



Main program

Session I. Research Advances in Drug Discoveries and Therapeutics

- Design, synthesis and preliminary activity assay of influenza virus neuraminidase inhibitors by *Wenfang Xu (Shandong University, China)*
- Infection disease models with silkworms to evaluate the therapeutic effects of drug candidates by *Kazuhisa Sekimizu (The University of Tokyo, Japan)*
- Japan's governmental approaches to facilitate drug development process by *Makoto Shimoaraiso (Ministry of Foreign Affairs of Japan, Japan)*
- Effective detection of the epidermal growth factor receptor mutation by the peptide nucleic acid-locked nucleic acid PCR Clamp by *Sakuo Hoshi (The University of Tokyo Hospital, Japan)*
- Design and synthesis of p53-MDM2 binding inhibitors by *Yongzhou Hu (Zhejiang University, China)*

Session II. Drug Synthesis/Clinical Therapeutics

- Pharmacogenomics-based clinical studies using a novel fully-automated genotyping system by *Setsuo Hasegawa (Sekino Clinical Pharmacology Clinic, Japan)*
- Synthesis and biological evaluation of pentacyclic triterpenes as anti-tumor agents by *Hongbin Sun (China Pharmaceutical University, China)*
- Drug discovery and therapeutics using silkworm as experimental animal by *Yasuyuki Ogata (The University of*

Tokyo, Japan)

- Novel selective estrogen receptor modulators (SERMs) with unusual structure and biological activities by Haibing Zhou (Wuhan University, China)

Session III. Medicinal Chemistry/Natural Products

- Synthesis and properties of isonucleosides incorporated oligonucleotides by Zhenjun Yang (Peking University, China)
- Isolation of antiviral compounds from plant resources using silkworm bioassay by Yutaka Orihara (The University of Tokyo, Japan)
- Synthesis and structural modification of tasiamide and the effect of these modifications on *in vitro* anticancer activity by Yingxia Li (Ocean University of China, China)
- Spirohexalines A and B, novel undecaprenyl pyrophosphate inhibitors produced by *Penicillium* sp. FK1-3368 by Junji Inokoshi (Kitasato University, Japan)
- Nosokomyocins, novel anti-MRSA antibiotics, produced by *Streptomyces* sp. K04-0144 by OR. Uchida (Kitasato University, Japan)
- *In vivo* screening for antimicrobial activity of Thai Herbal Medicines using silkworm model by Santad Chanprapaph (Chulalongkorn University, Thailand)
- Novel electrochemical sensor of nitric oxide for screening anti-aging Traditional Chinese Medicine by Zilin Chen (Wuhan University, China)
- Polysaccharide from green tea purified by silkworm muscle contraction assay induces innate immunity by increasing the expression of various inflammatory cytokine mRNA in human leukocytes by Saphala Dhital (The University of Tokyo, Japan)

Session IV. Anti-influenza Drugs

- Structure-activity relationship of flavonoids as influenza virus neuraminidase inhibitors and their *in vitro* anti-viral activities by Guanhua Du (Chinese Academy of Medical Sciences and Peking Union Medical College, China)
- Mechanisms and consequences of phagocytosis of influenza virus-infected cells by Yoshinobu Nakanishi (Kanazawa University, Japan)
- Nuclear export inhibitors; a possible target for novel anti-influenza viral drugs by Ken Watanabe (Nagasaki University, Japan)
- Catalytic asymmetric synthesis of oseltamivir phosphate directing toward its stable worldwide supply by Motomu Kanai (The University of Tokyo, Japan)
- Clinical effects of probiotic bifidobacterium in the prevention of influenza virus infections and allergic diseases by Jin-zhong Xiao (Morinaga Milk Industry Co., Ltd., Japan)
- Production of anti-influenza PR8-scFv using a phage display by Normaiza Zamri (Tokai University, Japan)

Session V. Anti-infection/Antiviral Drugs

- Emerging infectious diseases and anti-viral drugs: Urgent need to develop effective drugs which cause less resistant

virus by Nobuyuki Kobayashi (Nagasaki University, Japan)

- Design, synthesis and antiviral evaluation of novel heterocyclic compounds as HIV-1 NNRTIs by Xinyong Liu (Shandong University, China)
- Antiviral drug screening from microbial products by Eisaku Tsujii (Astellas Pharma Inc., Japan)
- Viral factors that determine the natural course of chronic hepatitis B viral infection by Hiroshi Yotsuyanagi (The University of Tokyo, Japan)
- Effect of andrographolide derivatives having α -glucosidase inhibition, on HBsAg, HBeAg secretion in HepG2 2.2.15 cells by Hongmin Liu (Zhengzhou University, China)
- Current and future antiviral therapy for influenza by Hideki Asanuma (Tokai University, Japan)
- Establishment of an HIV-based pseudotyping system as a safe model for screening inhibitors on bird flu H5N1 entry by Ying Guo (Peking Union Medical College Chinese Academy of Medical Sciences, China)
- Strategy of discovery for novel antibiotics using silkworm infection model by Hiroshi Hamamoto (The University of Tokyo, Japan)
- Potent neuraminidase inhibitors and anti-inflammatory substances from *Chaenomeles speciosa* by Li Zhang (Chinese Academy of Medical Sciences and Peking Union Medical College, China)
- High-throughput screening assay for hepatitis C virus helicase inhibitors using fluorescence-quenching phenomenon by Hidenori Tani (Waseda University and National Institute of Advanced Industrial Science and Technology, Japan)

Session VI. Biochemistry/Molecular Biology/Pharmacology

- A novel conjugate of low-molecular-weight heparin and Cu,Zn-superoxide dismutase: Study on its mechanism in preventing brain reperfusion injury after ischemia in gerbils by Fengshan Wang (Shandong University, China)
- A novel gene *fudoh* in SCCmec region regulates the colony spreading ability and virulence in *Staphylococcus aureus* by Chikara Kaito (The University of Tokyo, Japan)
- Water soluble fluorescent boronic acid sensors for tumor cell-surface saccharide by Hao Fang (Shandong University, China)
- Molecular characterization of the biosynthetic enzyme for the biotechnological production of tetrahydrocannabinol, the active constituent of marijuana by Futoshi Taura (Kyushu University, Japan)
- Galloyl cyclic-imide derivative CH1104I inhibits tumor invasion via suppressing matrix metalloproteinase activity by Xianjun Qu (Shandong University, China)
- Neuroprotection by inhibition of GAPDH-MAO B mediated cell death induced by ethanol by Xiao-Ming Ou (University of Mississippi Medical Center, USA)

(¹ Department of Applied Biochemistry, Tokai University, Kanagawa, Japan; ² Department of Surgery, Graduate School of Medicine, The University of Tokyo, Tokyo, Japan.)

Review

Pulmonary drug delivery: Implication for new strategy for pharmacotherapy for neurodegenerative disorders

Nazrul Islam^{1,*}, Shafiqur Rahman²

¹ Department of Pharmacy, School of Life Sciences, Queensland University of Technology, Brisbane, QLD, Australia;

² Department of Pharmaceutical Sciences, College of Pharmacy, South Dakota State University, Brookings, SD, USA.

ABSTRACT: Innovative drug delivery in the treatment of brain neurodegenerative disorders such as Parkinson's disease (PD) and Alzheimer's disease (AD) has the potential to avoid many unwanted side effects over current medications. Advances in understanding of these diseases and their treatments have led to the search for novel modes of drug delivery. In this review, we have highlighted new strategies and future prospects for pulmonary delivery of drugs for the management of these important neurological disorders. The advancement of knowledge on pulmonary drug delivery will provide novel therapeutic formulations for better management of the PD and AD patients throughout the world.

Keywords: Pulmonary drug delivery, Dry powder inhaler, Metered dose inhaler, Nebulizer, Parkinson's disease, Alzheimer's disease

1. Introduction

Pulmonary delivery, a non-invasive route of drug delivery is becoming a route of choice for most drugs. Pressurized metered dose inhalers (pMDI) and nebulisers (liquid jet and ultrasonic) are the preliminary devices to deliver drugs into lung; however, currently, breath actuated dry powder inhalers (DPI) are designed to deliver medicaments as a powder form through the airways in the lung to achieve both systemic and local effects.

Direct delivery of drugs into the pulmonary regions of the lung enables lower doses with an equivalent therapeutic action compared to oral or parenteral

routes because of the large surface area (~100 m²) of the lungs. Advantages of DPI formulations over other dosage forms (*i.e.*, parenteral and other liquid dosage forms) are solid dosage form stability, ease of use, less expensive, painless and user friendly. The inhaled route allows the delivery of small doses of drug directly to the alveoli attaining a high concentration of drug in the local area and minimizes systemic side effects resulting in a high therapeutic ratio of drugs compared with that of systemic delivery administered either by oral or parenteral routes. Oral tablets and capsules need to be swallowed which is sometime difficult for some patients especially for children. Respiratory delivery also offers effective therapy with minimum adverse effects by using small doses of drugs through inhalation and allows substantially greater bioavailability of polypeptides (1).

Currently, delivery of drugs for the management of neurological disorders especially PD and AD are done by oral, parenteral and transdermal routes. Pulmonary delivery of drugs is well established in the management of asthma and COPD (chronic obstructive pulmonary disorder). However, no DPI drugs are approved yet for the management of other diseases like AD and PD. This mini-review discusses advantages of pulmonary delivery of drugs, pulmonary delivery technologies, and current situations and future trends in managing major AD and PD by delivering drugs into the deep lung *via* DPI or MDI devices.

2. Pulmonary delivery technology

Aerosol delivery of drugs, formulated as liquid solutions, suspensions, emulsions or micronized dry powders, are aerosolised *via* some commonly used different types of delivery devices (nebulizer, pMDI, and DPI). In this section both the formulations as well delivery devices are discussed.

2.1. Nebulisers

Nebulisers are probably one of the oldest forms of pulmonary drug delivery, deliver large volumes of

*Correspondence to: Dr. Nazrul Islam, Section of Pharmacy, School of Life Sciences, Queensland University of Technology, Brisbane, QLD-4001, Australia;
e-mail: nazrul.islam@qut.edu.au

drug solutions or suspensions and are frequently used for those drugs which can not be formulated into pMDIs or DPIs. Currently, two categories of nebulisers are available on the market include air jet and ultrasonic nebulisers. Air jet nebulisers can generate both smaller particles (mass median aerodynamic diameter 2-5 μm) and coarse aerosols, and deliver medication quickly; however, it produces high oropharyngeal deposition of drugs. Most jet nebulisers operate by forcing pressurised gas (air or oxygen) through a nozzle or jet at high velocity so that the nebulizer solution is atomized. On actuation the gas expands resulting in the generation of a negative pressure which draws the liquid formulation into the gas stream. The aerosol mist impacts against a baffle, drains back into the reservoir incorporated with the nebuliser and recirculates. The ultrasonic nebulisers do not require compressed gas. The solution formulation is atomised by an energy source, piezoelectric crystal transducer, which vibrates at high frequency and these devices can generate slightly larger aerosols. However, the overall efficiency of the piezoelectric driven ultrasonic nebulisers is more or less similar to that of air-jet nebulisers. Patients who are seriously affected with obstructive lung conditions prefer to use nebuliser therapy. Nebulisers are suitable for drugs with high dose and little patient co-ordination or skill; however, treatment using nebuliser is time consuming and less efficient, resulting in the waste of active medicaments. They are not portable devices and have been limited to the treatment of hospitalised patients. A number of nebulisers include AeroDose[®] (Aerogen), AeroEclipse[®] (Trudell Medical International), Halolite[®] (Medic-Aid Limited), RespiMat[®] (Boeinger Ingheim),

etc are currently available on market to deliver various types of drugs.

2.2. Pressurised metered dose inhalers

Pressurised metered dose inhalers (pMDIs), also known as metered dose inhalers (MDIs), are the most commonly used delivery devices. In this device (Figure 1), drug is either dissolved or suspended in liquefied propellants (or a mixture of propellants) with other excipients and presented in a pressurised canister fitted with a metering valve. On actuation of the valve, a predetermined amount of drug is released as spray. Aerosol formulations are packed in tin-plated steel, plastic coated glass or aluminium containers. The propellants used in pMDI formulations are liquefied gases of chlorofluorocarbons (CFC), which are not environmentally friendly. This is the reason why currently hydrofluoroalkanes (HFAs), which have no remarkable effects on the ozone layers, are used in the formulation for MDIs. At room temperature and pressure these are gases but they are liquefied by applying high pressure or by lowering temperature. On spraying, drug formulation with propellants are expelled and aerosolised. Although pMDIs are widely used in respiratory drug delivery, some problems have been associated with these devices, including the need for coordination of inspiratory inhalation with valve actuation and the use of a propellant, which has possible adverse effects on the stratospheric ozone layer as mentioned before. Currently there are a good number of pMDIs available on the market such as Ventolin (albuterol, GlaxoSmithKline), Azmacort (triamcnenolone acetate, Aventis Pharma), Symbicort

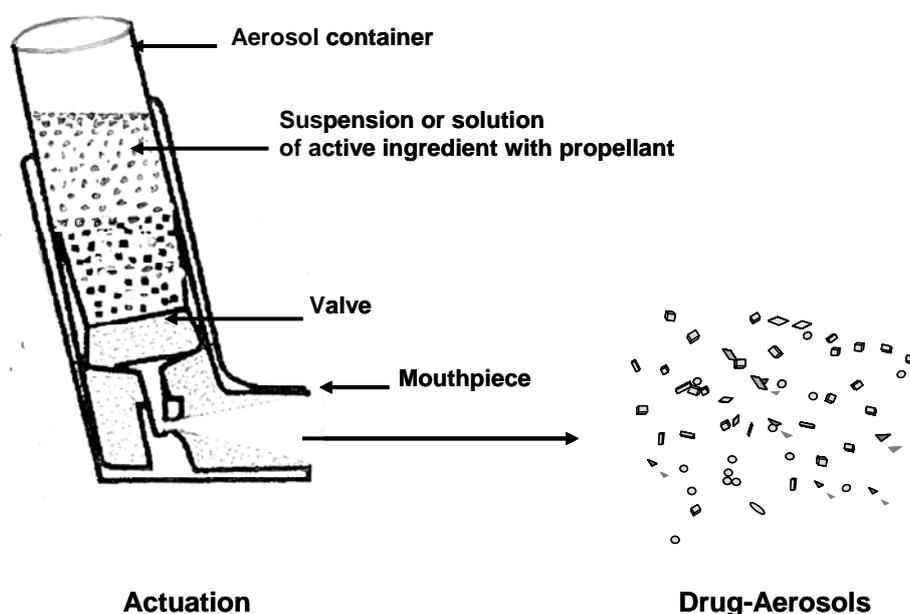


Figure 1. Schematic diagram of aerosol delivery of drugs from pMDI. Modified form Dalby *et al.* (2).

(Budesonide and formoterol, AstraZeneca), Flovent (Fluticasone, GlaxoSmithKline), *etc.* for the treatment of asthma.

There are some breath actuated and microprocessor controlled MDIs available on the market (Autohaler[®], Respimat[®]). These devices ensure the patient receives the drug at the correct point in the inspiration, and by slow inhalation with an indicator light to inform the patient whether the dose is inhaled or not. Anyway, as mentioned above, pMDIs have some disadvantages such as oropharyngeal deposition of drugs due to high velocity of propellants. The particles aerosolised from the MDIs have a high velocity, which exceeds the patients' inspiratory force, therefore, a large number of particles deposit onto the oropharyngeal areas. Thus a small fraction of drug deposits into the patients lungs (3) due to a lack of co-ordination between actuation and inhalation. To overcome this difficulty several inhalation aids like spacers incorporated with MDIs have been developed (4,5).

2.3. Dry powder inhaler (DPI) system

Dry powder inhalers contain the drug in a powder formulation, where drug particles (< 5 µm) are blended with a suitable large carrier (*e.g.* lactose) to improve flow properties and dose uniformity (6,7) and drug powders are delivered into the deep lung *via* a device known as dry powder inhaler (DPI). Powder de-agglomeration and aerosolisation from these formulations are achieved by the patient's inspiratory airflow, which needs to be sufficient to create an

aerosol containing respirable drug particles for lung deposition. Good flow properties of the formulation are necessary to ensure accurate dose metering of the drug. Advantages of DPI over other inhaler systems (pMDIs) are independence of breathing co-ordination with dose actuation, the absence of propellants, low innate initial velocity of particles (reducing inertial impaction at the back of the throat) and solid state drug stability. There are two types of DPI formulations; one is loose agglomerates of micronized drug particles having controlled flow properties, and the second one is carrier-based interactive mixtures (Figure 2) which consist of micronized (< 5 µm) drug particles mixed with larger carrier particles (8). Drug dispersion from the interactive mixtures can be enhanced by the addition of fine excipients (lactose) in the formulation (9,10). Drug particle size and powder formulation, breathing patterns and complex physiology of respiratory tract are major factors affect delivery of drugs into the deep lung. The redispersion of drug particles depends upon the interparticulate forces within the powder formulation. DPIs are highly portable, breath activated and relatively less expensive. Since drugs are kept in solid state in DPIs, they exhibit high physicochemical stability of drugs particularly proteins and peptides. In DPI formulation the device is an important factor in achieving adequate delivery of inhaled drug to lungs. The device must provide an environment where the drug can maintain its physicochemical stability and produce reproducible drug dosing.

There is a wide range of DPI devices, single, multi-unit or multiple dose devices, breath activated and

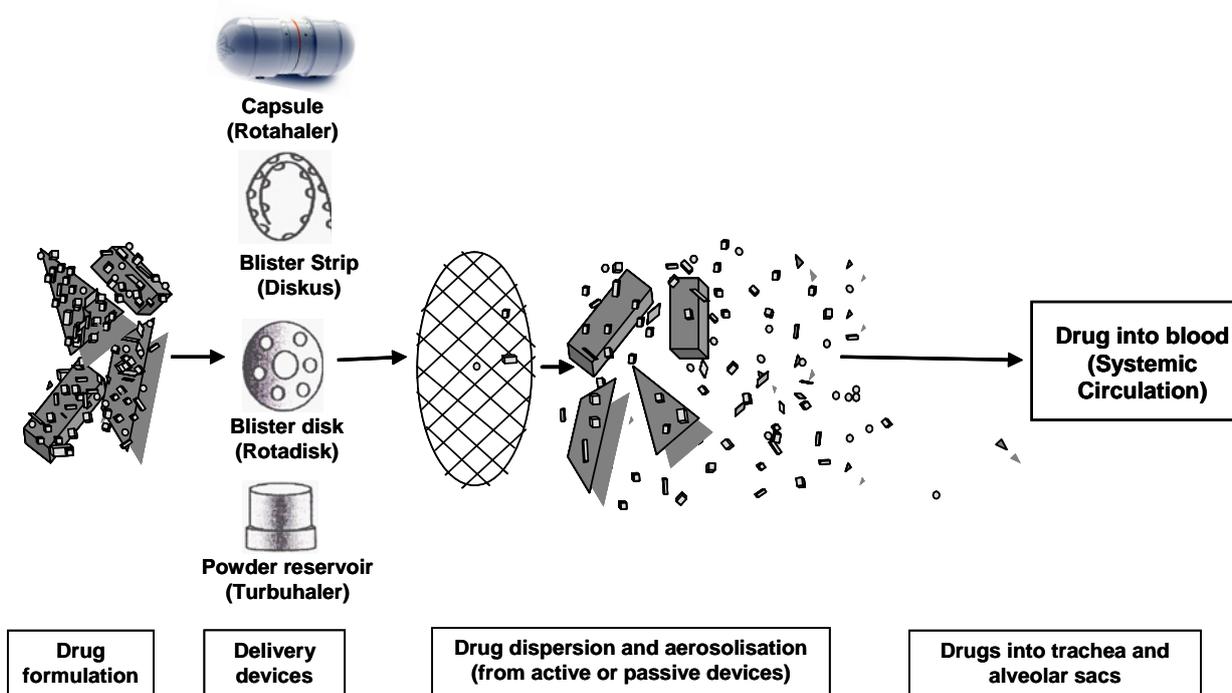


Figure 2. Schematic diagram of the pulmonary delivery of drugs from DPI formulations. The formulation consists of micronized drugs adhered on the surface of large carrier particles. Drug particles detached from the surface of large carriers and deposits into the patients airways by inhalation. Modified from Telko and Hickey (11).

power driven, available on the market; however, no devices showed efficiency in maximal drug delivery. Currently, based on the design, DPI devices may be classified into three broad categories *i.e.*, the first generation DPIs, the second generation DPIs and the third generation DPIs. The first generation DPIs were breath activated single unit dose (capsule) *i.e.*, the Spinhaler[®] and Rotahaler[®]. The second generation of DPIs use better technology *i.e.*, multi-dose DPIs (they measure the dose from a powder reservoir) or multi-unit dose (they disperse individual doses which are premeasured into blisters, disks, dimples, tubes and strip by the manufacturers) and multi-unit dose devices are likely to ensure the reproducibility of the formulation compared to that of multi-dose reservoir. The third generation DPIs, also known as active devices, which employ compressed gas or motor driven impellers or use electronic vibration (12,13) to disperse drug from the formulation. The very first approved active device (Exubera[®], Pfizer) with compressed air to aerosolise drug formulation for DPI insulin delivery was available on market; however, due to some unknown reasons, the production has been discontinued.

3. Mechanisms of drug deposition from aerosols

To achieve a desired therapeutic effect from aerosols, an adequate amount of drug must reach the alveolar sacs of the respiratory airways. The dynamic behavior of aerosol particles is governed by the laws of aerosol kinetics (14). The dominant mechanisms of depositing aerosol particles into the respiratory tract include inertial impaction, sedimentation (gravitational deposition), Brownian diffusion, interception and electrostatic precipitation (15). Inertial impaction and sedimentation are the most important for large particle deposition ($1\ \mu\text{m} < \text{MMAD} < 10\ \mu\text{m}$). A brief description of each mechanism of deposition is given below:

Inertial impaction: This is the main deposition mechanism at the tracheal bifurcation or successive branching points of airways. The airflow changes its direction at branching of the airways. The aerosol particles continue to move in their original direction and impact on any obstacle on the way. The deposition of aerosol particles by impaction increases with increasing air velocity, frequency of breathing and particle size (16). Large particles ($> 5\ \mu\text{m}$) with high velocity are mainly deposited by impaction (17).

Sedimentation: Sedimentation occurs when the gravitational force exerted on a particle overcomes the force of the air resistance. Particles of smaller size ($0.5\text{-}3.0\ \mu\text{m}$), which have tendency to escape from deposition by inertial impaction, may be deposited by sedimentation. Deposition of small particles by sedimentation mainly occurs in the smaller airways and alveolar regions and increased sedimentation

is observed during breath-holding or slow steady breathing (17).

Diffusion: Deposition of aerosolized particles less than $0.5\ \mu\text{m}$ occurs by diffusion due to Brownian movement. Deposition of aerosols by diffusion is independent of the density of particles but increases with decreasing size. Generally, the deposition of particles larger than $1.0\ \mu\text{m}$ is dominated by inertial impaction and particles smaller than $0.1\ \mu\text{m}$ are deposited by diffusion. Both sedimentation and diffusion are important for the particle size ranging between $0.1\text{-}1.0\ \mu\text{m}$ (18).

Interception: Although particle deposition by interception is not common, the deposition of elongated particles (particles large in one dimension but with small aerodynamic diameters) is believed to occur by this mechanism. Deposition of particles in the respiratory airways by interception is important when the dimensions of the anatomic spaces of airways become comparable to the dimensions of the particles (17).

Electrostatic precipitation: Electrostatic charges may be generated in a DPI on particles of an aerosol. Particles are inhaled immediately after charge generation and before neutralisation of the charge can occur. A charged particle may induce an image charge of opposite polarity on the airway walls. This image charge attracts the particle which is subsequently deposited by electrostatic precipitation (17,19). Only fibrous particles are believed to be deposited by this mechanism, therefore, this mechanism may not be significant for DPI formulations.

4. Pulmonary delivery of various drugs

4.1. Current drugs

Currently, local delivery of medicaments to the alveoli of lungs from both DPIs and pMDIs are mainly used for the treatment of lung disorders including asthma and bronchitis and a limited number of therapeutic compounds such as β -adrenoceptor agonist, muscarinic agonist, corticosteroids and mast cell stabilizers are available. Recently certain combinations of drugs are also formulated due to a synergistic therapeutic benefit. Corticosteroids and long acting β -adrenoceptor agonists formulations are available as both pMDIs and DPIs (20). Zanamavir, an antiviral agent has been introduced in the market as an aerosol product for the treatment of influenza (21). Aerosol delivery of recombinant human deoxyribonuclease (rhDNase) and tobramycin are available as nebuliser for the treatment of cystic fibrosis (21,22). The very first approved aerosol delivery of insulin as DPI formulation (Exubera[®], Pfizer) was introduced in the market; however, the production of this drug has been discontinued from market in early 2008 due to some unknown reasons.

Table 1. Drugs administered as aerosols against various diseases

Indication	Drug substances	Delivery method	References
Cystic fibrosis	Amiloride	Liquid	98,99
	Tobramycin	Powder	100,101
	DNAse	Powder	102,103
	Colistin sulphomethate	Powder	53
Cancer	Doxorubicin	Powder	104,105
Diabetes	Insulin	Microparticle/Liquid	106-108
Osteoporosis	Calcitonin	Microparticle	24,25
Sexual dysfunction	Apomorphine, Phosphodiesterase type 5 (PDEs) inhibitors	Microparticle	71,109
Vaccines	Malarial vaccine	DPI/ Microparticle	45
	Measles vaccine	Microparticle	44
	Influenza vaccine	Microparticle	46,47
	Zanamivir	Microparticle	110,111
Endometriosis, Pubertus praecox, Prostate carcinoma	Leuprolide	Powder/Liquid	26,27
Hormone replacement therapy	Testosterone	Liquid/Oral AREx	112
Immunosuppressor	Cyclosporin A	Microparticle/Liquid	113,114
Thrombosis and emphysema	Heparin	Microparticle	115,116

4.2. Drugs delivered as aerosol

Aerosol delivery offers the greatest potential to delivery drugs into the lower airway of the lungs of a wide range molecule for systemic diseases. A list of various drugs administered *via* pulmonary route has been presented in the Table 1.

Aerosol delivery of macromolecules is a potential non-invasive way of administering drug, to avoid frequent injections. Lung delivery of insulin has already been established; however, insulin loaded chitosan nanoparticles (23); nanoparticles of calcitonin (24); and nanospheres of elcatonin coated with chitosan (25), have been demonstrated for successful deep lung delivery. Aerosol delivery of leoprolide has been investigated as both MDI and DPI formulation for the management of prostate cancer (26,27). Dry powders of other proteins like parathyroid hormone for osteoporosis (28,29), glucagone (30), growth hormone (hGH) for dwarfism (31), vasoactive intestinal peptide (VIP) for pulmonary diseases like asthma (32) have been successfully investigated.

Lung delivery of genes that directly target the regions of interest by avoiding problems associated with intravenous delivery has been developed. Recently, successful gene delivery into lungs for cystic fibrosis has been demonstrated (12,33). Using nebuliser, delivery of liposomally encapsulated adenoviral vectors containing genes have been investigated; however, efficiency in low gene transfer at the cellular level has

been demonstrated (34,35). Lung delivery of genes complex with cationic lipids (lipoplex) and polymer-based (polyplex) are in progress (36,37) and a cationic lipid coupled with plasmid DNA (lipoplex), showed efficient lung delivery of gene (38). In another study, aerosol delivery of p53 and cytokine (IL-12) delivered *via* a nebuliser have been reported for therapeutic responses with reduced toxicity in animal lung tumor model (39-41). Based on the above mentioned researches it seems that there is a potential future of pulmonary gene therapy for various types of clinical applications.

Aerosol delivery of vaccines is an another area of interest and inhalation of measles vaccine was showed to be both safe and effective (42) and nebulised measles vaccine in human model found to produce better immunity with reduced side effects compared to that of subcutaneous injection (43). Dry powder inhaler formulation of measles vaccine (44), mucosal vaccination for influenza virus (45), malarial vaccine (46), and siRNA (47) have been investigated with significant success. Very recently, aerosol delivery of human immunodeficiency virus (HIV) treatments in infected patients found to be therapeutics with reduced toxicity and improved patient compliance (48). Therefore, it seems that pulmonary delivery of various genes is progressing and in future the world will see suitable vaccines against many pulmonary pathogens like *Mycobacterium tuberculosis*, respiratory syncytial virus (RSV), and severe acute respiratory syndrome (SARS).

Inhaled rifampicin antibiotic, and rifampicin loaded poly(lactide-co-glycolide) microparticles (49), colistin sulphate (50,51), and mucoactive agent Nacystelyn (52) have been found to be promising against cystic fibrosis (CF). Moreover, DPI formulation of colistin (53), gentamicin (54), azithromycin (55), tobramycin (56), have been effective method of treating CF. Furthermore, deep lung delivery of amphotericin B desoxycholate, liposomal amphotericin B, amphotericin B lipid complex and amphotericin B colloidal dispersion *via* nebulizers has been shown to be valuable in the prophylactic treatment of pulmonary aspergilosis (57). A nebulised dispersion of amorphous itraconazole nanoparticles (300 nm) produced by ultra-rapid freezing technique, showed improved bioavailability in mice (58). This outcome offers the application of itraconazole nanoparticles for the efficient treatment of fungal infections.

Lung delivery of aerosolised chemotherapeutic agents for the direct local treatment of lung tumors has been explored and found advantageous over other methods of drug delivery systems. Pulmonary delivery of aerosolised 9-nitrocamptothecin (9-NC) and cisplatin in patients with lung cancer have shown safety and promising antitumor effect (59,60). In addition, lung delivery of doxorubicin (61), paclitaxel (62), celecoxib and docetaxel (63,64), gemcitabine (65), liposomal camptothecin (66), *etc.*, has also been investigated and reduced toxicity was demonstrated. Recently, doxorubicin-loaded nanoparticles in dry powder aerosol form showed significant cytotoxicity in lung cancer (67). The researchers have indicated the potential of inhalation delivery of anticancer drugs in the treatment of lung cancer; however, further details investigation has been warranted.

Very recently, Dames and his co-workers developed targeted delivery of colloidal iron oxide nanoparticles (super magnetic iron oxide) (68), suspension of tocopherol nanoparticles coupled with biodegradable polymers for delayed release (69) and liposomal encapsulated cannabinoid for a prolonged psychoactive effect (70) have been demonstrated. In another study, aerosol delivery of apomorphine for sexual dysfunction (71), morphine and fentanyl for pain management (72), and ergotamine for migraine headaches (73,74). Furthermore, using nebuliser, aerosol delivery of radiopharmaceutical, ^{99m}Tc with phosphate buffer for lung ventilation imaging purposes is widely used (75). Inhalation of radiolabeled sulfur colloid (SC) aerosol (^{99m}Tc -SC, 0.2 μm) for studying particle uptake by airway surface macrophages has been demonstrated (76).

The aforementioned findings show the wider application of aerosol delivery of drug-aerosols; which offers the greatest potential to deliver drugs into the lower airway of lungs of a wide range of molecules (*i.e.*, antibiotics, genes, peptides and proteins, antibodies and

oligonuclides) for systemic diseases and put forwards the most promising inhalable platform for efficient systemic administration.

5. Inhaled drugs for neurodegenerative disorders

Pulmonary delivery of drugs for the management of neurological diseases is not currently approved. As deep lung delivery of different drugs showed potential benefit, researchers are now focusing to expand research on delivering drugs into the deep lungs *via* DPIs/MDIs. Some drugs administered as aerosols for the treatment of neurological disorders are shown in Table 2.

5.1. Pulmonary delivery of drugs for Parkinson's disease (PD)

PD is a chronic and progressive movement disorder (77). Millions of people suffer from PD in the developed and developing world (77). It is estimated that approximately 15 percent of people with PD are diagnosed before the age of 50 with incidence increasing with age. Evidence suggests that PD occurs when trouble sprouts in the basal ganglia, a segment of brain areas known for their contribution to movement. In general, nerve cells in the brain substantia nigra inexplicably die or become impaired. Normally these cells communicate *via* the chemical dopamine (DA) with cells in another one of the areas, the striatum, which includes subareas called the putamen and the caudate nucleus. Without DA, the striatum can't send out the electrical signals needed for normal movement, and consequently PD develops. Recently, a number of studies also indicate that the basal ganglia are involved with some of the cognitive problems that PD patients experience (78). Although there is presently no effective cure, there are many treatment options such as medication and surgery to manage the PD symptoms.

Levodopa is considered to be a temporary solution for minimising PD symptoms (79,80). Research suggests that the drug enters the brain and is transported into cells that can convert it into DA in the striatum. At first, symptoms diminish, but symptoms return in three to five years. Thus far, research suggests that the cells that convert levodopa die off. Higher doses of levodopa can make up for the decreasing number of cells but may cause jerking movements of the limbs, trunk and head as well as hallucinations. However, a number of attempts have been made to improve on the levodopa treatment by creating drugs that mimic DA by using DA agonists. Low doses of the DA agonists in combination with levodopa create less severe side effects and work for longer periods of time. Scientists are now taking this step further to determine if the drugs that mimic DA can replace levodopa altogether (81). At this point, researchers have developed drugs

Table 2. Drugs administered as aerosols against neurological disease

Indication	Drug substances	Delivery method	Details of delivery method	References
Pain, dyspnea	Opioids/Morphine	Powder	Micronized powders and aqueous formulation aerosolized	117-119
	Fentanyl/alfentanil and morphine	Liquid	Liposomal fentanyl delivered via a nebulizer and MDI	72,120
Smoking cessation	Nicotrol [®] Nicorette [®] (Nicotine)	Nicotine cartridge/ Liquid	Nicotine cartridges in an inhaler; pMDI	121,122
Migraine, Vascular cephalgia	Dihydroergotamine mesylate	Liquid	HFA 134a based suspension delivered by MDI	73
	Ergotamine tartrate	Liquid	Pulmonary delivery method is unavailable	123
	Detorelix	Liquid	Intratracheal (<i>i.t.</i>) instillation of Liposomal drugs	124
Parkinson's disease	Dopamine D-1 agonist, ABT-431	Liquid	HFA based drug formulation was delivered by MDI and AERx	85,86
Parkinson's disease	L-Dopa/ L-Dopa + agonist	Powder/liquid	Alkermes AIR, delivery of small or large molecules to the deep lung; Drugs in propellant delivered by MDI	83,84,89

Note: pMDI is pressurised metered dose inhaler; MDI: metered dose inhaler; HFC is hydrofluoro alkane; AERx is a DPI device that deliver aerosolised drugs from a dosage form that consists of liquid drug formulation and a nozzle array.

that each target a specific DA receptor site such as D1 or D2 (82). The drugs that are acting at D2 sites appear not to work effectively alone. But preliminary research on rodent models shows drugs that act on D1 sites work better than when administered without levodopa. Thus, these new drugs appear to work better than the use of levodopa alone because they continue to show benefits over time and cause lesser side effects. More importantly, several investigators have discovered other DA receptors sites such as, D3, D4, and D5. It is likely that in the future drugs could be developed that would act on these receptor sites (80-82).

Overall, levodopa (L-dopa) provides better therapeutic advantage for most early stage PD patients and current treatments of PD are primarily with oral formulation, such as levodopa/carbidopa, bromocriptine, selegiline, benzotropine and trihexyphenidyl; however, the efficacy of orally administered formulations becomes problematic with the progression of disease condition and due to a lost ability to control L-dopa's poor pharmacokinetics. Majority of the drugs approved and currently available on the market are oral (Tablet/capsules), some are parenteral (IV, IM, SC) and only one is a transdermal patch; however, no pulmonary delivery products been not developed.

Deep lung delivery of levodopa particles for treating a patient with PD has been reported (83); however, no further data is available for the readers.

Using MDI, delivery of a DA agonist to the airways has been demonstrated and the authors indicated that the inhalation route provided effective delivery of the drug to the receptor (84). Pulmonary administration of a drug (ABT-431, a selective D1 receptor agonist) was found to be significantly greater than that of oral administration (85). For example with intratracheal instillation of the drug solution, bioavailability of the drug was 75% in dogs and tetrafluoroethane (HFC-134a) based MDI formulation showed 40% bioavailability. The lung bioavailability of the aerosolised drug was 34% compared to intravenous injection in the same dogs. The authors emphasised that a single rising dose in human study demonstrated that the absorption of ABT-431 following oral inhalation administration (bioavailability 25%) resulted in a dose-dependent increase in the AUC (area under the curve) versus time profile at dosages from 3.3 mg to 13.2 mg. sing a novel delivery device (AERx), Okumu and his associates (86) delivered ABT-431 (a selective dopamine D-1 receptor agonist) to healthy male volunteers and plasma samples were analysed following lung delivery and intravenous administration and 82-107% of pulmonary bioavailability was observed. This outcome demonstrated that the aerosol inhalation of this drug was a proficient means for systemic delivery.

Dugs for pulmonary delivery for the management of PD have not currently been approved; however,

only one formulation containing L-dopa has been investigated (83,87). The inhalation data for pulmonary delivery of L-dopa showed at least two fold less dose compared to that of oral dose (88). In another study, demonstrated aerosol delivery of L-dopa dry powder formulation in a rat model and pulmonary administered L-dopa showed rapid and higher plasma levels ($C_{max} = 4.8 \pm 1.10$ mg/mL at 2 min) compared to that of oral administration where the drug produced delayed and lower plasma level ($C_{max} 1.8 \pm 0.40$ mg/mL) at 30 min (89). The authors acknowledged that an inhalable formulation of L-dopa may provide PD patients with effective form of rescue therapy as well as replacement for first-line oral therapy. However, this formulation is in preclinical stage. Innovative drug delivery in treating PD has the potential to reduce many adverse effects of currently available drugs. Pulmonary delivery of current drugs, genes and liposomes will help encourage patient specific treatment for PD. Improved treatments with pulmonary delivery system may involve drugs that target one specific site or a combination of sites for better pharmacotherapeutics. Therefore, these advances, along with investigations into gene transfer, surgical and transplantation techniques, hold great promise for those with PD.

5.2. Pulmonary delivery of drugs and Alzheimer's disease (AD)

Dementia is a brain disease that significantly affects a person's ability to carry out daily routine activities. It is well established that the most common form of dementia among older people is AD, which initially involves the various parts of the brain that control thought, memory, and language (90,91). In recent years some progress has been made, however, the causes of AD remain unknown, and there is no effective cure. Like PD (see above), it is estimated that millions of people worldwide suffer from AD (91). Previous research has found that other brain changes in people with AD. For example, brain neurons die in areas of the brain that are vital to memory and other mental abilities, and connections between neurons are disrupted. Furthermore, AD may impair thinking and memory by disrupting neuronal transmissions and functions. AD is a slow disease, starting with mild memory problems and ending with severe brain damage. The course the disease takes and how fast changes occur vary from person to person. On average, AD patients live from 8 to 10 years after they are diagnosed, though some people may live with AD for as many as 20 years (90,91).

Given the complex disease process with AD, there is no better therapeutic strategy to reduce or minimize AD. However, some drugs such as, tacrine, donepezil, rivastigmine, or galantamine may help prevent some symptoms from becoming worse for a limited time in some people in the early and middle stages of the

disorder (92,93). Furthermore, memantine has been marketed to treat moderate to severe AD, although it also is limited in its therapeutic benefits. There are some formulations that may help control behavioral symptoms of AD such as insomnia, agitation, anxiety, and depression. Despite the complexity of the symptom pattern in AD, treating these symptoms often makes patients more comfortable and makes their care easier for caregivers. In addition, many researchers have begun to search for ways to block the formation of amyloid deposits, an important biomarker for AD. In a recent study, investigators used the amyloid protein as a vaccine to prevent and clear existing plaques in mice that were engineered to develop large numbers of the deposits (93). Although it remains unclear, the vaccine appears to involve the immune system and clearing amyloid. Future human studies will test the level of the plaques' contribution to AD. More importantly, the research has the potential to show that the vaccine method can influence the clinical signs in patients. A number of research groups are investigating slight variations of the vaccine strategy (94). For example, one group found positive results with an amyloid vaccine that was delivered to mice in the form of nasal drops. It is suggested that other therapeutic strategy may help prevent plaques. Several investigators have uncovered some evidence that the estrogen hormone may influence the development of AD. A number of human studies demonstrated that older women who take estrogen supplements can reduce their risk of developing AD. It is possible that estrogen's benefits, at least in part, may result from an ability to reduce the levels of amyloid protein. In a recent study, estrogen reduced the levels of amyloid protein made in a cell culture model. Recently, a long-sought enzyme, beta-secretase which makes one of the cuts that leads to the formation of amyloid protein has been identified (95). The researchers plan to develop drugs that can inhibit the enzyme. Additionally, an enzyme involved in the formation of amyloid is gamma-secretase, which has not yet been purified. Overall, these significant discoveries shed light on AD treatment strategy and provide hope that amyloid plaque-directed therapies for humans may soon become available.

However, no specific medications have been formulated yet to deliver drugs into deep lung for the treatment of AD. Using nebuliser, pulmonary delivery of apolipoprotein, amphipathic compounds and apolipoprotein were found to efficiently reach the systemic circulation through the lung (96); however, no further data are available for the readers. The author claimed that pulmonary delivery of drugs can be used for the treatment of AD as well as cardiovascular disease. Most drugs for the treatment of AD are oral (very slow absorption) or parenteral (expensive) and aerosol formulations have not been studied. Thus, it would be worthwhile for the researchers to focus on

pulmonary delivery of drugs for the management of AD. Like PD, current treatments of AD are primarily with oral formulation as described above. Improved treatments with pulmonary delivery system may involve drugs that target one specific site or a combination of sites for better pharmacotherapeutics.

6. Future directions

Lung delivery of drugs offer the greatest potential to deliver drugs into the lower airway of the lungs and the delivery of a powder form of a wide range of molecules (*i.e.*, antibiotics, peptides, proteins, antibodies and oligonucleotides) to the deep lung for systemic diseases put forwards the most promising inhalable stage for increasing systemic administration. Pulmonary delivery of large molecules for chronic diseases is advancing tremendously and may become successful in the near future. Therefore, pulmonary drug delivery needs to focus not only on lung diseases but also on conditions in which fast onset is desirable such as cancer pain, allergic reactions, brain disorders, cardiovascular disorders, and sexual dysfunction. It has been reported that more than 40 drug formulations for widely varying designs of DPIs are in the pipeline for drug delivery into the deep lung; however, only four inhaled products (for the treatment of asthma only) include a long acting bronchodilator, two corticosteroids and a combination formulation (97). In addition, anticancer drugs, steroids, beta2 receptor agonists, antimuscarinics, antihistamines and anti-inflammatory agents, which are primarily administered by oral or parenteral route, may be considered for pulmonary delivery. Therefore, there is a promising future for lung delivery of drugs for the management of other systemic disorders along with pulmonary diseases. Local and systemic delivery of different drugs for chronic systemic diseases needs to be more focused on the use of aerosol formulations, which have a lot of potential. In future, biotechnology products will produce very small amount of potent drugs which will require smart devices that deliver drugs efficiently into the lower airway of lungs. The current trend in pulmonary drug delivery and potential benefits of this route will enable the continued development of smart but reliable DPI technology to enhance deposition of drugs into deep lungs with a better patient compliance.

7. Conclusions

Pulmonary delivery of various drugs by aerosolisation has been used for centuries to treat respiratory tract diseases and the pressurised metered-dose inhaler was the only delivery device of choice. Currently, aerosol therapy is expanding with the advancement of science and technology specially in developing dry powder inhaler formulations to target the systemic circulation

for the delivery of proteins and peptides, gene therapy, and influenza and measles vaccines. Moreover, advances in all of these areas have led to pulmonary delivery of medicaments being a route of choice for many drugs, not only for respiratory diseases but also for systemic delivery of drugs for other disorders. Now a days, mental health disorders and cardiac diseases are increasing with the changing world; however, pulmonary delivery of neuroactive and cardio active drugs has not been explored. Most drugs for treating AD or PD are in oral dosages forms (tablets, capsules and solutions); forms where drug absorption is very slow and only 70-90% is bioavailable. The longer range future of DPIs does include non-invasive and efficient delivery of large molecules for systemic conditions with improved patient compliance. Therefore, pulmonary drug delivery of drugs would extend the new era of drug delivery research, which will eventually extend the life of drugs (solid state stability), increase patient compliance, and reduce the total cost not only for brain neurodegenerative disorders but also for other chronic human diseases.

Acknowledgement

We gratefully acknowledge the contribution made by Gail Neilson in proofreading this review article.

References

1. Byron PR, Patton JS. Drug delivery *via* the respiratory tract. *J Aerosol Med* 1994; 7:49-75.
2. Dalby RN, Tiano SL, Hickey AJ. Medical devices for the delivery of therapeutic aerosols to the lungs. In: *Lung Biology in Health and Disease*. Vol. 94. Inhalation Aerosols. 2nd Edition (Hickey AJ, ed.). Marcel Dekker, Inc., New York, USA, 2007; pp. 417-444.
3. Newman SP, Pavia D, Clarke SW. How should a pressurized beta-adrenergic bronchodilator be inhaled? *Eur J Respir Dis* 1981; 62:3-21.
4. Ikeda A, Nishimura K, Koyama H, Tsukino M, Hajiro T, Mishima M, Izumi T. Comparison of the bronchodilator effects of salbutamol delivered *via* a metered-dose inhaler with spacer, a dry-powder inhaler, and a jet nebulizer in patients with chronic obstructive pulmonary disease. *Respiration* 1999; 66:119-123.
5. Bisgaard H. Automatic actuation of a dry powder inhaler into a nonelectrostatic spacer. *Am J Respir Crit Care Med* 1998; 157:518-521.
6. French DL, Edwards DA, Niven RW. The influence of formulation on emission, deaggregation and deposition of dry powders for inhalation. *J Aerosol Sci* 1996; 27:769-783.
7. Timsina MP, Martin GP, Marriott C, Ganderton D, Yianneskis M. Drug delivery to the respiratory tract using dry powder inhalers. *Int J Pharm* 1994; 101:1-13.
8. Hersey JA. Ordered mixing: a new concept in powder mixing practice. *Powder Technol* 1975; 11:41-44.
9. Islam N, Stewart P, Larson I, Hartley P. Lactose surface modification by decantation: are drug-fine lactose Ratios the key to better dispersion of salmeterol xinafoate

- from lactose-interactive mixtures? *Pharm Res* 2004; 21:492-499.
10. Islam N, Stewart P, Larson I, Hartley P. Effect of carrier size on the dispersion of salmeterol xinafoate from interactive mixtures. *J Pharm Sci* 2004; 93:1030-1038.
 11. Telko MJ, Hickey AJ. Dry powder inhaler formulation. *Respir Care* 2005; 50:1209-1227.
 12. Brown BAS, Rasmussen JA, Becker DP, Friend DR. A piezo-electronic inhaler for local & systemic applications. *Drug Deliv Technol* 2004; 4:90-93.
 13. Young PM, Thompson J, Woodcock D, Aydin M, Price R. The development of a novel high-dose pressurized aerosol dry-powder device (PADD) for the delivery of pumactant for inhalation therapy. *J Aerosol Med* 2004; 17:123-128.
 14. Newman SP, Clarke SW. Therapeutic aerosols 1--physical and practical considerations. *Thorax* 1983; 38:881-886.
 15. Gonda I. Aerosols for delivery of therapeutic and diagnostic agents to the respiratory tract. *Crit Rev Ther Drug Carrier Syst* 1990; 6:273-313.
 16. Carpenter RL. Aerosol deposition modeling using ACSL. *Drug Chem Toxicol* 1999; 22:73-90.
 17. Gonda I. Targeting by deposition. In: *Pharmaceutical Inhalation Aerosol Therapy* (Hickey AJ, ed.). Marcel Dekker, Inc., New York, USA, 1992; pp. 61-82.
 18. Brain JD, Blancard JD. Mechanisms of particle deposition and clearance. In: *Aerosols in Medicine, Principles, Diagnosis and Therapy*. 2nd Edition (Moren F, Dolovich MB, Newhouse MT, Newman SP, eds.), Elsevier Science Publishers, Amsterdam, Netherland, 1994; pp. 117-155.
 19. Chan TL, Yu CP. Charge effects on particle deposition in the human tracheobronchial tree. *Ann Occup Hyg* 1982; 26:65-75.
 20. Dhillon S, Keating GM. Beclometasone dipropionate/formoterol: in an HFA-propelled pressurised metered-dose inhaler. *Drugs* 2006; 66:1475-1483.
 21. Cass LMR, Brown J, Pickford M, Fayinka S, Newman SP, Johansson CJ, Bye A. Pharmacoscintigraphic evaluation of lung deposition of inhaled zanamivir in healthy volunteers. *Clin Pharmacokinet* 1999; 36 (Suppl. 1):21-31.
 22. Kuhn RJ. Pharmaceutical considerations in aerosol drug delivery. *Pharmacotherapy* 2002; 22 (3 Pt 2):80S-85S.
 23. Grenha A, Seijo B, Remuñán-López C. Microencapsulated chitosan nanoparticles for lung protein delivery. *Eur J Pharm Sci* 2005; 25:427-437.
 24. Kawashima Y, Takeuchi H, Yamamoto H, Mimura K. Powdered polymeric nanoparticulate system to improve pulmonary delivery of calcitonin with dry powder inhalation. *Proceedings of the International Symposium on Controlled Release of Bioactive Materials* 2000; 27:229-230.
 25. Yamamoto H, Kuno Y, Sugimoto S, Takeuchi H, Kawashima Y. Surface-modified PLGA nanosphere with chitosan improved pulmonary delivery of calcitonin by mucoadhesion and opening of the intercellular tight junctions. *J Control Release* 2005; 102:373-381.
 26. Anon. Method of and apparatus for effecting delivery of fine powders. *IP. Com J* 2008; 8 (1B):13.
 27. Shahiwala A, Misra A. A preliminary pharmacokinetic study of liposomal leuprolide dry powder inhaler: a technical note. *AAPS PharmSciTech* 2005; 6:E482-E486.
 28. Codrons V, Vanderbist F, Ucakar B, Pr at V, Vanbever R. Impact of formulation and methods of pulmonary delivery on absorption of parathyroid hormone (1-34) from rat lungs. *J Pharm Sci* 2004; 93:1241-1252.
 29. Codrons V, Vanderbist F, Verbeeck RK, Arras M, Lison D, Pr at V, Vanbever R. Systemic delivery of parathyroid hormone (1-34) using inhalation dry powders in rats. *J Pharm Sci* 2003; 92:938-950.
 30. Endo K, Amikawa S, Matsumoto A, Sahashi N, Onoue S. Erythritol-based dry powder of glucagon for pulmonary administration. *Int J Pharm* 2005; 290:63-71.
 31. Bosquillon C, Pr at V, Vanbever R. Pulmonary delivery of growth hormone using dry powders and visualization of its local fate in rats. *J Control Release* 2004; 96:233-244.
 32. Ohmori Y, Onoue S, Endo K, Matsumoto A, Uchida S, Yamada S. Development of dry powder inhalation system of novel vasoactive intestinal peptide (VIP) analogue for pulmonary administration. *Life Sci* 2006; 79:138-143.
 33. Lentz YK, Anchordoquy TJ, Lengsfeld CS. Rationale for the selection of an aerosol delivery system for gene delivery. *J Aerosol Med* 2006; 19:372-384.
 34. Laube BL. The expanding role of aerosols in systemic drug delivery, gene therapy, and vaccination. *Respir Care* 2005; 50:1161-1176.
 35. Li HY, Seville PC, Williamson IJ, Birchall JC. The use of amino acids to enhance the aerosolisation of spray-dried powders for pulmonary gene therapy. *J Gene Med* 2005; 7:343-353.
 36. Wightman L, Kircheis R, Rossler V, Carotta S, Ruzicka R, Kurska M, Wagner E. Different behavior of branched and linear polyethylenimine for gene delivery *in vitro* and *in vivo*. *J Gene Med* 2001; 3:362-372.
 37. Kichler A, Leborgne C, Coeytaux E, Danos O. Polyethylenimine-mediated gene delivery: a mechanistic study. *J Gene Med* 2001; 3:135-144.
 38. Deshpande D, Blanchard J, Srinivasan S, Fairbanks D, Fujimoto J, Sawa T, Wiener-Kronish J, Schreier H, Gonda I. Aerosolization of lipoplexes using AERx pulmonary delivery system. *AAPS Pharm Sci* 2002; 4:E3.
 39. Gautam A, Densmore CL, Waldrep JC. Inhibition of experimental lung metastasis by aerosol delivery of PEI-p53 complexes. *Mol Ther* 2000; 2:318-323.
 40. Lungwitz U, Breunig M, Blunk T, G pferich A. Polyethylenimine-based non-viral gene delivery systems. *Eur J Pharm Biopharm* 2005; 60:247-266.
 41. Densmore CL. Advances in noninvasive pulmonary gene therapy. *Curr Drug Deliv* 2006; 3:55-63.
 42. Wong-Chew RM, Islas-Romero R, Garc a-Garc a Mde L, Beeler JA, Audet S, Santos-Preciado JI, Gans H, Lew-Yasukawa L, Maldonado YA, Arvin AM, Valdespino-G mez JL. Immunogenicity of aerosol measles vaccine given as the primary measles immunization to nine-month-old Mexican children. *Vaccine* 2006; 24:683-690.
 43. Dilraj A, Cutts FT, de Castro JF, Wheeler JG, Brown D, Roth C, Coovadia HM, Bennett JV. Response to different measles vaccine strains given by aerosol and subcutaneous routes to schoolchildren: a randomised trial. *Lancet* 2000; 355:798-803.
 44. LiCalsi C, Maniaci MJ, Christensen T, Phillips E, Ward GH, Witham C. A powder formulation of measles vaccine for aerosol delivery. *Vaccine* 2001; 19:2629-2636.
 45. Edwards DA, Sung J, Pulliam B, Wehrenberg-Klee E, Schwartz E, Dreyfuss P, Kulkarni S, Lieberman E. inventors; (President and Fellows of Harvard College,

- USA). assignee. Pulmonary delivery of malarial vaccine in the form of particulates. Application: WO patent 2005-US16082 2005110379. 2005 20050509.
46. Smith DJ, Bot S, Dellamary L, Bot A. Evaluation of novel aerosol formulations designed for mucosal vaccination against influenza virus. *Vaccine* 2003; 21:2805-2812.
 47. Brito L, Chen D, Ge Q, Treco D, inventors; (Nastech Pharmaceutical Company Inc., USA). assignee. Dry powder compositions for RNA influenza therapeutics. Application: US patent 2007-623306 2007172430. 2007 20070115.
 48. Cipolla D, inventor (Zogenix, Inc., USA). assignee. Needle-free delivery of HIV therapeutics through aerosols and inhalants. Application: WO patent 2007-US9745 2007124104. 2007 20070420.
 49. Sethuraman VV, Hickey AJ. Powder properties and their influence on dry powder inhaler delivery of an antitubercular drug. *AAPS PharmSciTech* 2002; 3:1-10.
 50. Le Brun PP, de Boer AH, Mannes GP, de Fraiture DM, Brimicombe RW, Touw DJ, Vinks AA, Frijlink HW, Heijerman HG. Dry powder inhalation of antibiotics in cystic fibrosis therapy: part 2 Inhalation of a novel colistin dry powder formulation: a feasibility study in healthy volunteers and patients. *Eur J Pharm Biopharm* 2002; 54:25-32.
 51. Westerman EM, Le Brun PPH, Touw DJ, Frijlink HW, Heijerman HGM. Effect of nebulized colistin sulphate and colistin sulphomethate on lung function in patients with cystic fibrosis: a pilot study. *J Cyst Fibros* 2004; 3:23-28.
 52. Vanderbist F, Wery B, Baran D, Van Gansbeke B, Schoutens A, Moes AJ. Deposition of Nacystelyn from a dry powder inhaler in healthy volunteers and cystic fibrosis patients. *Drug Dev Ind Pharm* 2001; 27:205-212.
 53. Westerman EM, de Boer AH, Le Brun PPH, Touw DJ, Frijlink HW, Heijerman HGM. Dry powder inhalation of colistin sulphomethate in healthy volunteers: A pilot study. *Int J Pharm* 2007; 335:41-45.
 54. Crowther Labiris NR, Holbrook AM, Chrystyn H, Macleod SM, Newhouse MT. Dry powder versus intravenous and nebulized gentamicin in cystic fibrosis and bronchiectasis. A pilot study. *Am J Respir Crit Care Med* 1999; 160:1711-1716.
 55. Hickey Anthony J, Lu D, Ashley Elizabeth D, Stout J. Inhaled azithromycin therapy. *J Aerosol Med* 2006; 19:54-60.
 56. Geller David E, Konstan Michael W, Smith J, Noonberg Sarah B, Conrad C. Novel tobramycin inhalation powder in cystic fibrosis subjects: pharmacokinetics and safety. *Pharmacol Res* 2007; 42:307-313.
 57. Ruijgrok Elisabeth J, Fens Marcel HA, Bakker-Woudenberg Irma AJM, van Etten Els WM, Vulto Arnold G. Nebulization of four commercially available amphotericin B formulations in persistently granulocytopenic rats with invasive pulmonary aspergillosis: evidence for long-term biological activity. *J Pharm Pharmacol* 2005; 57:1289-1295.
 58. Yang W, Tam J, Miller DA, Zhou J, McConville JT, Johnston KP, Williams RO 3rd. High bioavailability from nebulized itraconazole nanoparticle dispersions with biocompatible stabilizers. *Int J Pharm* 2008; 361:177-188.
 59. Gagnadoux F, Hureauux J, Vecellio L, Urban T, Le Pape A, Valo I, Montharu J, Leblond V, Boisdron-Celle M, Lerondel S, Majoral C, Diot P, Racineux JL, Lemarie E. Aerosolized Chemotherapy. *J Aerosol Med Pulm Drug Deliv* 2008; 21:61-70.
 60. Smyth HDC, Saleem I, Donovan M, Verschraegen CF. Pulmonary delivery of anti-cancer agents. In: *Drugs and the Pharmaceutical Sciences*. Vol. 172. *Advanced Drug Formulation Design to Optimize Therapeutic Outcomes* (Williamms III RO, Taft DR, McConville JT, eds.). Informa healthcare, New York, USA, 2008; pp. 81-111.
 61. Johnston MR, Minchen RF, Dawson CA. Lung perfusion with chemotherapy in patients with unresectable metastatic sarcoma to the lung or diffuse bronchioloalveolar carcinoma. *Thorac Cardiovasc Surg* 1995; 110:368-373.
 62. Hershey AE, Kurzman ID, Forrest LJ, Bohling CA, Stonerook M, Placke ME, Imondi AR, Vail DM. Inhalation chemotherapy for macroscopic primary or metastatic lung tumors: proof of principle using dogs with spontaneously occurring tumors as a model. *Clin Cancer Res* 1999; 5:2653-2659.
 63. Haynes A, Shaik MS, Chatterjee A, Singh M. Formulation and evaluation of aerosolized celecoxib for the treatment of lung cancer. *Pharm Res* 2005; 22:427-439.
 64. Fulzele Suniket V, Shaik Madhu S, Chatterjee A, Singh M. Anti-cancer effect of celecoxib and aerosolized docetaxel against human non-small cell lung cancer cell line, A549. *J Pharm Pharmacol* 2006; 58:327-336.
 65. Koshkina NV, Kleinerman ES. Aerosol gemcitabine inhibits the growth of primary osteosarcoma and osteosarcoma lung metastases. *Int J Cancer* 2005; 116:458-463.
 66. Knight JV, Koshkina N, Gilbert B, Verschraegen CF, inventors; (USA). assignee. Small particle liposome aerosols for delivery of anticancer drugs. Application: US patent 2003-663573 2006204447. 2006 20030916.
 67. Azarmi S, Tao X, Chen H, Wang Z, Finlay WH, Löbenberg R, Roa WH. Formulation and cytotoxicity of doxorubicin nanoparticles carried by dry powder aerosol particles. *Int J Pharm* 2006; 319:155-161.
 68. Dames P, Gleich B, Flemmer A, Hajek K, Seidl N, Wiekhorst F, Eberbeck D, Bittmann I, Bergemann C, Weyh T, Trahms L, Rosenecker J, Rudolph C. Targeted delivery of magnetic aerosol droplets to the lung. *Nat Nanotechnol* 2007; 2:495-499.
 69. Bonnet-Gonnet C, inventor (Flamel Technologies, Inc., Fr.). assignee. Pharmaceutical formulations comprising colloidal suspensions, for the prolonged release of active principle(s), and their applications, especially therapeutic applications. Application: US patent 2007-808456 2008026070. 2008 20070611.
 70. Hung O, Zamecnik J, Shek PN, Tikuisis P, inventors; (Her Majesty the Queen as Represented by the Minister of National Defence of, Can.). assignee. Pulmonary delivery of liposome-encapsulated cannabinoids. Application: WO patent 2000-CA805 2001003668. 2001 20000707.
 71. Staniforth JN, Morton D, Toby M, Eason S, Harmer Q, Ganderton D, inventors; (UK). assignee. Pharmaceutical compositions comprising apomorphine for pulmonary inhalation. Application: US patent 2006-552231 2006178394. 2006 20060421.
 72. Mather LE, Woodhouse A, Ward ME, Farr SJ, Rubsamen RA, Eltherington LG. Pulmonary administration of aerosolized fentanyl: pharmacokinetic analysis of

- systemic delivery. *Br J Clin Pharmacol* 1998; 46:37-43.
73. Pavkov RM, Armer TA, Mohsen NM, inventors; (Map Pharmaceuticals, Inc., USA). assignee. Aerosol formulations for delivery of dihydroergotamine to the systemic circulation via pulmonary inhalation. Application: WO patent 2004-US29632 2005025506. 2005 20040910.
 74. Armer TA, Shrewsbury SB, Newman SP, Pitcairn G, Ramadan N. Aerosol delivery of ergotamine tartrate *via* a breath-synchronized plume-control inhaler in humans. *Curr Med Res Opin* 2007; 23:3177-3187.
 75. Ballinger JR, Andrey TW, Boxen I, Zhang ZM. Formulation of technetium-99m-aerosol colloid with improved delivery efficiency for lung ventilation imaging. *J Nucl Med* 1993; 34:268-271.
 76. Alexis Neil E, Lay John C, Zeman Kirby L, Geiser M, Kapp N, Bennett William D. *In vivo* particle uptake by airway macrophages in healthy volunteers. *Am J Respir Cell Mol Biol* 2006; 34:305-313.
 77. Olanow CW, Tatton WG. Etiology and pathogenesis of Parkinson's disease. *Annu Rev Neurosci* 1999; 22:123-144.
 78. Weintraub D, Comella Cynthia L, Horn S. Parkinson's disease--Part 1: Pathophysiology, symptoms, burden, diagnosis, and assessment. *Am J Manag Care* 2008; 14: S40-48.
 79. Fahn S. Does levodopa slow or hasten the rate of progression of Parkinson's disease? *J Neurol* 2005; 252 (Suppl 4):IV37-IV42.
 80. Jankovic J, Stacy M. Medical management of levodopa-associated motor complications in patients with Parkinson's disease. *CNS Drugs* 2007; 21:677-692.
 81. Stoessl AJ. Potential therapeutic targets for Parkinson's disease. *Expert Opin Ther Targets* 2008; 12:425-436.
 82. Wu Stacy S, Frucht Steven J. Treatment of Parkinson's disease: what's on the horizon? *CNS Drugs* 2005; 19:723-743.
 83. Jackson B, Bennett DJ, Bartus RT, Emerich DF, inventors; (Advanced Inhalation Research, Inc., USA). assignee. Pulmonary delivery for levodopa. Application: WO patent 2003-US8659 2003079992. 2003 20030319.
 84. Adjei AL, Zheng J, Gupta PK, Marsh KC, Wu V, Lee DY, inventors; (Abbott Laboratories, USA). assignee. Formulations for pulmonary delivery of dopamine agonists. Application: US patent 97-822631 6193954. 2001 19970321.
 85. Zheng Y, Marsh KC, Bertz RJ, El-Shourbagy T, Adjei AL. Pulmonary delivery of a dopamine D-1 agonist, ABT-431, in dogs and humans. *Int J Pharm* 1999; 191:131-140.
 86. Okumu FW, Lee RY, Blanchard JD, Queirolo A, Woods CM, Lloyd PM, Okikawa J, Gonda I, Farr SJ, Rubsamen R, Adjei AL, Bertz RJ. Evaluation of the AERx pulmonary delivery system for systemic delivery of a poorly soluble selective D-1 agonist, ABT-431. *Pharm Res* 2002; 19:1009-1012.
 87. Bartus RT, Emerich DF, inventors; (Advanced Inhalation Research, Inc., USA). assignee. Pulmonary delivery in treating disorders of the central nervous system. Application: US patent 2001-877734 2002058009. 2002 20010608.
 88. Bartus RT, Emerich DR, inventors; (Advanced Inhalation Research, Inc., USA). assignee. Pulmonary delivery in treating disorders of the central nervous system. Application: WO patent 2001-US29311 2002024158. 2002 20010919.
 89. Bartus RT, Emerich D, Snodgrass-Belt P, Fu K, Salzberg-Brenhouse H, Lafreniere D, Novak L, Lo ES, Cooper T, Basile AS. A pulmonary formulation of L-dopa enhances its effectiveness in a rat model of Parkinson's disease. *J Pharmacol Exp Ther* 2004; 310:828-835.
 90. Sloane PD, Zimmerman S, Suchindran C, Reed P, Wang L, Boustani M, Sudha S. The public health impact of Alzheimer's disease, 2000-2050: potential implication of treatment advances. *Annu Rev Public Health* 2002; 23:213-231.
 91. Harciarek M, Kertesz A. The prevalence of misidentification syndromes in neurodegenerative diseases. *Alzheimer Dis Assoc Disord* 2008; 22:163-169.
 92. Lleo A. Current therapeutic options for Alzheimer's disease. *Curr Genomics* 2007; 8:550-558.
 93. Moreira PI, Zhu X, Nunomura A, Smith MA, Perry G. Therapeutic options in Alzheimer's disease. *Expert Rev Neurother* 2006; 6:897-910.
 94. Farlow MR, Miller ML, Pejovic V. Treatment options in Alzheimer's disease: Maximizing benefit, managing expectations. *Dement Geriatr Cogn Disord* 2008; 25:408-422.
 95. Ghosh AK, Gemma S, Tang J. beta-Secretase as a therapeutic target for Alzheimer's disease. *Neurotherapeutics* 2008; 5:399-408.
 96. Wagle N, inventor (USA). assignee. Non-invasive treatment of diseases using amphipathic compounds. Application: US patent 2005-57091 2005176623. 2005 20050211.
 97. Byron PR. Drug delivery devices: issues in drug development. *Proc Am Thorac Soc* 2004; 1:321-328.
 98. Chougule MB, Padhi BK, Misra A. Nano-liposomal dry powder inhaler of Amiloride Hydrochloride. *J Nanosci Nanotechnol* 2006; 6:3001-3009.
 99. Thomas SH, O'Doherty MJ, Graham A, Page CJ, Blower P, Geddes DM, Nunan TO. Pulmonary deposition of nebulised amiloride in cystic fibrosis: comparison of two nebulisers. *Thorax* 1991; 46:717-721.
 100. Newhouse MT, Hirst PH, Duddu SP, Walter YH, Tarara TE, Clark AR, Weers JG. Inhalation of a dry powder tobramycin PulmoSphere formulation in healthy volunteers. *Chest* 2003; 124:360-366.
 101. Le Conte P, Potel G, Peltier P, Horeau D, Caillon J, Juvin ME, Kerguéris MF, Bugnon D, Baron D. Lung distribution and pharmacokinetics of aerosolized tobramycin. *Am Rev Respir Dis* 1993; 147:1279-1282.
 102. Milla CE. Long-term effects of aerosolised rhDNase on pulmonary disease progression in patients with cystic fibrosis. *Thorax* 1998; 53:1014-1017.
 103. Hodson ME. Aerosolized dornase alfa (rhDNase) for therapy of cystic fibrosis. *Am J Respir Crit Care Med* 1995; 151:S70-S74.
 104. Haynes A, Shaik MS, Chatterjee A, Singh M. Evaluation of an aerosolized selective COX-2 inhibitor as a potentiator of doxorubicin in a non-small-cell lung cancer cell line. *Pharm Res* 2003; 20:1485-1495.
 105. Tian Y, Klegerman ME, Hickey AJ. Evaluation of microparticles containing doxorubicin suitable for aerosol delivery to the lungs. *PDA J Pharm Sci Technol* 2004; 58:266-275.
 106. Laube BL. Treating diabetes with aerosolized insulin. *Chest* 2001; 120:S99-S106.
 107. Katz IM, Schroeter JD, Martonen TB. Factors affecting the deposition of aerosolized insulin. *Diabetes Technol*

- Ther 2001; 3:387-397.
108. Clark A, Foulds GH, inventors; (Inhale Therapeutic Systems, Inc., USA). assignee. Aerosolized active agent delivery. Application: WO patent 99-US4654 9947196. 1999 19990311.
 109. Cheatham WW, Leone-Bay A, Grant M, Fog PB, Diamond DC, inventors; (Mannkind Corporation, USA). assignee. Pulmonary delivery of inhibitors of phosphodiesterase type 5. Application: WO patent 2005-US30028 2006023944. 2006 20050823.
 110. Calfee DP, Hayden FG. New approaches to influenza chemotherapy. Neuraminidase inhibitors. *Drugs* 1998; 56:537-553.
 111. Cass LM, Gunawardena KA, Macmahon MM, Bye A. Pulmonary function and airway responsiveness in mild to moderate asthmatics given repeated inhaled doses of zanamivir. *Respir Med* 2000; 94:166-173.
 112. Davison S, Thippahawong J, Blanchard J, Liu K, Morishige R, Gonda I, Okikawa J, Adams J, Evans A, Otulana B, Davis S. Pharmacokinetics and acute safety of inhaled testosterone in postmenopausal women. *J Clin Pharmacol* 2005; 45:177-184.
 113. Matilainen L, Järvinen K, Toropainen T, Näsi E, Auriola S, Järvinen T, Jarho P. *In vitro* evaluation of the effect of cyclodextrin complexation on pulmonary deposition of a peptide, cyclosporin A. *Int J Pharm* 2006; 318:41-48.
 114. Waldrep JC, Arppe J, Jansa KA, Vidgren M. Experimental pulmonary delivery of cyclosporin A by liposome aerosol. *Int J Pharm* 1998; 160:239-250.
 115. Yang T, Mustafa F, Bai S, Ahsan F. Pulmonary delivery of low molecular weight heparins. *Pharm Res* 2004; 21:2009-2016.
 116. Qi Y, Zhao G, Liu D, Shriver Z, Sundaram M, Sengupta S, Venkataraman G, Langer R, Sasisekharan R. Delivery of therapeutic levels of heparin and low-molecular-weight heparin through a pulmonary route. *Proc Natl Acad Sci U S A* 2004; 101:9867-9872.
 117. Dershwitz M, Walsh JL, Morishige RJ, Connors PM, Rubsamen RM, Shafer SL, Rosow CE. Pharmacokinetics and pharmacodynamics of inhaled versus intravenous morphine in healthy volunteers. *Anesthesiology* 2000; 93:619-628.
 118. Verkerk V, Blom-Ross ME, Van Dort K, De Vos D, inventors; (Pharmachemie B.V., Neth.). assignee. Pharmaceutical preparation for inhalation of an opioid. Application: WO patent 98-NL713 2000035417. 2000 19981211.
 119. Ward ME, Woodhouse A, Mather LE, Farr SJ, Okikawa JK, Lloyd P, Schuster JA, Rubsamen RM. Morphine pharmacokinetics after pulmonary administration from a novel aerosol delivery system. *Clin Pharmacol Ther* 1997; 62:596-609.
 120. Shafer SL, Hung OR, Pliura DH, inventors; (Delex Therapeutics Inc., Can.). assignee. Opioid delivery system for pulmonary administration. Application: US patent 2004-927145 2005084523. 2005 20040827.
 121. Shiffman S, Fant RV, Buchhalter AR, Gitchell JG, Henningfield JE. Nicotine delivery systems. *Expert Opin Drug Deliv* 2005; 2:563-577.
 122. Andrus PG, Rhem R, Rosenfeld J, Dolovich MB. Nicotine microaerosol inhaler. *Can Respir J* 1999; 6:509-512.
 123. Graham AN, Johnson ES, Persaud NP, Turner P, Wilkinson M. The systemic availability of ergotamine tartrate given by three different routes of administration to healthy volunteers. *Progress in Migraine Research* 1984; 2:283-292.
 124. Bennett DB, Tyson E, Mah S, de Groot JS, Hedge JS, Jerao S, Teitelbaum Z. Sustained delivery of detirelix after pulmonary administration of liposomal formulations. *J Cont Rel* 1994; 32:27-35.

(Received November 4, 2008; Accepted November 6, 2008)

Brief Report

Effect of drug-polymer binary mixtures on the *in-vitro* release of ibuprofen from transdermal drug-in-adhesive layers

Kwong Yat Ho, Michael Ord, Kalliopi Dodou*

Department of Pharmacy Health & Well-being, University of Sunderland, UK.

ABSTRACT: We report on the formation of eutectic mixtures of ibuprofen using two different polymers together with investigations on the *in-vitro* release of ibuprofen from drug-in-adhesive layers. Ibuprofen, literature melting point (m.p.) = 73.5-76.5°C, was tested together with Pluronic F127, literature m.p. = 54.4-60.5°C, and polyethylene glycol 1000 (PEG 1000), literature m.p. = 37-40.9°C, as second components in binary mixtures, incorporated into an acrylic adhesive, either as solid physical mixtures (PM) or molten mixtures (MM). Studies of how the type of mixture preparation (PM versus MM) and the ratio of components in binary mixtures affecting the *in-vitro* drug release of ibuprofen, compared with ibuprofen-adhesive layers without polymer addition were conducted. Ibuprofen release did not improve using the eutectic composition with Pluronic F127, possibly due to increased ibuprofen solubilisation in the adhesive and a subsequent decrease in the thermodynamic activity of the formulation. A significant increase in ibuprofen release ($P < 0.05$) was shown for compositions adjacent to the eutectic one, with ibuprofen: Pluronic F127 (40:60) and ibuprofen: PEG 1000 (20:80, 25:75, 30:70), from both PM- and MM-adhesive formulations, compared to the ibuprofen-adhesive formulations.

Keywords: Transdermal patches, Drug-in-adhesive, Ibuprofen, Eutectic mixture, Thermodynamic activity

1. Introduction

The advantages of transdermal drug delivery include avoidance of the gastrointestinal tract, sustained drug release and increased patient compliance.

*Correspondence to: Dr. Kalliopi Dodou, Department of Pharmacy Health & Well-being, University of Sunderland, Wharnccliffe Street, SR1 3SD, Sunderland, UK;
e-mail: kalliopi.dodou@sunderland.ac.uk

The barrier properties of the stratum corneum mean however that only certain drugs with specific physicochemical properties can be formulated into therapeutically effective passive transdermal patches. There are several strategies to further enhance the passive transdermal delivery of drugs including: the incorporation of chemical enhancers to the formulation, prodrug designs, attainment of maximum thermodynamic activity *via* ensuring saturated concentration of the drug in the formulation and a decrease of the melting point of the drug in the formulation (1).

The transdermal flux of a drug is proportional to the concentration of dissolved drug in the formulation; maximum flux being achieved at saturated drug concentrations. The tendency of the drug to crystallise on storage however eventually renders the formulation thermodynamically unstable with a subsequent decrease in drug flux (2). Several additives in monolayer transdermal (drug-in-adhesive) patches have been shown to decrease or prevent crystallisation of the drug (3). The melting point suppression strategy is based on the "ideal solution theory". This states that the lower the melting point of a drug, the greater the drug solubility in skin lipids (4). The formation of a eutectic binary mixture between the drug and an additional component is a well-known technique by which the melting point of the drug can be suppressed in the formulation. This has been previously examined as a potential method to enhance topical and transdermal drug delivery (5,6). There is however no published literature on the incorporation of eutectic mixtures into transdermal adhesive monolayers.

A eutectic binary mixture is a solid dispersion of two components at a specific ratio at which it possesses a lower melting point than either of the components and the other component ratios. At temperatures below this melting point the two components exist as an intimate microcrystalline mixture that melts uniformly at the melting temperature. At temperatures above the melting point the two components exist as a uniform melt that inhibit crystallisation of one another. Therefore a

eutectic composition may confer stability to the drug against crystallisation.

In our study we used ibuprofen as the model drug and two hydrophilic polymers with low melting points, PEG 1000 and Pluronic F127 that would enable a considerable suppression of the melting point of the drug. Ibuprofen is a non steroidal anti-inflammatory drug (NSAID) with analgesic and anti-inflammatory properties. The main side effect of the oral administration of NSAIDs is irritation of the gastrointestinal wall lining, which can lead to the development of ulcers following long-term administration. For this reason ibuprofen and other NSAIDs have been studied extensively as candidates for systemic delivery *via* the transdermal route (7). PEGs are non-irritant nor toxic to healthy skin and do not readily penetrate it (8). Poloxamers have previously been used as a vehicle for the topical delivery of NSAIDs due to their low toxicity and irritation (9).

Binary eutectic mixtures of ibuprofen with Pluronic F127 and PEG 1000 were formulated into drug-in-adhesive layers containing binary drug-polymer mixtures at several ratios, including the eutectic ratio. Hot stage microscopy (HSM) was used to study the melting properties of the solid dispersions and identify the eutectic composition. The HSM technique has been shown to be more efficient than differential scanning calorimetry in detecting the presence of drug crystals in solid dispersions and differences in the melting behaviour among samples, especially when a polymer with a low melting point is used as a drug carrier (10,11). The binary mixtures were prepared and incorporated in the adhesive layer according to two different methods; either as physical mixtures (PM) or as molten mixtures (MM) of the two components that would solidify after incorporation into the adhesive layer. The aim of our work was to study how the method of mixture preparation, as a PM or MM, and the ratio of components in the binary mixtures influence the *in-vitro* drug release of ibuprofen, compared with ibuprofen-adhesive layers without additive.

2. Materials and Methods

2.1. Materials

Ibuprofen was obtained from Knoll Pharmaceuticals (Nottingham, UK). DURO-TAK[®] 87-4287 was a gift from National Adhesives-Henkel (Slough, UK). Polyethylene glycol with an average molecular weight of 1000 Da (PEG 1000) was supplied by Sigma (St. Louis, USA). Pluronic F127 was supplied as Lutrol[®] F127 and was a gift from BASF AG (Ludwigshafen, Germany). The Scotchpak 9742 release liner was a gift from 3M Corporation (St. Paul, USA).

2.2. Preparation of solid dispersions

Solid dispersions of ibuprofen with either Pluronic F127 or PEG 1000 were prepared in ratios ranging from 90%:10% to 10%:90% (w/w) according to the fusion method. Appropriate amounts of ibuprofen and polymer to give a 2 g mixture were accurately weighed in test tubes and were placed in a water bath (Techne Inc., Princeton, USA) with a VMR 1122S temperature control. The initial temperature of the water bath was 30°C, gradually increased to 75°C, at a rate of 3°C/min, while the drug-polymer mixtures were gently stirred with a glass rod. The molten mixtures were then allowed to solidify at 20°C for a week.

2.3. Hot stage microscopy

The melting temperature of the solid dispersions, ibuprofen, Pluronic F127 and PEG 1000 were recorded using a Vickers microscope attached to a Mettler FP5 hot stage temperature control and recorder. The temperature range was set from 25 to 80°C with a heating rate of 2°C/min. Each solid dispersion was tested in triplicate ($n = 3$). Two temperatures were recorded per sample, the first being the initial temperature when melting began (lower limit) and the second being the temperature that melting was complete (upper limit). Phase diagrams were then plotted and the eutectic compositions were identified.

2.4. Preparation of the adhesive layers

The following binary mixtures of ibuprofen:polymer were prepared as physical mixtures and solid dispersions: 60:40, 40:60, 30:70 with Pluronic F127, and 60:40, 30:70, 25:75, 20:80 with PEG 1000. A binary mixture that would contain 0.05 g of ibuprofen was accurately weighed and added to the required amount of liquid acrylic adhesive to produce dried circular adhesive layers with an ibuprofen concentration of 10% (w/w). Ibuprofen-adhesive layers without polymer were also prepared by mixing 0.05 g of either solid or molten ibuprofen with the acrylic adhesive to produce layers of 10% (w/w) ibuprofen concentration. All layers had a mean surface area of 4.5 ± 0.35 cm², with one side attached to a release liner. The layers were stored for a week at 20°C before dissolution testing.

2.5. In-vitro drug release studies

In-vitro drug release studies were conducted according to the B.P. Dissolution method for transdermal patches. The release of ibuprofen from each set of layers ($n = 3$) was tested for 5 h using a paddle dissolution apparatus (Copley instruments Ltd, Nottingham, UK) with 900 mL of citrophosphate buffer (pH 5.6) as the dissolution medium under sink conditions. The temperature of the

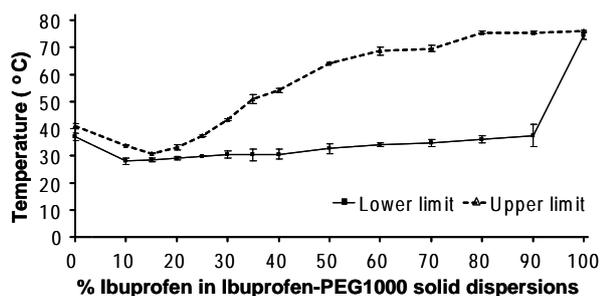


Figure 1. Phase diagram showing the melting behaviour of ibuprofen-PEG 1000 solid dispersions. Lower limit points show the mean temperature ($n = 3$) reading where melting of the sample started; upper limit points show the mean temperature ($n = 3$) reading where melting of the sample was complete. Error bars represent the standard deviation from each mean temperature reading.

dissolution medium was maintained at $32 \pm 0.5^\circ\text{C}$ and the paddle stirring rate was set at 50 rpm. 10 mL samples of dissolution medium were withdrawn from each vessel every 10 min during the 1st hour and then every 30 min up to 5 h. The ibuprofen content of the samples was analysed using a CE272 Linear Readout Ultraviolet (UV) Spectrophotometer (CECIL instruments Ltd, Cambridge, UK) at a wavelength of 272 nm.

2.6. Statistical analysis

The % cumulative amount of ibuprofen released ($n = 3$) at 5 h was plotted against time. Statistical differences were determined using a Student *t*-test (two independent samples) with significance at $P < 0.05$.

3. Results and Discussion

The eutectic composition of ibuprofen with PEG 1000 was found at ibuprofen:PEG 1000 ratio of 15:85, with a melting point of 30.9°C (Figure 1). This temperature is lower than normal skin temperature (32°C), implying that ibuprofen will be in liquid form when applied onto the skin. Theoretically this would favour drug permeation into the stratum corneum (4). The eutectic composition of ibuprofen with Pluronic F127 was found at the ibuprofen:Pluronic F127 ratio of 30:70, with a melting point of 46°C (Figure 2).

The drug concentration in the adhesive layers (10%, w/w) was selected to be lower than the saturation solubility of ibuprofen in the adhesive polymer, in order to avoid suppression of drug release by drug crystallization and, thus, observe only the effect of binary mixtures on drug release. All layers prepared with the MM method were transparent in appearance with no drug crystals observed. The layers prepared with the PM method contained undissolved polymer dispersed in the adhesive layer. When the eutectic composition of ibuprofen with PEG 1000 (15:85) was incorporated into the adhesive as a molten mixture, it did not solidify on cooling but remained as liquid and leaked out of the borders of the adhesive layers

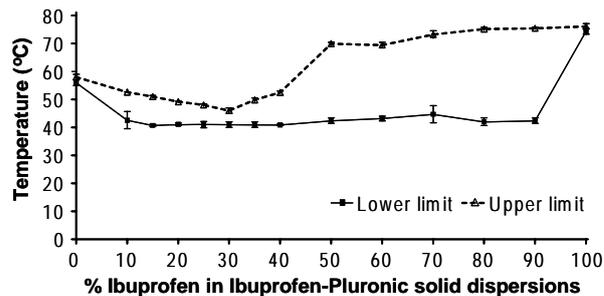


Figure 2. Phase diagram showing the melting behaviour of ibuprofen-Pluronic F127 solid dispersions. Lower limit points show the mean temperature ($n = 3$) reading where melting of the sample started; upper limit points show the mean temperature ($n = 3$) reading where melting of the sample was complete. Error bars represent the standard deviation from each mean temperature reading.

on storage. For this reason it was not possible to carry out dissolution studies with the ibuprofen-PEG 1000 eutectic composition.

% Cumulative ibuprofen release at 5 h was statistically higher ($P < 0.05$) for the ibuprofen:PEG 1000 ratios 30:70, 25:75 and 20:80 compared to the ibuprofen monolayer 100:0 and the 60:40 ratio, for both PM and MM (Figures 3 and 4). Similarly, the ibuprofen:Pluronic F127 composition with the significantly higher ibuprofen release ($P < 0.05$) for both MM and PM was the 40:60 ratio, which is adjacent to the eutectic composition (Figures 5 and 6).

The eutectic composition of ibuprofen with Pluronic F127 (30:70) showed lower drug release compared to the formulation containing ibuprofen alone (Figures 5 and 6). This could be attributed to the fact that the eutectic mixture increases the solubility of ibuprofen in the adhesive layer and so simultaneously decreases the thermodynamic activity of the formulation. This is in agreement with previous observations demonstrating that a decrease in the melting point of a compound *via* formation of a binary eutectic system can be used as an approach for increasing the drug solubility in the vehicle (5).

The agreement between MMs and PMs on the order of enhancing drug release using either PEG 1000 or Pluronic F127 is noteworthy indicating an interaction taking place. An interaction between the components of binary physical mixtures during mixing has been previously reported (12). In our case, the incorporation of the binary physical mixtures in the adhesive solution may have resulted in ibuprofen-polymer solid dispersion formation during drying of the monolayer.

In conclusion, our results showed that ibuprofen release was enhanced by binary mixtures adjacent to the eutectic composition that contain a higher proportion of ibuprofen than the eutectic. This enhanced ibuprofen release could be observed up to a certain ratio, after which any further increase in the amount of ibuprofen in the binary mixture showed no significant difference ($P > 0.05$) on drug release compared to the drug-adhesive alone. Our

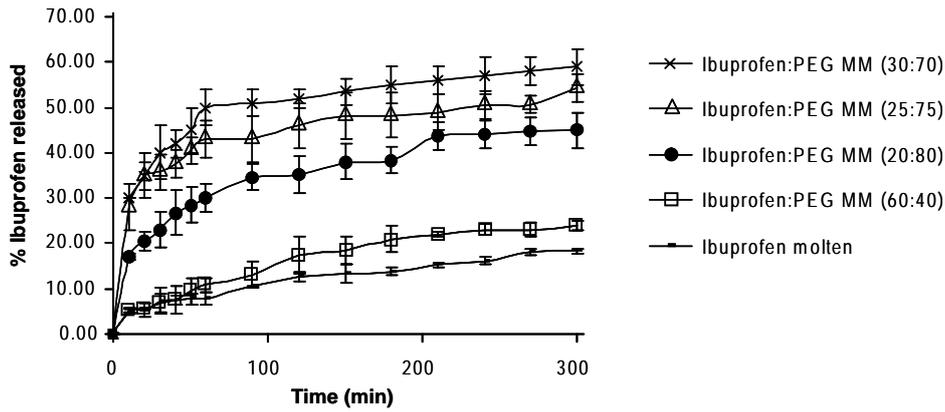


Figure 3. % Ibuprofen released from the adhesive layers containing MM of ibuprofen with PEG 1000. Error bars represent the standard deviation from the mean ($n = 3$).

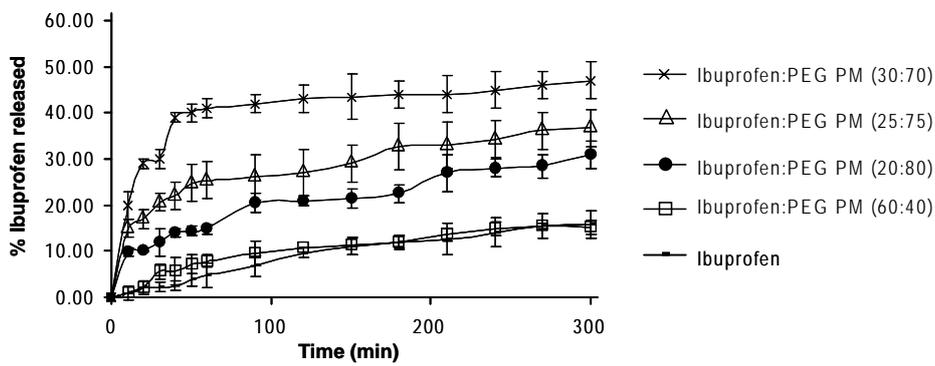


Figure 4. Ibuprofen released from the adhesive layers containing PM of ibuprofen with PEG 1000. Error bars represent the standard deviation from the mean ($n = 3$).

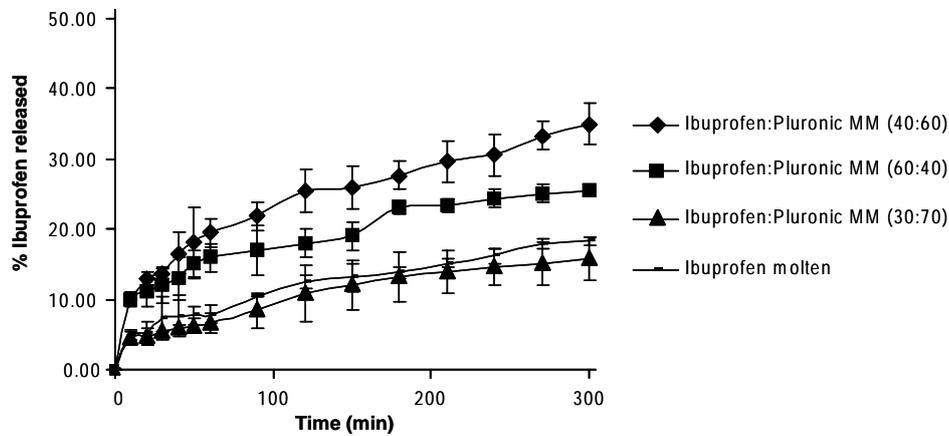


Figure 5. % Ibuprofen released from the adhesive layers containing MM of ibuprofen with Pluronic F127. Error bars represent the standard deviation from the mean ($n = 3$).

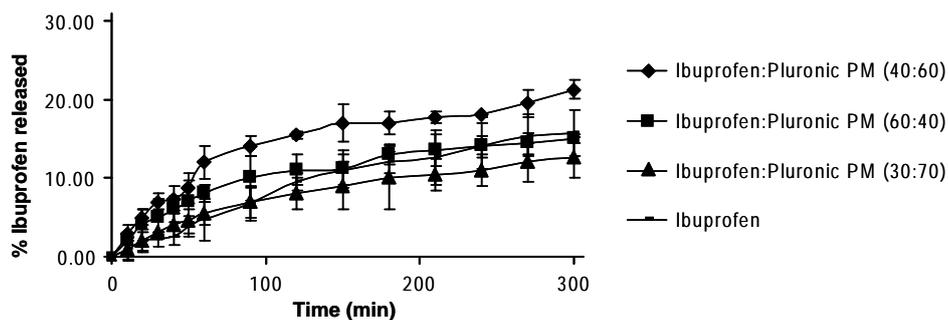


Figure 6. % Ibuprofen released from the adhesive layers containing PM of ibuprofen with Pluronic F127. Error bars represent the standard deviation from the mean ($n = 3$).

results also support the hypothesis that incorporation of an additional component as a eutectic mixture with the drug in the adhesive monolayer will increase the solubility of the drug in the adhesive, with a subsequent decrease in thermodynamic activity for the given ibuprofen concentration in the monolayer. This may indicate that using the eutectic composition, higher ibuprofen concentrations can be accommodated in the transdermal monolayer without compromising the stability of the formulation, considering the inherent stability of eutectic mixtures against crystallization.

Acknowledgements

The authors would like to thank National Adhesives-Henkel, 3M and BASF for their kind provision of materials.

References

1. Hadgraft J. Passive enhancement strategies in topical and transdermal drug delivery. *Int J Pharm* 1999; 184:1-6.
2. Iervolino M, Cappello B, Raghavan SL, Hadgraft J. Penetration enhancement of ibuprofen from supersaturated solutions through human skin. *Int J Pharm* 2001; 212:131-141.
3. Cilurzo F, Minghetti P, Casiraghi A, Tosi L, Pagani S, Montanari L. Polymethacrylates as crystallization inhibitors in monolayer transdermal patches containing ibuprofen. *Eur J Pharm Biopharm* 2005; 60:61-66.
4. Kasting GB, Smith RL, Cooper ER. Effects of lipid solubility and molecular size on percutaneous absorption. *Pharmacol Skin* 1987; 1:138-153.
5. Kaplun-Frischoff Y, Touitou E. Testosterone skin permeation enhancement by menthol through formation of eutectic with drug and interaction with skin lipids. *J Pharm Sci* 1997; 86:1394-1399.
6. Stott PW, Williams AC, Barry BW. Transdermal delivery from eutectic systems: enhanced permeation of a model drug, ibuprofen. *J Controlled Release* 1998; 50:297-308.
7. Beetge E, Du Plessis J, Müller DG, Goosen C, Van Rensberg FJ. The influence of the physicochemical characteristics and pharmacokinetic properties of selected NSAIDs on their transdermal absorption. *Int J Pharm* 2000; 193:261-264.
8. Price JC. Polyethylene Glycol. In: *Handbook of pharmaceutical excipients* (Kibbe AH ed.). Pharmaceutical Press & American Pharmaceutical Association, London, Washington, 2000; pp. 392-398.
9. Shin SC, Cho CW, Oh IJ. Enhanced efficacy by percutaneous absorption of piroxicam from the poloxamer gel in rats. *Int J Pharm* 2000; 193:213-218.
10. Fini A, Moyano JR, Ginés JM, Perez-Martinez JJ, Rabasco AM. Diclofenac salts II. Solid dispersions in PEG6000 and Gelucire 50/13. *Eur J Pharm Biopharm* 2005; 60:99-111.
11. Bikiaris D, Papageorgiou GZ, Stergiou A, Pavlidou E, Karavas E, Kanaze F, Georgarakis M. Physicochemical studies on solid dispersions of poorly water-soluble drugs: Evaluation of capabilities and limitations of thermal analysis techniques. *Thermochimica Acta* 2005; 439:58-67.
12. Shakhtshneider TP, Vasilchenko MA, Politov AA, Boldyrev VV. The mechanochemical preparation of solid disperse systems of ibuprofen-polyethylene glycol. *Int J Pharm* 1996; 130:25-32.

(Received October 28, 2008; Accepted October 31, 2008)

Brief Report**Effect of benzyl-*N*-acetyl- α -galactosaminide on KL-6 mucin expression and invasive properties of a human pancreatic carcinoma cell line****Huanli Xu^{1,2}, Yoshinori Inagaki¹, Fengshan Wang^{2,*}, Norihiro Kokudo¹, Munehiro Nakata³, Wei Tang^{1,2,*}**¹ Hepato-Biliary-Pancreatic Surgery Division, Department of Surgery, Graduate School of Medicine, the University of Tokyo, Tokyo, Japan;² Institute of Biochemical and Biotechnological Drugs, School of Pharmaceutical Sciences, Shandong University, Ji'nan, China;³ Department of Applied Biochemistry, Tokai University, Kanagawa, Japan.

ABSTRACT: KL-6 mucin is a type of MUC1 mucin and its aberrant expression has been shown to be associated with aggressive metastasis and poor clinical outcome in tumors. The present study is to investigate the effects of benzyl-*N*-acetyl- α -galactosaminide (GalNAc-*O*-bn), an *O*-glycosylation inhibitor, on KL-6 mucin expression and invasive properties of a human pancreatic carcinoma cell line, Suit-2 cells. Expression profiles of KL-6 mucin in the cells pretreated with or without 5 mM GalNAc-*O*-bn for 48 h were examined by Western blotting and immunocytochemical staining and invasive properties were examined by transwell chamber assay. Western blotting and immunocytochemical staining showed that the expression profiles of KL-6 mucin changed significantly after GalNAc-*O*-bn treatment. Meanwhile, the invasive ability of Suit-2 cells decreased significantly after GalNAc-*O*-bn treatment ($p < 0.05$). These results suggest that glycosylation of KL-6 mucin may be closely related to aggressive behaviors of pancreatic cancer cells like metastasis and invasion.

Keywords: KL-6 mucin, *O*-glycosylation, Benzyl-*N*-acetyl- α -galactosaminide, Invasion, Pancreatic carcinoma

*Correspondence to: Dr. Wei Tang, Hepato-Biliary-Pancreatic Surgery Division, Department of Surgery, Graduate School of Medicine, the University of Tokyo, Hongo 7-3-1, Tokyo 113-8655, Japan; e-mail: TANG-SUR@h.u-tokyo.ac.jp;
Dr. Fengshan Wang, Institute of Biochemical and Biotechnological Drugs, School of Pharmaceutical Sciences, Shandong University, Ji'nan 250012, Shandong Province, China; e-mail: fswang@sdu.edu.cn

1. Introduction

Patients with pancreatic cancer still have a poor prognosis, a 5-year survival rate of ~3% and a median survival of < 6 months (1), although recent efforts have improved cancer prevention, screening, and therapy. The major problem in the management of postsurgical cases is failure to control cancer metastases, which results from a lack of early detection and effective treatment.

MUC1 is a polymorphic, highly glycosylated, type I transmembrane glycoprotein expressed by ductal epithelial cells of secretory organs, including the pancreas, breast, lung, and gastrointestinal tract, that is overexpressed and aberrantly glycosylated in most cases of adenocarcinoma (2). The deduced amino acid sequence of MUC1 mucin reveals four distinct domains: an NH₂-terminal domain consisting of a hydrophobic signal sequence, a highly *O*-glycosylated tandem-repeat domain, a transmembrane domain, and a cytoplasmic domain (3). A number of studies have suggested that overexpression of MUC1 plays an important role in pancreatic cancer metastasis and that MUC1 seems to be an attractive target for treatment of pancreatic cancer (4,5). Therefore, having MUC1 target monoclonal antibody (mAb) with high specificity and affinity may represent an effective strategy.

KL-6 mucin, a type of MUC1 categorized as cluster 9, is recognized by KL-6 mAb, and its epitope includes sialo-oligosaccharide moiety in MUC1 molecules (6,7). This mucin was first established in the serum of patients with intestinal pneumonia but has recently been detected in various cancer tissues (8,9). Previous immunohistochemical studies by the current authors have shown that overexpression of KL-6 mucin was associated with worse tumor behaviors such as invasion and metastasis in ampullary carcinoma, primary

colorectal carcinoma, and metastatic liver cancer tissues (10-12). In addition, the relationship between expression of KL-6 mucin and metastatic potential has also been reported in colorectal carcinoma cell lines (13). However, the role of glycosylation in KL-6 mucin in tumor behaviors remains to be elucidated. Reports have indicated that extension of *O*-glycosylation in cultured cells can be blocked in the presence of benzyl-*N*-acetyl- α -galactosaminide (GalNAc-*O*-bn) (14-16). The present study aimed to evaluate the effect of the *O*-glycosylation inhibitor GalNAc-*O*-bn on the expression profiles of KL-6 mucin and the invasive properties of the human pancreatic carcinoma Suit-2 cell line.

2. Materials and Methods

2.1. Reagents

GalNAc-*O*-bn, trypsin, cell lysis reagents, trypan blue, and culture media were purchased from Sigma-Aldrich Japan, Tokyo, Japan. KL-6 mAb was provided by Eisai Co., Ltd, Tokyo, Japan. The Histofine SAB-PO kit and BD BioCoat™ Tumor Invasion System were from Nichirei Corporation, Tokyo, Japan and BD Biosciences, San Jose, CA, USA, respectively.

2.2. Cells and culture conditions

Human pancreatic carcinoma Suit-2 cell line was obtained from JCRB Cell Bank, Tokyo, Japan. The cells were cultured in RPMI 1640 supplemented with 10% heat-inactivated fetal bovine serum (FBS) in a humidified atmosphere with 5% CO₂ in air at 37°C. For maintenance and subculture, cells in the exponential phase were treated with 0.25% trypsin solution containing 0.02% EDTA. Cells reaching 80% of confluence were collected and subjected to the following experiment.

2.3. Cell proliferation assay

Cells were reseeded in 96-well plates (3 × 10³ cells per well) incubated with 0, 1, 2, 4, 8, and 16 mM of GalNAc-*O*-bn for 48 h. Cell proliferation was assessed by 3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyl-tetrazolium bromide (MTT) assay as described elsewhere (17). Absorbance was detected at 550 nm with 750 nm as a reference wavelength.

2.4. Western blotting

Cells treated with or without 5 mM of GalNAc-*O*-bn for 48 h at 37°C were lysed in lysis buffer (50 mM Tris-HCl, pH 7.4, containing 1% NP-40, 0.25% Na-deoxycholate, 150 mM NaCl, 1 mM EDTA, 1 mM phenylmethylsulfonyl fluoride, 1 mM Na₃VO₄, 1 mM

NaF, and 1 µg/mL each of aprotinin, leupeptin, and pepstatin) with occasional sonication. After centrifuging at 12,000 × g for 30 min at 4°C, cell extracts (10 µg protein each) were subjected to SDS-PAGE (8% gel) and then electrotransferred onto polyvinylidene difluoride membranes. After blocking with 20 mM Tris-buffered saline containing 5% nonfat dry milk and 0.1% Tween-20 overnight at 4°C, the membranes were incubated with KL-6 mAb (1:750 dilution) for 1 h at room temperature. After they were washed three times, the membranes were incubated with horse radish peroxidase-conjugated secondary antibodies for 1 h at room temperature. Reactivity was visualized by enhanced chemiluminescence using the ECL Western Blotting Starter Kit (RPN2108; GE Healthcare Bio-Sciences, Piscataway, NJ, USA). Data were obtained from three independent experiments.

2.5. Immunocytochemistry

Cells (1 × 10⁴) treated with or without GalNAc-*O*-bn were seeded on MAS-coated slides (Matsunami, Inc., Osaka, Japan) and incubated for 4 h. Cells were then fixed with 3.7% paraformaldehyde for 30 min followed by treatment with 0.2% Triton X-100 for 10 min. Next, immunohistochemical staining by KL-6 mAb was done as described before (10). Briefly, cells were blocked with normal goat serum for 30 min at room temperature, and then incubated with or without KL-6 antibody (1:200 dilution) for 60 min at room temperature. After the incubation of biotin-labeled secondary antibody, detection of KL-6 mucin was achieved by the biotin-streptavidin-peroxidase complex method using Histofine SAB-PO kit. 3,3'-Diaminobenzidine was used as the chromogen, and haematoxylin was used as a counterstain.

2.6. Transwell chamber assay

Motility and the invasive abilities of cells were assessed in 24-well transwell plates (Corning, NY, USA). Cells were pre-incubated with or without 5 mM of GalNAc-*O*-bn for 48 h at 37°C in a CO₂ incubator and then detached and resuspended in serum-free RPMI-1640. A suspension of cells (2 × 10⁵ cells/mL) was placed on matrigel-coated filters and control filters, respectively. The lower chambers were filled with 0.75 mL of RPMI-1640 medium supplemented with 5% FBS. Cells were allowed to migrate for 22 h at 37°C. Cells that invaded the matrigel and reached the opposite surface of the filter were stained with a Diff-Quik kit (Dade Behring, Newark, DE, USA) in accordance with the manufacturer's instructions. The invaded cells were quantified by counting the number of cells in eight random microscopic fields per filter at a magnification of ×100. Each data point was calculated from two separate experiments performed

in triplicate. A *p* value less than 0.05 was considered statistically significant.

3. Results and Discussion

Control of metastatic pancreatic cancer remains a major objective in pancreatic cancer treatment. The overexpression of MUC1 mucin plays an important role in the process of pancreatic cancer metastasis and invasion. MUC1 targeting mAb with high specificity and affinity might represent a useful targeted therapy. KL-6 mucin is a type of MUC1 recognized by KL-6 mAb, and its epitope includes sialo-oligosaccharide moiety in MUC1 molecules (6-9). Many studies have shown that overexpression of KL-6 mucin may be associated with worse tumor behaviors such as invasion and metastasis in many kinds of cancers (6,10-12). Since MUC1 is a highly *O*-glycosylated cell surface glycoprotein, GalNAc-*O*-bn, an *O*-glycosylation inhibitor, was used to inhibit *O*-linked oligosaccharide of KL-6 mucin. GalNAc-*O*-bn is a synthetic analogue of *N*-acetylgalactosamine and inhibits elongation of *O*-glycans. The inhibition is competitive and instead of monosaccharide transfer in which GalNAc is bound to serine or threonine, the elongation of the *O*-glycan chain occurs in benzyl-*N*-acetyl- α -galactosaminide molecules (18,19).

First, the inhibitory effects of GalNAc-*O*-bn on Suit-2 cell proliferation were examined using MTT assay. Inhibition of Suit-2 cell proliferation was not observed in the presence of up to 16 mM GalNAc-*O*-bn (data not shown). Therefore, a concentration of 5 mM was used in the following experiments as reported previously (18). Cell viability, as judged by trypan blue exclusion, was also unaffected in the presence of 5 mM GalNAc-*O*-bn (data not shown).

Using the KL-6 antibody, KL-6 mucin expression was next detected in Suit-2 cells with or without GalNAc-*O*-bn treatment by means of Western blotting. As shown in Figure 1A, KL-6 mucin was detected with variations in molecular weight. Notably, the intensity of the major band around 209 kDa was significantly lower after treatment with 5 mM GalNAc-*O*-bn. In contrast, the intensity of other two bands with a molecular weight of around 110 and 178 kDa, respectively, seemed to be higher in GalNAc-*O*-bn treated cells than in the untreated cells. To examine the subcellular expression of KL-6 mucin, Suit-2 cells with or without GalNAc-*O*-bn treatment were subjected to immunocytochemical staining. Before GalNAc-*O*-bn treatment, strong KL-6 mucin stains were observed in Suit-2 cells in the entire cell, including its membrane and cytoplasm (Figure 1B). After GalNAc-*O*-bn treatment, KL-6 mucin stains decreased significantly especially in the cell membrane. Cell morphology was also found to have changed after GalNAc-*O*-bn treatment (Figure 1B).

One of the earliest steps in metastasis is the invasion

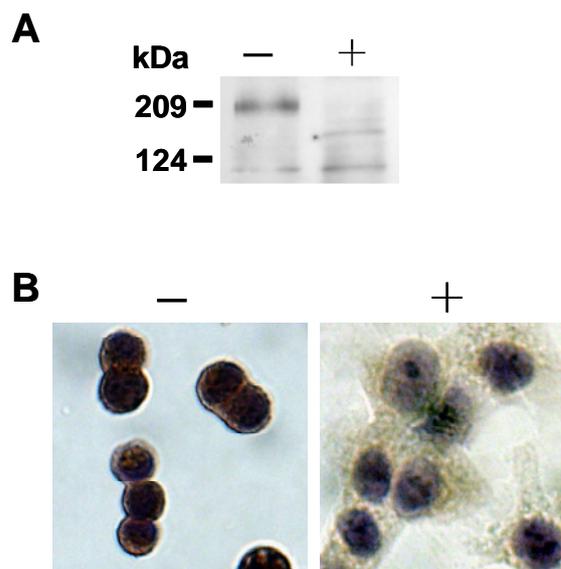


Figure 1. Western blotting and immunocytochemical analyses for KL-6 mucin expression in Suit-2 cells treated with (+) or without (-) GalNAc-*O*-bn. Suit-2 cells were pretreated with or without 5 mM GalNAc-*O*-bn for 48 h at 37°C, and then subjected to Western blotting (A) and immunocytochemical analysis (B) as described in Materials and Methods. Original magnification of B, $\times 400$.

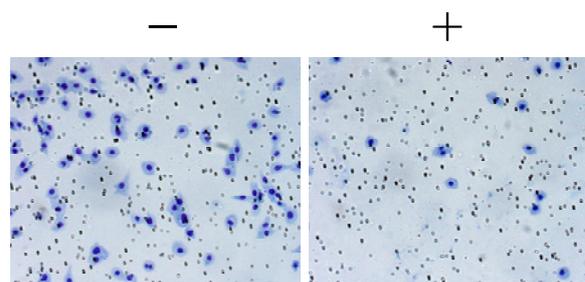


Figure 2. Inhibitory effect of GalNAc-*O*-bn on Suit-2 cell invasion. Suit-2 cells pretreated with (+) or without (-) 5 mM GalNAc-*O*-bn were placed on matrigel-coated chambers and incubated for 22 h at 37°C. Original magnification, $\times 100$.

of the basement membrane. Next evaluated was the effect of GalNAc-*O*-bn on the motility and invasive ability of Suit-2 cells migrating through a matrigel-coated polycarbonate membrane. As shown in Figure 2, the invasive potential of Suit-2 cells decreased significantly after 5 mM GalNAc-*O*-bn treatment. The percentage of cells penetrating the matrigel-coated polycarbonate filters without or with GalNAc-*O*-bn treatment was 93.5% and 10.3%, respectively ($p < 0.05$).

The present study suggests that GalNAc-*O*-bn, an *O*-glycosylation inhibitor, may significantly alter the expression profiles of KL-6 mucin, especially on the cell surface, and it may alter the invasive ability of Suit-2 cells. Since KL-6 mAb recognizes sialo-oligosaccharides in addition to part of the core polypeptide of KL-6 mucin (8), the decrease in KL-6 mucin expression as a result of treatment with GalNAc-*O*-bn may be due to the insufficient elongation of oligosaccharide chains. Although the present study did

not examine the changes in levels of expression of the KL-6 mucin core polypeptide for GalNAc-*O*-bn-treated and untreated cells, the current findings suggest that an extracellular domain containing sialo-oligosaccharide chains of KL-6 mucin may play a role in the invasive ability of the cells.

GalNAc-*O*-bn has been reported to inhibit *O*-glycosylation in other cell surface glycoprotein such as brush border glycoprotein sucrase-isomaltase (20) and to cause morphological changes with an accumulation of GalNAc terminal glycoproteins that may be mucin precursors at the cell surface (21). Further study is needed to clarify the detailed mechanism by which GalNAc-*O*-bn acts on KL-6 mucin expression and the underlying role of KL-6 mucin in the metastatic progression of the pancreatic carcinoma. These experiments may provide *in vitro* evidence for the *O*-glycosylation of KL-6 mucin playing a role in the invasion of cancer cells and portend that therapeutic strategies targeting oligosaccharide moieties of KL-6 mucin should be useful in the treatment of aggressive pancreatic cancer.

Acknowledgments

This study was supported in part by Grants-in-Aid from the Ministry of Education, Science, Sports and Culture of Japan, and JSPS and CAMS under the Japan-China Medical Exchange Program.

References

- Schneider G, Siveke JT, Eckel F, Schmid RM. Pancreatic cancer: basic and clinical aspects. *Gastroenterology* 2005; 128:1606-1625.
- Kim YS, Gum J, Brockhausen I. Mucin glycoproteins in neoplasia. *Glycoconjugate J* 1996; 13:693-707.
- Hollingsworth MA, Swanson BJ. Mucins in cancer: protection and control of the cell surface. *Nature Rev* 2004; 4:45-60.
- Kohlgraf KG, Gawron AJ, Higashi M, Meza JL, Burdick MD, Kitajima S, Kelly DL, Caffrey TC, Hollingsworth MA. Contribution of the MUC1 tandem repeat and cytoplasmic tail to invasive and metastatic properties of a pancreatic cancer cell line. *Cancer Res* 2003; 63:5011-5020.
- Luttges J, Feyerabend B, Buchelt T, Pacena M, Kloppel G. The mucin profile of noninvasive and invasive mucinous cystic neoplasms of the pancreas. *Am J Surg Pathol* 2002; 26:466-471.
- Ogawa Y, Ishikawa T, Ikeda K, Nakata B, Sawada T, Ogasawa K, Kato Y, Hirakawa K. Evaluation of serum KL-6, a mucin-like glycoprotein, as a tumor marker for breast cancer. *Clin Cancer Res* 2006; 6:4069-4072.
- Kohno N, Inoue Y, Hamada H, Fujioka S, Fujino S, Yokoyama A, Hiwada K, Ueda N, Akiyama M. Difference in sero-diagnostic values among KL-6-associated mucins classified as cluster 9. *Int J Cancer* 1994; 8:81-83.
- Kohno N, Akiyama M, Kyoizumi S, Hakoda M, Kobuke K, Yamakido M. Detection of soluble tumor-associated antigens in sera and effusions using novel monoclonal antibodies, KL-3 and KL-6, against lung adenocarcinoma. *Jpn J Clin Oncol* 1988; 18:203-216.
- Kohno N. Serum marker KL-6/MUC1 for the diagnosis and management of interstitial pneumonitis. *J Med Invest* 1999; 46:151-158.
- Tang W, Inagaki Y, Kokudo N, Guo Q, Seyama Y, Nakata M, Imamura H, Sano K, Sugawara Y, Makuuchi M. KL-6 mucin expression in carcinoma of the ampulla of Vater: Association with cancer progression. *World J Gastroenterol* 2005; 11:5450-5454.
- Guo Q, Tang W, Inagaki Y, Midorikawa Y, Kokudo N, Sugawara Y, Nakata M, Konishi T, Nagawa H, Makuuchi M. Clinical significance of subcellular localization of KL-6 mucin in primary colorectal adenocarcinoma and metastatic tissues. *World J Gastroenterol* 2006; 12:54-59.
- Xu HL, Inagaki Y, Tang W, Guo Q, Wang FS, Seyama Y, Midorikawa Y, Gai RY, Kokudo N, Sugawara Y, Nakata M, Makuuchi M. Elevation of serum KL-6 mucin levels in patients with cholangiocarcinoma. *Hepatogastroenterology* 2008; 88: *in press*.
- Guo Q, Tang W, Inagaki Y, Kokudo N, Sugawara Y, Karako H, Nakata M, Makuuchi M. Subcellular localization of KL-6 mucin in colorectal carcinoma cell lines: association with metastatic potential and cell morphology. *Oncol Rep* 2007; 17:1057-1060.
- Delacour D, Gouyer V, Leteurtre E, Ait-Slimane T, Drobecq H, Lenoir C, Moreau-Hannedouche O, Trugnan G, Huet G. 1-Benzyl-2-acetamido-2-deoxy-alpha-D-galactopyranoside blocks the apical biosynthetic pathway in polarized HT-29 cells. *J Biol Chem* 2003; 278:37799-37809.
- Gouyer V, Leteurtre E, Delmotte P, Steelant WFA, Krzewinski-Recchi MA, Zanetta JP, Lesuffleur T, Trugnan G, Delannoy P, Huet G. Differential effect of GalNAc- α -*O*-bn on intracellular trafficking in enterocytic HT-29 and Caco-2 cells: relation with the glycosyltransferase expression pattern. *J Cell Sci* 2001; 114:1455-1471.
- Huang J, Byrd JC, Yoor WH, Kim YS. Effect of benzyl- α -GalNAc, an inhibitor of mucin glycosylation, on cancer-associated antigens in human colon cancer cells. *Oncol Res* 1992; 4:507-515.
- Zhang L, Hu JJ, Du GH. Establishment of a cell-based assay to screen insulin-like hypoglycemic drugs. *Drug Discov Ther* 2008; 2:229-233.
- Satoh S, Hinoda Y, Hayashi T, Burdick MD, Imai K, Hollingsworthman MA. Enhancement of metastatic properties of pancreatic cancer cells by MUC1 gene encoding an anti-adhesion molecule. *Int J Cancer* 2000; 88:507-518.
- Paszkiwicz-Gadek A, Porowska H, Lemancewicz D, Wolczynski S, Gindzienski A. The influence of *N*- and *O*-glycosylation inhibitors on the glycosylation profile of cellular membrane proteins and adhesive properties of carcinoma cell lines. *Int J Mol Med* 2006; 17:669-674.
- Alfalah M, Jacob R, Preuss U, Zimmer KP, Naim H, Naim HY. *O*-linked glycans mediate apical sorting of human intestinal sucrase-isomaltase through association with lipid rafts. *Curr Biol* 1999; 9:593-596.
- Kuan SF, Byrd JC, Basbaum C, Kim YS. Inhibition of mucin glycosylation by aryl-*N*-acetyl-a-galactosaminides in human colon cancer cells. *J Biol Chem* 1989; 264:19271-19277.

(Received November 9, 2008; Accepted November 21, 2008)

Brief Report**Antioxidant activity of wild plants collected in Beni-Sueif governorate, Upper Egypt**Sameh AbouZid^{1,*}, Abdelaaty Elshahaat², Sajjad Ali³, Mohamed I. Choudhary³¹ Department of Pharmacognosy, Faculty of Pharmacy, Beni-Sueif University, Beni-Sueif, Egypt;² Department of Phytochemistry, National Research Centre, Cairo, Egypt;³ H.E.J. Research Institute of Chemistry, International Center for Chemical and Biological Sciences, University of Karachi, Karachi, Pakistan.

ABSTRACT: Antioxidant activity of a selection of commonly occurring wild plants growing in Beni-Sueif governorate, Upper Egypt, has been tested. The plants selected are *Tamarix nilotica*, *Ambrosia maritima*, *Zygophyllum coccenium*, *Conyza dioscoridis*, *Chenopodium ambrosioides*, and *Calotropis procera*. The *in vitro* antioxidant assays used in this study were 1,1-diphenyl-2-picryl hydrazyl (DPPH) radical scavenging activity, superoxide anion scavenging activity and iron chelating activity. Extracts prepared from the leaves and flowers of *Tamarix nilotica* have shown the highest antioxidant activity in the three kinds of assay.

Keywords: Screening, Wild plant, DPPH, Superoxide anion, Iron chelation, *Tamarix nilotica*

1. Introduction

Plants are a valuable source of natural products. These plant metabolites can be new sources of such economic materials as oils, gums or tannins, new therapeutic agents and precursors of synthesis of complex chemical substances. Of the several hundred thousand plant species present on earth, only small proportion has been studied both chemically and biologically. The combination of both chemical and biological screening will provide important information about plant natural products (1).

Beni-Sueif governorate occupies a land area of approximately 10,954 km² in north part of Upper Egypt, with a total inhabitancy of 1,369.41 km². It boasts a population of over 2,315,512. Three phytogeographical

regions can be distinguished in Beni-Sueif; the desert on the western side of the Nile that is considered as extension of the African Sahara region, the desert of the eastern side of the Nile that extends with the official border of the governorate to Red sea and the fertile land on both sides of the Nile including canal banks distributed throughout the governorate. In our continued efforts for chemical and biological screening of wild plants growing in Beni-Sueif (2), antioxidant activity of extracts prepared from commonly occurring wild plants was tested.

2. Materials and Methods**2.1. Plant material**

The plant materials used in this study consisted of mature leaves, flowers and latex of different plants. The plants have been collected from the wild in Gerza village area, Beni-Sueif, Egypt, during flowering stage of each plant (2007). Voucher specimens were deposited in the Herbarium of Faculty of Pharmacy, Beni-Sueif University. The plant materials were rinsed with tap water and air dried in shade. Aqueous methanol extract (80%) of different plants was prepared by extracting the plant material twice. The extracts were stored in refrigerator at 4°C till use.

2.2. DPPH radical scavenging activity

The free radical scavenging activity (hydrogen donation) of plant extracts and *n*-propyl gallate was measured using 1,1-diphenyl-2-picryl-hydrazil (DPPH) radical. A solution of DPPH in methanol (0.08 mM) was prepared. Then, 1 mL of this solution was added to 0.3 mL of extracts or *n*-propyl gallate solutions at concentration of 500 µg/mL. The mixture was then shaken vigorously and allowed to stand at room temperature for 30 min, with the absorbance measured at 517 nm in a spectrophotometer against blank samples.

*Correspondence to: Dr. Sameh AbouZid, Department of Pharmacognosy, Faculty of Pharmacy, Beni-Sueif University, Beni-Sueif, Egypt;
e-mail: wssmd3000@yahoo.com

2.3. Superoxide anion scavenging activity

Superoxide radicals were generated in phenazine methosulphate (PMS)-nicotinamide adenine dinucleotide (NADH) systems by NADH oxidation and assayed by nitroblue tetrazolium (NBT) reduction. In this experiment, the superoxide radicals were generated in 3 mL of Tris-HCl buffer (16 mM, pH 8.0) containing 0.5 mL of NBT (300 μ M) solution, 0.5 mL NADH (936 μ M) solution, and 0.5 mL of plant extracts or *n*-propyl gallate solution at concentration of 500 μ g/mL. The reaction was started by adding 0.5 mL of PMS solution (120 μ M) to the mixtures. The reaction mixture was incubated at 25°C for 5 min, and the absorbance at 560 nm was measured against blank samples using a spectrophotometer.

2.4. Iron chelating activity

Plant extracts or EDTA solution (0.94 mL) at concentration of 500 μ g/mL was added to a solution of 0.02 mL FeCl₂ (2 mM). The reaction was initiated by the addition of 0.04 mL ferrozine (5 mM), and then the mixture was shaken vigorously and left standing at room temperature for 10 min. After equilibrium had been reached, absorbance of the solution was measured spectrophotometrically at 562 nm.

3. Results and Discussion

Free radicals are involved in a number of pathological conditions such as inflammatory diseases,

atherosclerosis, cerebral ischemia, AIDS, and cancer (3). The free radicals are induced in the human body due to environmental pollutants, chemicals, physical stress, radiations, *etc.* Catalase and hydroperoxidase enzymes are among the most important antioxidants produced by the immune system. Consumption of antioxidants or free radical scavengers is necessary to compensate depletion of antioxidants of the immune system.

There is an increasing interest in the use of medicinal plants as antioxidants. Silymarin and wheat germ oil are well-known plant extracts used as antioxidants in the pharmaceutical market. In the present study, plant extracts prepared from commonly occurring wild plants in Beni-Sueif governorate, have been screened for their *in vitro* antioxidant activity. The reported medicinal properties of these plants are described in Table 1.

The antioxidant activity of aqueous methanol (80%) extracts tested is listed in Table 2. In the DPPH radical scavenging assay, only extracts of leaves and flowers of *Tamarix nilotica* showed significant activity. In superoxide anion scavenging activity the same extracts and that of *Conyza dioscoridis* have shown significant activity. In iron chelating activity extracts of leaves and flowers of *T. nilotica* and that of *Calotropis procera* leaves have shown the highest activity. Extracts prepared from *T. nilotica* flowers have shown the highest antioxidant activities in the three used assays. Extracts of *T. nilotica* have been used in traditional Egyptian medicine as antiseptic agents. The polyphenolic and flavonoids of *T. nilotica* have been previously investigated (4-7). The methanol extract of

Table 1. Plants tested for their antioxidant activity in this study

Plant name	Family	Parts used	Reported medicinal values
<i>Tamarix nilotica</i>	Tamaricaceae	Leaves and flowers	Antiseptic in traditional Egyptian medicine (4)
<i>Ambrosia maritima</i>	Asteraceae	Leaves	Hepatoprotective (8), Molluscicidal (9)
<i>Zygophyllum coccenium</i>	Zygophyllaceae	Leaves	Anidiarrheal (10), Antidiabetic (11)
<i>Conyza dioscoridis</i>	Asteraceae	Leaves and flowers	–
<i>Chenopodium ambrosioides</i>	Chenopodiaceae	Leaves and flowers	Trypanocidal (12), Antileishmania (13)
<i>Calotropis procera</i>	Asclepiadaceae	Leaves	Protection against acetaminophen induced liver damage (14)
<i>Calotropis procera</i>		Latex	Contractions of gastrointestinal smooth muscle (15)

Table 2. Antioxidant activity of aqueous methanol (80%) extracts^a of different plants

Plant name (Plant part)	Type of assay		
	DPPH radical ^b	Superoxide anion ^b	Fe ³⁺ chelation ^c
<i>Tamarix nilotica</i> (leaves)	73.13 \pm 1.15	96.34 \pm 0.83	79.30 \pm 4.49
<i>Tamarix nilotica</i> (flowers)	89.34 \pm 0.82	92.82 \pm 3.88	79.56 \pm 2.94
<i>Ambrosia maritima</i> (leaves)	– ^d	21.87 \pm 1.52	60.24 \pm 1.81
<i>Zygophyllum coccenium</i> (leaves)	– ^d	33.79 \pm 2.19	22.63 \pm 3.16
<i>Conyza dioscoridis</i> (leaves and flowers)	28.01 \pm 0.69	93.98 \pm 3.17	40.99 \pm 3.19
<i>Chenopodium ambrosioides</i> (leaves and flowers)	– ^d	55.78 \pm 1.10	73.73 \pm 1.57
<i>Calotropis procera</i> (leaves)	– ^d	23.07 \pm 7.59	84.13 \pm 1.05
<i>Calotropis procera</i> (latex)	– ^d	– ^d	22.42 \pm 3.33
<i>n</i> -propyl gallate	90.31 \pm 0.24	91.2 \pm 0.21	
EDTA			98.46

^a Extract concentration used was 500 μ g/mL. ^b Percent of radical scavenging activity. ^c Percent inhibition. ^d Insignificant results (< 20%).

T. nilotica have shown higher DPPH radical scavenging activity ($51.5 \pm 8.14\%$) than that of silymarin ($40.4 \pm 2.05\%$). Further work is underway to characterize the active principles acting as antioxidants in these promising plant extracts.

Acknowledgments

Sameh AbouZid gratefully acknowledges receiving a postdoctoral fellowship from the International Center for Chemical and Biological Sciences (ICCBS) and the academy of sciences for the developing world (TWAS). Thanks are due to Dr. Emad Elshereef, Faculty of Science, Beni-Sueif University, Egypt for his kind assistance and advice in collection and identification of the plants.

References

- Hostettmann K. Strategy for the biological and chemical evaluation of plant extracts. *Pure App Chem* 1999; 70:1-9.
- Ezeldin S, Ibrahim I, Wahba H, Hassan M, Zaki M, AbouZid S. Phytochemical studies on plants growing in Beni-Sueif governorate, Egypt. Abstract Book of The 30th International Conference of Pharmaceutical Science Organized by The Egyptian Pharmaceutical Society. 1999; p. 43 – Egypt.
- Thomas CE, Kalyanaraman B. Oxygen radicals and the disease process. Harwood Academic Publishers, Amsterdam, Netherlands, 1997.
- Nawwar MAM, Buddrus J, Bauer H. Dimeric phenolic constituents from the roots of *Tamarix nilotica*. *Phytochemistry* 1982; 21:1755-1758.
- Nawwar MAM, Souleman AMA, Buddrus J, Linscheid M. Flavonoids of the flowers of *Tamarix nilotica*. *Phytochemistry* 1984; 23:2347-2349.
- Nawwar MAM, Souleman AMA, Buddrus J, Bauet H, Linscheid M. Phenolic constituents of the flowers of *Tamarix nilotica*: The structure of nilocitin, a new digalloylglucose. *Tetrahedron Lett* 1984; 25:49-52.
- El-Sisi HI, Nawwar MAM, Saleh NAM. Plant constituents of *Tamarix nilotica* leaves (Tamaricaceae). *Experientia* 1973; 29:1064-1065.
- Bastawy MA, Khater MR. Evaluation of the protective potential of *Ambrosia maritima* extract on acetaminophen-induced liver damage. *J Ethnopharmacol* 2001; 75:169-174.
- Belot J, Geerts S, Sarr S, Polderman AM. Field trials to control schistosome intermediate hosts by the plant molluscicide *Ambrosia maritima* L. in the Senegal River Basin. *Acta Tropi* 1993; 52:275-282.
- Atta AH, Mouneir SM. Antidiarrhoeal activity of some Egyptian medicinal plant extracts. *J Ethnopharmacol* 2004; 92:303-309.
- Mansour HA, Newairy AA, Yousef MI, Sheweita SA. Biochemical study on the effects of some Egyptian herbs in alloxan-induced diabetic rat. *Toxicol* 2002; 170:221-228.
- Kiuchi F, Itano Y, Uchiyama N, Honda G, Tsubouchi A, Nakajima-Shimada J, Aoki T. Monoterpene hydroperoxides with trypanocidal activity from *Chenopodium ambrosioides*. *J Nat Prod* 2002; 65:509-512.
- Monzotea L, M. Montalvoa A, Almanonnic S, Scullb R, Mirandab M, Abreub J. Activity of the essential oil from *Chenopodium ambrosioides* grown in Cuba against *Leishmania amazonensis*. *Chemotherapy* 2006; 52:130-136.
- Olaleye MT, Rocha BT. Acetaminophen-induced liver damage in mice: effects of some medicinal plants on the oxidative defense system. *Exp Toxicol Pathol* 2008; 59:319-327
- Kumar VL, Shivkar YM. *In vivo* and *in vitro* effect of latex of *Calotropis procera* on gastrointestinal smooth muscles. *J Ethnopharmacol* 2004; 93:377-379.

(Received November 15, 2008; Accepted November 23, 2008)

Original Article**The neuroprotective effect of antidepressant drug *via* inhibition of TIEG2-MAO B mediated cell death****Deyin Lu, Chandra Johnson, Shakevia Johnson, Shawna Tazik, Xiao-Ming Ou****Department of Psychiatry and Human Behavior, University of Mississippi Medical Center, Jackson, MS 39216, USA.*

ABSTRACT: Alcohol use disorders are common in the world. However, the development of novel drugs to prevent alcohol-induced brain damage is based upon an improved neurobiological understanding on the cellular changes that take place in the brain. We previously reported that ethanol exposure lowered cell proliferation and increased cell apoptosis in all cell types, but affects brain cell lines the most, while ethanol and the anti-depressant drug deprenyl, an monoamine oxidase B (MAO B) inhibitor, exposure in unison increases cell viability. Here we investigated the molecular mechanism of the neuroprotective effect of deprenyl (0.25 nM) on ethanol (75 mM)-induced harmful effect. Transforming growth factor-beta-inducible early gene 2 (TIEG2) is an activator for MAO B. MAO B levels increase has been shown to contribute to neuronal cell death. This study uses the neuronal cell line to address whether ethanol induced cell death is through the activation of TIEG2-MAO B apoptotic pathway, and whether deprenyl protects cells from the effects of alcohol through the inhibition of this pathway. We have found that ethanol exposure increases the levels of mRNA and protein/catalytic activity for both TIEG2 and MAO B, while ethanol and deprenyl exposure in unison reduce the expression of both TIEG2 and MAO B, however it increases cell viability. Additionally, TIEG2-overexpressed cells display more cellular death-induced by ethanol than control cells. In summary, this study demonstrates the role of TIEG2 in ethanol induced cell death. The inhibition of the TIEG2-MAO B pathway may be one of the mechanisms for the neuroprotective effect of deprenyl.

Keywords: Alcohol, Neuroprotection, Transforming growth factor-beta-inducible early gene 2, Monoamine oxidase B, Cell viability

*Correspondence to: Dr. Xiao-Ming Ou, Division of Neurobiology & Behavioral Research G-109, Department of Psychiatry and Human Behavior, University of Mississippi Medical Center, 2500 N. State Street, Jackson, MS 39216, USA;
e-mail: xou@psychiatry.umsmed.edu

1. Introduction

Alcohol use disorders are common around the world and also have a high correlation between alcohol use disorders and other psychiatric problems, such as major depression (1). Although short-term alcohol drinking causes euphoric and stress-relieving effects, numerous clinical and experimental studies have shown that alcohol use is a major risk factor for neurobehavioural diseases, inflammation disorders and enhanced susceptibility to bacterial infection (2-4). In particular, it affects the brain and alters its normal function (5). This includes altering the effects of neurotransmitters, suppressing nerve signals and causing cell death (6). In rodents, ethanol exposure during development significantly reduces the size of the brain as well as brain/body weight ratios (7). There are many adverse physical effects from long-term exposure to alcohol including the increased activity in the liver that causes cell death and chronic hepatic diseases (alcoholic fatty liver, alcoholic hepatitis and cirrhosis, etc) (7,8) and an increase in the number of apoptotic cells in various brain areas (9).

Monoamine oxidase (MAO) metabolizes biogenic and dietary amines in the central nervous system and peripheral tissues, including monoamine neurotransmitters serotonin, norepinephrine, dopamine, and phenylethylamine. MAO plays important roles in several psychiatric and neurological disorders (10). MAO exists in two forms, MAO A and MAO B. Their catalytic activity generates H₂O₂ and nitrogen species, which are toxic products and may cause oxidative damage to mtDNA and have potential implications for apoptosis, aging, and neurodegenerative processes.

Deprenyl (selegiline), an irreversible inhibitor of monoamine oxidase B (MAO B), was synthesized as an antidepressant and used to treat Parkinson's disease (11). Because MAO degrades serotonin and produces reactive oxygen that may cause cell death, an MAO inhibitor prevents cell apoptosis (12,13). Deprenyl or related compounds may be neuroprotective in general through the inhibition of "death" signal transduction-

mediated by MAO, induced by endogenous and environmental factors (11). Deprenyl in low concentrations that induce MAO B inhibition potentially inhibits serum withdrawal induced apoptosis in tissue cultures of neuro-ectodermal origin (14). This report is consistent with our previous studies (15) that ethanol can induce apoptosis in neuronal cells, and deprenyl in a low concentration can protect cells from the harmful effects of ethanol.

Transforming growth factor-beta-inducible early gene 2 (TIEG2, also called KLF11--Kruppel-Like Factor 11) is a human transforming growth factor-beta-(TGF- β) inducible early gene. It is a recently identified human TGF- β -inducible zinc finger protein belonging to Sp1-like family of transcription factors. TIEG2 protein is a negative regulator of cell growth and induces apoptosis (16-18) by binding to GC-rich sequences (19) located in the promoter region of several genes including MAO B promoter. Ethanol has been shown to potentiate TGF- β 1-mediated growth inhibition in the rat neuroblastoma cells (20), and ethanol exposure increases TGF- β 1 signal (21) that may increase TIEG2 protein level and lead to apoptotic death of cells (22). Our previous data have shown that TIEG2 activates MAO B gene expression (23). This study investigates the neuroprotective effect of antidepressant drug (deprenyl) on ethanol-induced apoptosis possibly mediated by TIEG2 and MAO B.

2. Materials and Methods

2.1. Cell lines, DNA plasmids and reagents

SH-SY5Y, a human neuroblastoma cell line, was purchased from The American Type Culture Collection (ATCC). SH-SY5Y was cultured in RPMI1640 supplemented with 10% fetal bovine serum and antibiotics. TIEG2-expression vector was a gift from Dr. Raul Urrutia, Mayo Clinic. TIEG2 coding sequence was cloned into pcDNA3.1 His A expression vector. MAO B inhibitor, selegiline (deprenyl), was purchased from Sigma-Aldrich USA. The antibodies used in this study were purchased from Santa Cruz Biotechnology, except that anti-TIEG2 antibody was from BD Transduction Laboratory.

2.2. TIEG2-stably transfected cell line

In generating the TIEG2-stable cell line, SH-SY5Y were plated at a density of 5×10^6 cells in a 10-cm dish. The next day the TIEG2 expression vector or pcDNA 3.1 was transfected into cells with a Superfect transfection reagent (Qiagen Inc). After 24 h, cells were treated with Geneticin (G418; 600 μ g/mL). Resistant clones isolated into separate dishes after 6 days and cultured under continuous G418 selection (13).

2.3. Cell culture and treatments with ethanol and deprenyl

Before treatments, SH-SY5Y cells were seeded on 10-cm dishes or 6-well plates. After overnight culture in medium, the medium was replaced with new medium containing 75 mM of ethanol with or without 0.25 nM of deprenyl for three days. As ethanol is volatile, a closed chamber system was utilized to stabilize the ethanol concentration in the culture medium (24,25). With this system, ethanol concentrations are maintained at steady ethanol levels (more than ~90% of the original concentration) for 3 days in medium. Briefly, cell culture dishes or 6-well plate containing SH-SY5Y cells were placed on a rack inside a plastic container that could be tightly sealed. A separate sealed container was used for each ethanol concentration. The bottom of each container was a reservoir that was filled with 200 mL of an aqueous solution with the same ethanol concentration that was present in the culture medium. A nonethanol control had a bath of water only. The underlying principle of this method is that the alcohol in the bath evaporates into the air inside the sealed container establishing a stable vapor pressure so that there is no net loss of ethanol from the culture medium. Before sealing the containers, a small amount (60 cc) of CO₂ was injected into each container. The concentration of CO₂ in the chamber was routinely tested and was determined to be stable at 5%. The containers were sealed and maintained in an incubator at 37°C for up to 3 days as needed (25).

The ethanol concentration we used (75 mM for examining the effect of deprenyl) was within the standard range of *in vitro* study (26). When a heavy drinker's ethanol concentration in blood reaches ~50-100 mM, he probably shows slurred speech and unsteadiness (27). Therefore the ethanol concentration for this study is around the physiological effect of ethanol in alcoholics.

2.4. Real-time PCR (RT-PCR)

Total RNAs were extracted with Trizol from cultured cells. Reverse transcription was carried out with SuperScript first strand synthesis system for RT-PCR (Invitrogen Inc) following the manufacturer's instruction. Specific primers for the human MAO B and TIEG2 were designed as follows:

MAO B Sense,
5'-GACCATGTGGGAGGCAGGACTTAC-3'
Antisense,
5'-CGCCACAAATTTCTCTCTCTG-3'
TIEG2 Sense,
5'-CCTGTTGCGGATAAGACCCCTCAC-3'
Antisense,
5'-AAAGCCGCAATCTGGAGTCTGGA-3'

The mRNA quantitative analyses for each group were

performed by Real-Time PCR using a Bio-Rad iCycler system. The real-time PCR was performed with a SYBR supermix kit (Bio-Rad). The data were analyzed by the software from Bio-Rad as described previously (23).

2.5. MAO B catalytic activity assay

SH-SY5Y was grown to confluence, harvested, and washed with phosphate-buffered saline. One hundred micrograms of total proteins were incubated with 10 μ M 14 C-labeled PEA (Amersham Biosciences) in the assay buffer (50 mM sodium phosphate buffer, pH 7.4) at 37°C for 20 min and terminated by the addition of 100 μ L of 6 N HCl. The reaction products were then extracted with ethyl acetate/toluene (1:1) and centrifuged for 7 min. The organic phase containing the reaction product was extracted, and its radioactivity was obtained by liquid scintillation spectroscopy (28).

2.6. Western blot

Cells were cultured in medium with ethanol (75 mM) for 3 days, washed by PBS (pH 7.4), and sonicated in 500 μ L of RIPA lysis buffer (10 mM Tris-HCl, pH 7.4/160 mM NaCl/1% Triton/1% Na dextrylate/0.1% SDS/1 mM EDTA/1 mM EGTA) supplemented with protease inhibitors (Sigma). The samples were then freeze thawed and centrifuged for 2 min at 12,500 rpm. The supernatant was then kept and transferred to a new tube. Thirty micrograms (for TIEG2) of total proteins were separated by 10.5% SDS-polyacrylamide gel electrophoresis and transferred to PVDF membranes. After the transfer, membranes were blocked at room temperature for 2 h with 5% nonfat dry milk in TBS (10 mM Tris-HCl, pH 7.5, 150 mM NaCl). The membranes were then incubated with mouse anti-TIEG2 antibody (1:500) or mouse anti-actin antibody (1:1,000) overnight at 4°C. After incubation with respective secondary antibody at room temperature for 2 h, the bands were visualized by horseradish peroxidase (HRP) reaction using SuperSignal West Pico Chemiluminescent Substrate (PIERCE).

2.7. MTT assay for proliferation rate/cell viability evaluation

Cell viability and proliferation was measured by tetrazolium salt (MTT) (13,15). The medium in excess of 2 mL (6-well plates) was removed and 40 μ L of MTT dye (5 mg/mL) in sterile PBS was added to 360 μ L of medium or PBS depending on cell confluence. Plates were incubated for 4 to 5 hours, during which time the mitochondria in living cells converted the soluble yellow dye (MTT) into an insoluble purple formazan crystal. Cells and dye were then solubilized by the addition of 800-1,000 μ L of DMSO to the 6-well plates. Optical density of each well at 572 nm was determined using the NanoDrop Spectrophotometer.

2.8. Statistical analysis

All values are presented as means \pm SD. A one-way ANOVA followed by a post hoc Bonferroni's *t*-test was employed when three or more groups were to be compared. A paired *t*-test was performed for the statistical analysis of two groups. A *P* value less than 0.05 was considered to be statistically significant.

3. Results

3.1. Ethanol increases the MAO B mRNA level and enzymatic activity, but deprenyl reverses the effect of ethanol

SH-SY5Y cells were treated with 75 mM ethanol for three days. Then the cellular mRNA and MAO B activity were determined by real time-PCR and traditional 14 C method assay. The ethanol concentration we used (75 mM) was clinically relevant, because the ethanol at 50-100 mM reflects blood ethanol levels in chronic alcoholics (29,30). Therefore, the ethanol concentration in our study was within the levels that results in physiological effects observed in alcoholics.

The results showed that ethanol induced cellular MAO B mRNA level increased about 3.5 times more than the control; this increase could be reduced by 0.25 nM deprenyl treatment (Figure 1A, lanes 2 vs. 1 and 3 vs. 1). At the same time, we found MAO B activity in ethanol treated cells increased 1.8 fold more than control cells (Figure 1B, lanes 2 vs. 1). However, deprenyl significantly decreased the MAO B catalytic activity (Figure 1B, lanes 3 vs. 2).

3.2. Ethanol increases the MAO B mRNA and protein levels, but deprenyl reverses the effect of ethanol

To test the possibility that TIEG2 may take part in the ethanol-induced cell death, TIEG2 mRNA levels and protein levels were detected with RT-PCR and western blot. The results show that ethanol could induce TIEG2 mRNA expression 4-fold more than that of the control cells (Figure 2A lanes 2 vs. 1), deprenyl plus ethanol could just increase 2.8-fold (Figure 2A, lanes 3 vs. 1). Similarly, TIEG2 protein expression level was increased around 3.5 times, however deprenyl could inhibit TIEG2 expression (Figure 2B, a and b, lanes 2 and 3 vs. 1).

3.3. TIEG2 enhances, but MAO B inhibitor (deprenyl) protects, cell death induced by ethanol

Ethanol has been found to increase the MAO B gene expression and catalytic activity in the human glioma 1242-MG cells (16). With our previous experiment, 75 mM ethanol treatment in conjunction with 0.25 nM deprenyl provided the most protection against apoptotic activity for brain cells SH-SY5Y and U-118 MG (31).

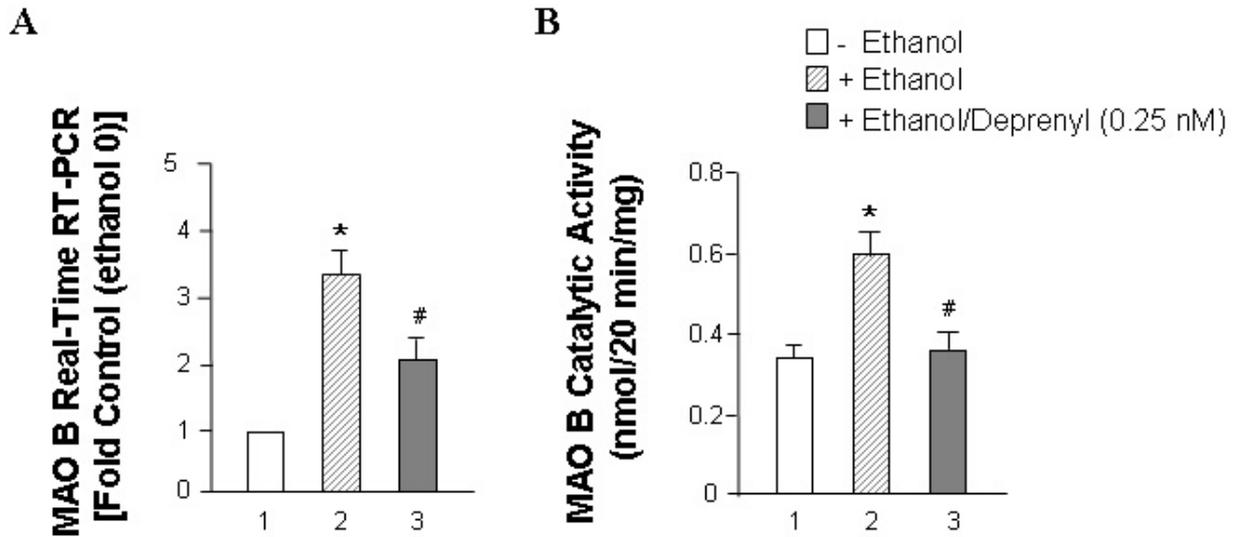


Figure 1. Effects of ethanol on MAO B mRNA level and catalytic activity. SHSY5Y cells were treated with or without 75 mM ethanol or with ethanol plus 0.25 nM deprenyl for three days. **(A)** Cellular mRNA was extracted and quantitative real-time RT-PCR was performed. **(B)** MAO B enzyme activity was determined by enzymatic activity assay. Data represent the mean \pm S.D. of three independent experiments. Controls were untreated cells (0 mM of ethanol) which were taken as 1. * $P < 0.01$ versus control cells and # < 0.05 versus cells treated with ethanol alone (one-way ANOVA followed by a post hoc Bonferroni's *t*-test).

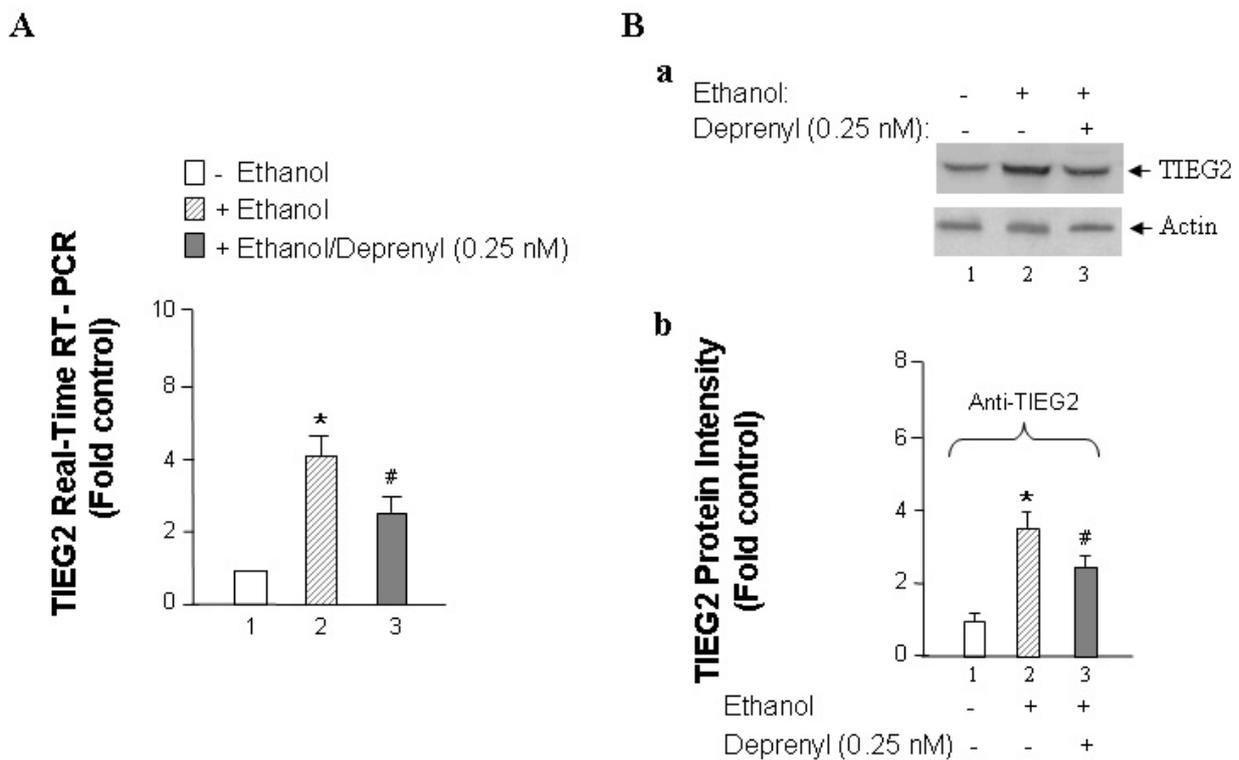


Figure 2. Effects of ethanol on TIEG2 mRNA and protein levels. SHSY5Y cells were treated with or without 75 mM ethanol or with ethanol plus 0.25 nM deprenyl for three days. **(A)** Cellular TIEG2 mRNA level was detected with quantitative real-time RT-PCR, and **(B)** TIEG2 protein was examined by western blot. **(a)** TIEG2 protein levels in cells treated with or without ethanol or with ethanol plus deprenyl were shown by western blot, and actin was used as the internal control. **(b)** The optical density analysis with western blot shows TIEG2 has about 3.5 fold expression in ethanol treated cells, whereas, deprenyl inhibited the TIEG2 expression. Data represent the mean \pm S.D. of three independent experiments. * $P < 0.01$ versus control cells without ethanol treatment and # < 0.05 versus cells treated with ethanol (one-way ANOVA followed by a post hoc Bonferroni's *t*-test).

Here, we use pcDNA and TIEG2 stably transfected SH-SY5Y cell lines (Figure 3Aa) to investigate the role of TIEG2 in ethanol-induced cell death and the neuroprotective effect of deprenyl. We have previously shown that TIEG2 is a transcriptional activator for MAO B (23). As shown in Figure 3Ab, the MAO B catalytic

activity is increased by ~2-fold in TIEG2-overexpressed cells as compared to that in pcDNA3.1-transfected cells (Figure 3Ab, lanes 2 vs. 1).

Next, cells were exposed to 75 mM ethanol in conjunction with 0.25 nM deprenyl for three days, and the cell viability (in survival rate) was observed in

SH-SY5Y cells compared to that in cells treated with 75 mM ethanol alone. The results show that TIEG2-overexpression could induce more cell death with the presence of 75 mM ethanol than that in the control group which was stably transfected with empty pcDNA3.1 vector (Figure 3B, lanes 7 vs. 5). However, deprenyl could protect cells from ethanol's harmful effect (Figure 3B, lanes 6 vs. 5 and 8 vs. 7).

Previously, we have shown that 0.25 nM of deprenyl produced the best neuroprotective effect on SH-SY5Y cells (15). In order to examine whether 0.25 nM deprenyl is also the most appropriate dosage in this study using TIEG2-overexpressed cells, the different concentrations (0, 0.125, 0.25, 0.5, and 1 nM) of deprenyl were used to test the inhibitory effects on MAO B catalytic activity and cell death. As shown in Figure 3Ca, the ethanol treatment in conjunction with deprenyl for three days

exhibited the inhibition on MAO B catalytic activity in a concentration dependent manner. Furthermore, MTT assay was performed (Figure 3Cb) to determine the effects of different concentrations (0, 0.125, 0.25, 0.5, and 1 nM) of deprenyl on cell viability in TIEG2-overexpressed cells. The result showed that the ethanol treatment in conjunction with 0.25 nM deprenyl for three days increased the cell survival rate by 175% as compared to that of control cells (Figure 3Cb, lanes 3 vs. 1), suggesting that 0.25 nM of deprenyl has the most protection against apoptotic activity.

4. Discussion

An understanding of the molecular mechanisms of cellular apoptosis toward excessive alcohol consumption is crucial for the development of new

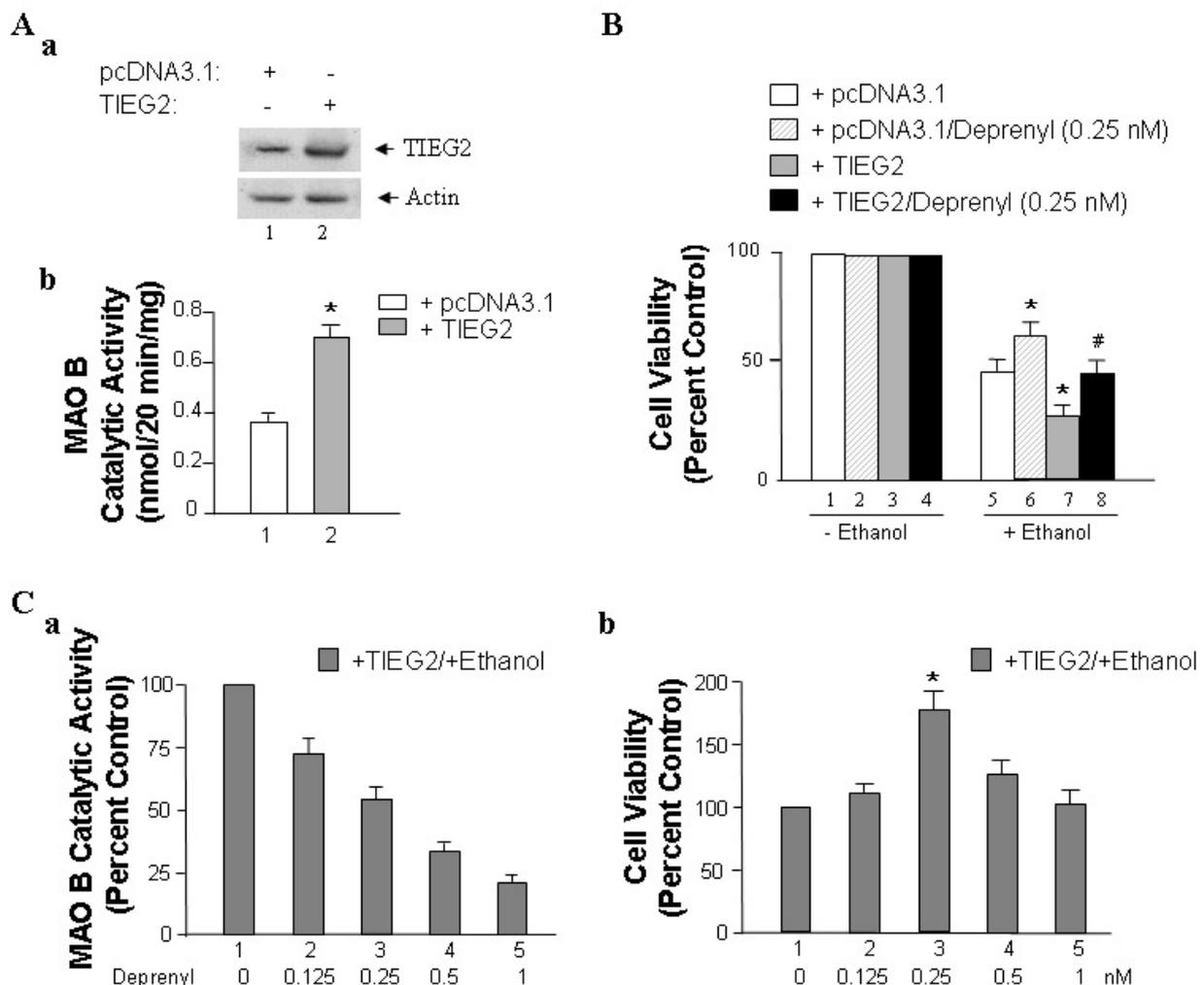


Figure 3. The effect of TIEG2 on ethanol-induced cell death. (A) Compare the expression of TIEG2 and MAO B catalytic activity in TIEG2/pcDNA stably transfected SH-SY5Y cell lines. (a) Western Blot analysis of the expression of TIEG2 in TIEG2/pcDNA overexpressed cell lines. (b) MAO B catalytic activity is about 2 times higher in TIEG2 stably transfected cells than pcDNA stably transfected cells. Values are expressed as means \pm S.D. of at least three independent experiments. * $P < 0.02$ (paired *t*-test). (B) Effects of ethanol, ethanol plus deprenyl and TIEG2 on cell survival rates. pcDNA3.1 stably transfected cells or TIEG2 stably transfected cells were treated with 75 mM ethanol with/without 0.25 nM deprenyl for three days. Then the cell viability was determined by MTT assay. (C) Effects of different dosage of deprenyl on MAO B catalytic activity and the protection of ethanol induced cell death in TIEG2-stably transfected SH-SY5Y cell line. Cells were treated with or without 75 mM ethanol in the conjunction with 0.125, 0.25, 0.5 or 1 nM of deprenyl as indicated in the figure for three days. Then (a) the MAO B catalytic activity or (b) cell viability (MTT assay) was determined. All data are presented as the mean \pm S.D. of at least three independent experiments. Controls were untreated cells (0 mM of ethanol in B and 0 nM of deprenyl in C) which were taken as 100%. * $P < 0.05$ versus control cells and # < 0.05 versus cells-stably expressing TIEG (one-way ANOVA followed by a post hoc Bonferroni's *t*-test).

treatments for alcoholism. In this study, we examine the TIEG2-MAO B role in the ethanol induced apoptosis, and the neuroprotective effect of deprenyl *via* inhibition of TIEG2-MAO B mediated cell death with SH-SY5Y cell line.

The MAO B gene is located on the Xp11.2-11.4 chromosome and consists of 15 exons with identical exon-intron organization (32), and its activity increases progressively in the brain throughout adult life (33,34). An aberrant increase of MAO B activity has been implicated in several psychiatric and neurodegenerative disorders (35,36). Thus one predicted mechanism for cell death is an abnormal increase in monoamine oxidase (37). Previously, the physiologically relevant concentration of ethanol has been found to increase the MAO gene expression and catalytic activity in the human glioma 1242-MG cells (31). The increased activity of MAO may thereby increase production of hydrogen peroxide (H₂O₂, a major source for oxidative stress) and cause apoptosis (38). The SH-SY5Y, a human neuroblastoma cell line, treated with 75 mM ethanol, could increase the expression of mRNA and protein, in particular, MAO B catalytic activity also increased. Previously, we showed that the level of Caspase 3, an apoptotic marker protein, was increased significantly by ethanol treatment, suggesting that ethanol-induced cell death is mediated at least partially by apoptotic pathway (15).

Transforming growth factor-beta-inducible early gene 2 (TIEG2) is an activator for MAO B through Sp1 overlapping sites (GC-rich sequence) located at the promoter region of MAO B. Sp1-like protein plays key roles in the regulation of MAO B gene expression (23). It has been reported that TIEG2 induces apoptosis in murine OLI-neu cells (39). With the ethanol treated SH-SY5Y system, we were able to show that the mRNA and protein levels for TIEG2 were increased significantly along with the increase in MAO B activity. Using TIEG2-overexpressed stable cell line, we further demonstrated that TIEG2 could increase the MAO B catalytic activity, and also enhance the cellular apoptosis triggered by ethanol, whereas, deprenyl, an MAO B inhibitor, could protect cell death induced by ethanol, because ethanol and deprenyl exposure in unison reduced the expression of both TIEG2 and MAO B.

Deprenyl is an irreversible inhibitor of MAO B which is an antidepressant drug, and is now also used in the treatment of Parkinson's disease. Deprenyl in much lower concentrations needed to induce MAO B inhibition (less than ~1 nM) potently inhibits serum withdrawal (40) and nitric oxide (41) induced apoptosis. However, in high concentration, deprenyl induces apoptosis in cell cultures (14). Our findings suggest that 0.25 nM deprenyl and ethanol exposure in unison for three days is able to inhibit MAO B catalytic activity and produce the best neuroprotective effect comparing to other concentrations (0.125, 0.5, and 1 nM) of deprenyl. This may be due to the higher concentration (more than 0.25 nM) of

deprenyl used to start to induce apoptosis in cell culture (14).

In summary, TIEG2-MAO B-mediated apoptotic pathway may contribute to ethanol induced neurotoxicity. The inhibition of this apoptotic signaling pathway may be one of the mechanisms for the neuroprotective effect of deprenyl.

Acknowledgements

This study was supported by Public Health Service Grants P20 RR17701 and a NARSAD Young Investigator Award. The contribution by Dr. Raul Urrutia for providing TIEG2-pcDNA3.1 expression vector is also acknowledged. Shakevia Johnson and Shawna Tazik were supported by the Neuroscience Summer Scholars Program, Department of Psychiatry, University of Mississippi Medical Center.

References

1. Hasin DS, Goodwin RD, Stinson FS, Grant BF. Epidemiology of major depressive disorder: results from the National Epidemiologic Survey on Alcoholism and Related Conditions. *Arch Gen Psychiatry* 2005; 62:1097-1106.
2. Bruno A. Cerebrovascular complications of alcohol and sympathomimetic drug abuse. *Curr Neurol Neurosci Rep* 2003; 3:40-45.
3. Frank J, Witte K, SchrodL W, Schutt C. Chronic alcoholism causes deleterious conditioning of innate immunity. *Alcohol Alcohol (Oxford, Oxfordshire)* 2004; 39:386-392.
4. GullO L, Migliori M, Brunetti MA, Manca M. Alcoholic pancreatitis: new insights into an old disease. *Curr Gastroenterol Rep* 2005; 7:96-100.
5. Herrera DG, Yague AG, Johnsen-Soriano S, Bosch-Morell F, Collado-Morente L, Muriach M, Romero FJ, Garcia-Verdugo JM. Selective impairment of hippocampal neurogenesis by chronic alcoholism: protective effects of an antioxidant. *Proc Natl Acad Sci U S A* 2003; 100:7919-7924.
6. Ikonomidou C, Bosch F, Miksa M, Bittigau P, Vockler J, Dikranian K, Tenkova TI, StefovskA V, Turski L, Olney JW. Blockade of NMDA receptors and apoptotic neurodegeneration in the developing brain. *Science* 1999; 283:70-74.
7. Jiang Q, Hu Y, Wu P, Cheng X, Li M, Yu D, Deng J. Prenatal alcohol exposure and the neuroapoptosis with long-term effect in visual cortex of mice. *Alcohol Alcohol (Oxford, Oxfordshire)* 2007; 42:285-290.
8. Tsukamoto H, Lu SC. Current concepts in the pathogenesis of alcoholic liver injury. *Faseb J* 2001; 15:1335-1349.
9. Pignataro L, Miller AN, Ma L, Midha S, Protiva P, Herrera DG, Harrison NL. Alcohol regulates gene expression in neurons *via* activation of heat shock factor 1. *J Neurosci* 2007; 27:12957-12966.
10. Shih JC, Chen K, Ridd MJ. Monoamine oxidase: from genes to behavior. *Annu Rev Neurosci* 1999; 22:197-217.
11. Maruyama W, Naoi M. Neuroprotection by (-)-deprenyl and related compounds. *Mech Ageing Dev* 1999; 111:189-200.

12. Tatton WG, Ju WY, Holland DP, Tai C, Kwan M. (-)-Deprenyl reduces PC12 cell apoptosis by inducing new protein synthesis. *J Neurochem* 1994; 63:1572-1575.
13. Ou XM, Chen K, Shih JC. Monoamine oxidase A and repressor R1 are involved in apoptotic signaling pathway. *Proc Natl Acad Sci U S A* 2006; 103:10923-10928.
14. Magyar K, Szende B. (-)-Deprenyl, a selective MAO-B inhibitor, with apoptotic and anti-apoptotic properties. *Neurotoxicology* 2004; 25:233-242.
15. Johnson S, Williams AN, Johnson C, Ou XM. The effects of antidepressant drug on ethanol-induced cell death. *Drug Discov Ther* 2007; 1:130-135.
16. Tachibana I, Imoto M, Adjei PN, Gores GJ, Subramaniam M, Spelsberg TC, Urrutia R. Overexpression of the TGFbeta-regulated zinc finger encoding gene, TIEG, induces apoptosis in pancreatic epithelial cells. *J Clin Invest* 1997; 99:2365-2374.
17. Cook T, Gebelein B, Mesa K, Mladek A, Urrutia R. Molecular cloning and characterization of TIEG2 reveals a new subfamily of transforming growth factor-beta-inducible Sp1-like zinc finger-encoding genes involved in the regulation of cell growth. *J Biol Chem* 1998; 273:25929-25936.
18. Zhang JS, Moncrieffe MC, Kaczynski J, Ellenrieder V, Prendergast FG, Urrutia R. A conserved alpha-helical motif mediates the interaction of Sp1-like transcriptional repressors with the corepressor mSin3A. *Mol Cell Biol* 2001; 21:5041-5049.
19. Cook T, Gebelein B, Belal M, Mesa K, Urrutia R. Three conserved transcriptional repressor domains are a defining feature of the TIEG subfamily of Sp1-like zinc finger proteins. *J Biol Chem* 1999; 274:29500-29504.
20. Luo J, Miller MW. Transforming growth factor beta1-regulated cell proliferation and expression of neural cell adhesion molecule in B104 neuroblastoma cells: differential effects of ethanol. *J Neurochem* 1999; 72:2286-2293.
21. Chen CP, Kuhn P, Chaturvedi K, Boyadjieva N, Sarkar DK. Ethanol induces apoptotic death of developing beta-endorphin neurons *via* suppression of cyclic adenosine monophosphate production and activation of transforming growth factor-beta1-linked apoptotic signaling. *Mol Pharmacol* 2006; 69:706-717.
22. Kuhn P, Sarkar DK. Ethanol induces apoptotic death of beta-endorphin neurons in the rat hypothalamus by a TGF-beta 1-dependent mechanism. *Alcohol Clin Exp Res* 2008; 32:706-714.
23. Ou XM, Chen K, Shih JC. Dual functions of transcription factors, transforming growth factor-beta-inducible early gene (TIEG)2 and Sp3, are mediated by CACCC element and Sp1 sites of human monoamine oxidase (MAO) B gene. *J Biol Chem* 2004; 279:21021-21028.
24. Adickes ED, Mollner TJ, Lockwood SK. Closed chamber system for delivery of ethanol to cell cultures. *Alcohol Alcohol (Oxford, Oxfordshire)* 1988; 23:377-381.
25. Luo J, Miller MW. Differential sensitivity of human neuroblastoma cell lines to ethanol: correlations with their proliferative responses to mitogenic growth factors and expression of growth factor receptors. *Alcohol Clin Exp Res* 1997; 21:1186-1194.
26. Ku BM, Joo Y, Mun J, Roh GS, Kang SS, Cho GJ, Choi WS, Kim HJ. Heme oxygenase protects hippocampal neurons from ethanol-induced neurotoxicity. *Neurosci Lett* 2006; 405:168-171.
27. Paton A. Alcohol in the body. *BMJ* 2005; 330:85-87.
28. Geha RM, Rebrin I, Chen K, Shih JC. Substrate and inhibitor specificities for human monoamine oxidase A and B are influenced by a single amino acid. *J Biol Chem* 2001; 276:9877-9882.
29. Yao Z, Zhang J, Dai J, Keller ET. Ethanol activates NFkappaB DNA binding and p56lck protein tyrosine kinase in human osteoblast-like cells. *Bone* 2001; 28:167-173.
30. Henriksen JH, Gronbaek M, Moller S, Bendtsen F, Becker U. Carbohydrate deficient transferrin (CDT) in alcoholic cirrhosis: a kinetic study. *J Hepatol* 1997; 26:287-292.
31. Ekblom J, Zhu QS, Chen K, Shih JC. Monoamine oxidase gene transcription in human cell lines: treatment with psychoactive drugs and ethanol. *J Neural Transm* 1996; 103:681-692.
32. Lan NC, Heinzmann C, Gal A, Klisak I, Orth U, Lai E, Grimsby J, Sparkes RS, Mohandas T, Shih JC. Human monoamine oxidase A and B genes map to Xp 11.23 and are deleted in a patient with Norrie disease. *Genomics* 1989; 4:552-559.
33. Fowler CJ, Wiberg A, Orelund L, Marcusson J, Winblad B. The effect of age on the activity and molecular properties of human brain monoamine oxidase. *J Neural Transm* 1980; 49:1-20.
34. Saura J, Luque JM, Cesura AM, Da Prada M, Chan-Palay V, Huber G, Loffler J, Richards JG. Increased monoamine oxidase B activity in plaque-associated astrocytes of Alzheimer brains revealed by quantitative enzyme radioautography. *Neuroscience* 1994; 62:15-30.
35. Schneider G, Oepen H, von Wedel HR. Monoamine oxidase activity in brain regions and organs of patients with Parkinson's disease and Huntington's disease and serum MAO activity of patients with Huntington's disease as compared with neurologically healthy individuals (author's transl). *Arch Psychiatr Nervenkr* 1981; 230:5-15.
36. Mann JJ, Kaplan RD, Bird ED. Elevated postmortem monoamine oxidase B activity in the caudate nucleus in Huntington's disease compared to schizophrenics and controls. *J Neural Transm* 1986; 65:277-283.
37. De Zutter GS, Davis RJ. Pro-apoptotic gene expression mediated by the p38 mitogen-activated protein kinase signal transduction pathway. *Proc Natl Acad Sci U S A* 2001; 98:6168-6173.
38. Nagatsu T, Sawada M. Molecular mechanism of the relation of monoamine oxidase B and its inhibitors to Parkinson's disease: possible implications of glial cells. *J Neural Transm* 2006; 71:53-65.
39. Wang Z, Spittau B, Behrendt M, Peters B, Krieglstein K. Human TIEG2/KLF11 induces oligodendroglial cell death by downregulation of Bcl-X(L) expression. *J Neural Transm* 2007; 114:867-875.
40. Tatton WG, Chalmers-Redman RM, Ju WJ, Mammen M, Carlile GW, Pong AW, Tatton NA. Propargylamines induce antiapoptotic new protein synthesis in serum- and nerve growth factor (NGF)-withdrawn, NGF-differentiated PC-12 cells. *J Pharmacol Exp Ther* 2002; 301:753-764.
41. Hara MR, Thomas B, Cascio MB, Bae BI, Hester LD, Dawson VL, Dawson TM, Sawa A, Snyder SH. Neuroprotection by pharmacologic blockade of the GAPDH death cascade. *Proc Natl Acad Sci U S A* 2006; 103:3887-3889.

(Received August 29, 2008; Revised October 15, 2008; Accepted October 19, 2008)

Original Article

Regulation of the nitric oxide synthesis pathway and cytokine balance contributes to the healing action of *Myristica malabarica* against indomethacin-induced gastric ulceration in miceBiswanath Maity¹, Debashish Banerjee¹, Sandip K. Bandyopadhyay¹, Subrata Chattopadhyay^{2,*}¹ Department of Biochemistry, Dr. B.C. Roy Post Graduate Institute of Basic medical Sciences & IPGME&R, 244B, Acharya Jagadish Chandra Bose Road, Kolkata, India;² Bio-Organic Division, Bhabha Atomic Research Centre, Mumbai, India.

ABSTRACT: The role of the arginine-metabolism in the healing action of the methanol extract of *Myristica malabarica* (rampatri) (RM) and omeprazole (Omez) against indomethacin-induced stomach ulceration in mouse was investigated. Indomethacin (18 mg/kg) was found to induce maximum stomach ulceration in Swiss albino mice on the 3rd day of its administration, which was associated with reduced arginase activity (38.5%, $p < 0.05$), eNOS expression, along with increased iNOS expression, total NOS activity (5.37 fold, $p < 0.001$), NO generation (55.1%, $p < 0.01$), and ratio of pro-/anti-inflammatory cytokines. Besides providing comparable healing as Omez (3 mg/kg \times 3 d), RM (40 mg/kg \times 3 d, p.o.) shifted the iNOS/NO axis to the arginase/polyamine axis as revealed from the increased arginase activity (59.5%, $p < 0.01$), eNOS expression, and reduced iNOS expression, total NOS activity (73%, $p < 0.001$), and NO level (49.8%, $p < 0.01$). These could be attributed to a favourable anti/pro inflammatory cytokines ratio, generated by RM. The healing by Omez was however, not significantly associated with those parameters.

Keywords: Arginase, Cytokine balance, Gastric ulcer healing, Indomethacin, NOS

1. Introduction

Gastric ulcer is a complex pluricausal disease and is known to develop due to loss of balance between aggressive and protective factors (1). The non-steroidal anti-inflammatory drugs (NSAIDs) are known to cause gastric ulceration and delay ulcer healing, which is

generally explained in terms of cyclooxygenase (COX) inhibition, reduced prostaglandin (PG) synthesis, and the impaired PG-mediated angiogenesis. However, the complete mechanism underlying this effect is not completely understood. Various other factors, especially the nitrogen-metabolizing enzymes are also key contributors in host immune defence mechanisms and wound healing (2,3). In acute inflammatory responses, such as wound healing, heat stroke and glomerulonephritis, arginase has been implicated as an important regulator of diverse pathways including generation of polyamines and the cytostatic free radical molecule, nitric oxide (NO) (4). Arginine pathway plays a vital role in wound healing since L-arginine becomes an essential amino acid after wounding with almost undetectable levels in the wound milieu (5). Studies have shown that arginine itself has advantageous effects on cutaneous healing by enhancing cell proliferation and collagen synthesis as well as breaking strength (6). Further, nitric oxide (NO), produced from arginine also plays an important role in inflammatory processes, being a mediator of macrophage function (4,6,7). The temporal switch of arginine as a substrate for the inducible nitric oxide synthase (iNOS)/NO axis to arginase/polyamine axis is subject to regulation by inflammatory cytokines. However, little is known on the interplay of cytokines and the NO synthesis pathway during indomethacin-induced gastric ulceration. After trauma, the Th1/Th2 imbalance with Th2 predominance is reflected by a predominance of the arginase inducing cytokines such as IL-4, IL-10, and TGF β (8).

Very recently, we have documented impressive healing activity of the fruit rinds of the plant, *Myristica malabarica* (Myristicaceae), popularly known as rampatri (9). It was found that oral administration of the methanol extract of rampatri (designated as RM) at a dose of 40 mg/kg for three days could effectively heal the indomethacin (18 mg/kg, p.o., single dose)-induced stomach ulceration in mice, reducing the

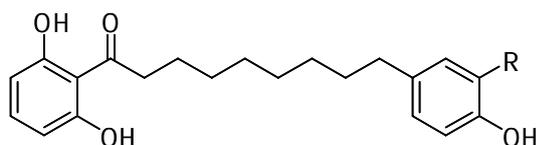
*Correspondence to: Dr. Subrata Chattopadhyay, Bio-Organic Division, Bhabha Atomic Research Centre, Mumbai 400 085, India; e-mail: schatt@barc.gov.in

ulcer index from 3.0 to 0.8 (~74%). The healing activity of RM could be partly attributed to its ability to prevent oxidative damages to lipids, thiols and antioxidant enzymes, as well as augmenting mucin status. Subsequently, malabaricones B and C (the chemical structures shown in Figure 1) were found as its active constituents (10). However, the extract was more potent than the individual malabaricones at their respective concentrations in the extract (9). Hence for the present study, we have used RM to understand the mechanisms of its healing action in terms of its capacity to regulate the arginine metabolism by modulating the balance of cytokines in the process. To this end, we have investigated the effect of RM in elevating arginase activity, and regulating NO production through modulation of NOS expression. In addition, the important role of the pro- and anti-inflammatory, as well as regulatory cytokines *vis-a-vis* their putative role during wound healing was also investigated.

2. Materials and Methods

2.1. Chemicals and reagents

The dry fruit rinds of *M. malabarica* were purchased from the local market. L-arginine, indomethacin, isonitrosopropiophenone, Bradford reagent, Triton X-100, leupeptin, phenylmethylsulfonyl fluoride (PMSF), glycine, sodium dodecyl sulfate (SDS), acrylamide, bis-acrylamide, Tween 20, ethylene diamine tetraacetic acid (EDTA), 3,3',5,5'-tetramethyl benzidine (TMB), MnCl₂, urea, omeprazole (Omez), Trizma base, cetyl trimethylammonium bromide (CTAB), and nitrocellulose membrane were procured from Sigma Chemicals (St. Louis, MO, USA). Other reagents used were disodium hydrogen phosphate and sodium dihydrogen phosphate (BDH, UK), sulphuric acid, hydrochloric acid, phosphoric acid, sodium chloride (Thomas Becker, Mumbai, India), horseradish peroxidase (HRPO, Sisco Research Laboratory, Mumbai, India), rabbit polyclonal iNOS and eNOS antibodies (Santacruz Biotechnology, Delaware, USA), peroxidase conjugated anti-rabbit IgG antibody, enhanced chemiluminescence detection kit (Roche, Mannheim, Germany), NOS and NO assay kits (Calbiochem, CA, USA), TGF-β1 set (Promega



Malabaricone B: R = H
Malabaricone C: R = OH

Figure 1. The chemical structures of malabaricone B and malabaricone C present in *M. malabarica*.

Corporation, Madison, USA) and cytokine ELISA kits (Pierce Biotechnology, Rockford, IL, USA).

2.2. Instrumentation

The absorbance spectrophotometry was carried out at 25°C using an ELISA reader (Biotech Instruments, USA). The bands obtained from western blots were quantified using the Gelquant software (DNR Bioimaging System, version 2.7.0, Israel).

2.3. Preparation of plant extract

The dry fruit rinds (20 g) of *M. malabarica* were powdered with a grinder and extracted successively with ether, methanol, and water (60 mL × 4 d with each solvent) at room temperature. The supernatants in each case were decanted. The entire process was repeated three times, each of the combined supernatants was filtered through a nylon mesh and evaporated at < 40° C *in vacuo* to obtain the respective extracts, which were stored in a vacuum desiccator. The methanol extract (designated as RM, 28.9%) was used for the present work.

2.4. Preparation of the drugs

The drugs were prepared from RM and Omez as aqueous suspensions in 2% gum acacia as the vehicle, and administered to the mice orally.

2.5. Protocol for ulceration and biochemical studies

The mice, bred at the BARC Laboratory Animal House Facility, Mumbai, India were procured after obtaining clearance from the BARC Animal Ethics Committee, and were handled following International Animal Ethics Committee Guidelines. Male swiss albino mice (25-30 g) were reared on a balanced laboratory diet as per NIN, Hyderabad, India and given tap water ad libitum. They were kept at 20 ± 2°C, 65-70% humidity, and day/night cycle (12 h/12 h). To perform all the experiments in a blinded fashion, the animals were identified by typical notches in the ear and limbs, and randomized, before the experiments. The mice were divided into four groups (each containing five mice), and each experiment was repeated three times. Ulceration was induced in the groups II-IV mice by administering indomethacin (18 mg/kg, p.o.) dissolved in distilled water and suspended in the vehicle, gum acacia (2%) as a single dose. The animals were deprived of food but had free access to tap water, 24 h before ulcer induction. The mice of groups I and II, serving as normal and ulcerated controls respectively were given the daily oral dose of vehicle (gum acacia in distilled water, 0.2 mL) only. The groups III and IV mice were given RM (40 mg/kg × 3 d, p.o.) and Omez (3 mg/kg

× 3 d, p.o.) respectively, starting the first dose 6 h post indomethacin administration. After 6 h of the last dose of the drugs, the mice were sacrificed after an overdose of thiopental, the stomach was opened along the greater curvature, and the wet weights of the tissues were recorded. The glandular portion from five animals were pooled, rinsed with appropriate buffer, homogenized in the same buffer under cold condition and used for assessing the expression of different NOSs and assaying arginase and MPO activities. Other biochemical (total NOS and NO) and immunological parameters were assayed using the serum samples.

2.6. Assessment of ulcer healing

The ulcerated portions of the stomach were fixed in 10% formol saline solution for 24 h, embedded in a paraffin block, and cut into 5 µm sections, which were placed onto glass slides, and stained with haematoxyline and eosin for histological examination under a light microscope. One centimetre lengths of each histological section was divided into three fields. The histological damage score (DS) was assessed as described previously (9).

2.7. Determination of myeloperoxidase (MPO) activity

The MPO activity was determined following a reported method (11) with slight modifications. Gastric ulcer tissues were homogenized for 30 sec in a 50 mM phosphate buffer (pH 6.0) containing 0.5% CTAB and 10 mM EDTA, followed by freeze thawing three times. The homogenate was centrifuged at 12,000 × g for 20 min at 4°C. The supernatant was collected, and the protein content determined. The supernatant (50 µL) was added to 80 mM phosphate buffer, pH 5.4 (250 µL), 0.03 M TMB (150 µL) and 0.3 M H₂O₂ (50 µL). After incubating the mixture at 25°C for 25 min, the reaction was terminated by adding 0.5 M H₂SO₄ (2.5 mL). The absorbance of the mixture at 450 nm was recorded using HRPO as the standard. The MPO activity was expressed as mU/mg protein.

2.8. Arginase assay

Following a known method (12) with minor modifications, the assay was carried out using the tissue homogenate prepared in ice-cold 25 mM Tris-HCl buffer (pH 7.5) followed by centrifugation at 12,000 × g for 30 min at 4°C. The reaction mixture (200 µL) containing 0.5 M L-arginine (pH 9.7), 1 mM MnCl₂, and the tissue extract (100 µL) was incubated for 20 min at 37.4°C. The reaction was stopped by adding an acid mixture (800 µL, H₂SO₄-H₃PO₄-H₂O, 1:3:7) and 3% isonitrosopropiophenone, followed by heating at 100°C for 45 min, and the absorbance at 540 nm was read. The data were quantified from a calibration curve prepared

using urea (1.5-120 µg), and normalized for tissue protein. One unit (U) of enzyme activity is defined as the amount of enzyme that catalyses the formation of 1 µmol of urea/min.

2.9. Total NOS assay

The serum NOS activity was measured using the commercially available colorimetric kit following manufacturer's protocol.

2.10. Western blot analyses of tissue iNOS and eNOS expressions

The glandular part of the gastric mucosa after being washed with PBS containing protease inhibitors was minced and homogenized in a lysis buffer (10 mM Tris-HCl, pH 8.0, 150 mM NaCl, 1% Triton X-100) containing leupeptin (0.005 µg/mL) and PMSF (0.4 µM/mL). Following centrifugation at 15,000 × g for 30 min at 4°C, the supernatant was collected, and the protein concentration measured. The proteins (40 µg) were resolved by 10% SDS-polyacrylamide gel electrophoresis and transferred to nitrocellulose membrane. The membrane was blocked for 2 h in TBST buffer (20 mM Tris-HCl, pH 7.4, 150 mM NaCl, 0.02% Tween 20) containing 99% fat-free milk powder and incubated overnight at 4°C with rabbit polyclonal iNOS or eNOS antibody. The membrane was washed over a period of 2 h with TBST and incubated with peroxidase conjugated anti-rabbit IgG (1:2,500 dilution). The bands were detected using an enhanced chemiluminescence detection kit and quantified using the Gelquant software.

2.11. Assay of NO

Following manufacturer's instruction, the serum NO level was measured using a commercially available colorimetric kit that measures the total nitrite concentration of the sample.

2.12. Assay of cytokines

The serum IL-4, IL-6 and TNF-α levels were estimated using commercially available ELISA kits following manufacturer's protocols. The method of TGF-β1 estimation (13) in sera was adopted after acidification to include the active and latent forms of the cytokine. Briefly, 96-well high binding ELISA plates were coated with anti-mouse TGF-β1 monoclonal antibody and incubated overnight at 4°C. After blocking for 30 min at 37°C, the wells were washed once with TBST buffer, the samples were activated by acid treatment followed by neutralization. The samples along with the standards were seeded to each well at an appropriate dilution, and incubated at room temperature for 90 min. The wells were washed (5 times), diluted polyclonal antibody

(100 μ L) added, and the mixture incubated further for 2 h at room temperature. The wells were washed, and incubated for 2 h after addition of TGF- β HRPO conjugate (100 μ L). After the final wash, TMB (100 μ L) was added to each well, the mixture incubated for 15 min, the reaction was stopped by 1 N HCl, and the absorbance at 450 nm was read.

2.13. Statistical analysis

The values are expressed as the mean \pm S.E.M. The data were analyzed by a paired Student's *t* test for the paired data, or one-way analysis of variance (ANOVA) followed by Dunnett's multiple comparisons post hoc test.

3. Results

Earlier we have observed peak ulceration in mice on the 3rd day after indomethacin (18 mg/kg, single dose) administration, and a three-day treatment with RM (40 mg/kg/day) and Omez (3 mg/kg/day) provided optimal ulcer healing (9). The extent of stomach ulceration and healing by the drugs were quantified from the same histological slide. The optimized healing data, assessed in terms of histological damage scores are provided in Table 1. Hence, the present experiments were also carried out under the above conditions. We preferred using the crude extract (RM) since it was more potent than the individual active components on equivalent basis. This would also be economically beneficial.

3.1. Regulation of the mucosal MPO activity

Compared to the normal mice, the mucosal MPO status in the gastric tissues of ulcerated untreated mice increased by 34.3% ($p < 0.05$) (Figure 2). This was brought down significantly by both RM (22.4%, $p < 0.05$) and Omez (20.3%, $p < 0.05$). The effect of Omez was not significantly different from that of RM.

3.2. Regulation of the mucosal arginase activity

The indomethacin-mediated stomach ulceration

Table 1. The optimized healing capacities of RM and Omez against indomethacin-induced stomach ulceration in mice as revealed from the histological damage scores^a

Group	Damage score
Untreated	3.0 \pm 0.07
Extract-treated	0.8 \pm 0.03 (73.3) ^b
Omez-treated	0.4 \pm 0.02 (86.7) ^b

^a Stomach ulceration in mice was induced by indomethacin (18 mg/kg, p.o.). RM (40 mg/kg \times 3 d) and Omez (3.0 mg/kg \times 3 d) were used for the experiments. The histologic damage (DS) were measured on the 3rd day after indomethacin administration by analyzing the data from the review of a minimum of three sections per animal. The values are mean \pm S.E.M. ($n = 15$). Figures in parenthesis show % reduction from the experimental control.

^b $p < 0.001$, compared to the 3rd day ulcerated control.

depleted (38.5%, $p < 0.05$) the arginase activity considerably, compared to the normal mice (Figure 3). Three-day treatment with RM and Omez enhanced it by 59.5% ($p < 0.01$) and 19.6% respectively compared to the untreated mice. The results of RM and Omez were significantly different ($p < 0.05$).

3.3. Regulation of the NOS activity

Compared to the normal mice, a significant increase (5.37 fold, $p < 0.001$) in the total NOS activity was noticed in the ulcerated mice. Both RM and Omez reduced it by \sim 73% ($p < 0.001$) (Figure 4).

3.4. Modulation of the mucosal eNOS and iNOS expressions

The Western blots of the eNOS and iNOS expressions

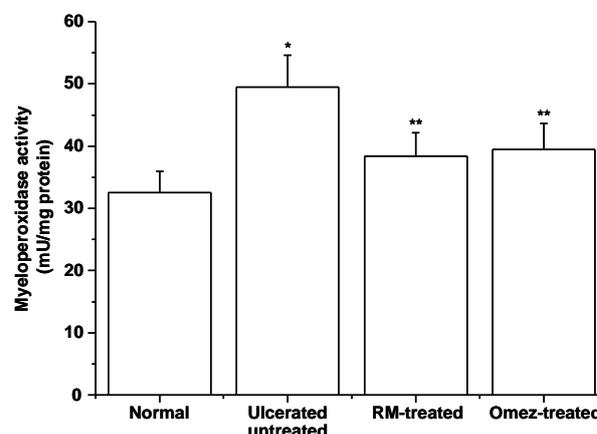


Figure 2. Effects of RM and Omez in modulating the mucosal MPO level in the indomethacin-induced ulcerated mice. The supernatant of the gastric tissue homogenate was incubated with TMB in a suitable buffer and the MPO activity (mean \pm S.E.M., $n = 15$) was assayed from the absorbance at 450 nm. * $p < 0.05$ compared to normal mice; ** $p < 0.05$ compared to untreated mice.

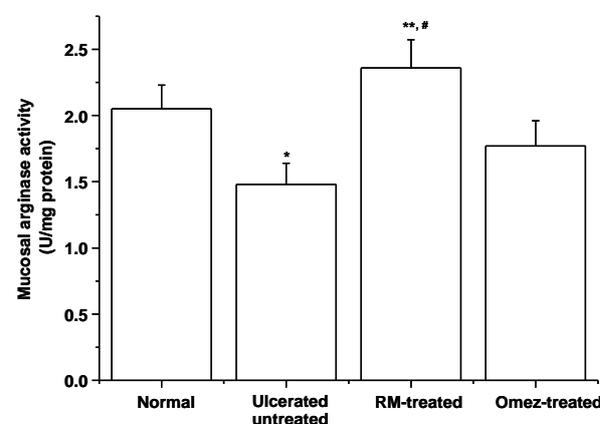


Figure 3. Effects of RM and Omez in modulating the mucosal arginase activity in the indomethacin-induced ulcerated mice. The supernatant of the gastric tissue homogenate was incubated with L-arginine and MnCl₂ in a suitable buffer and the arginase activity (mean \pm S.E.M., $n = 15$) was assayed from the absorbance at 540 nm. * $p < 0.05$ compared to normal mice; ** $p < 0.01$ compared to untreated mice; # $p < 0.01$ compared to Omez-treated mice.

in the gastric mucosa of the normal, ulcerated and drug (RM or Omez)-treated mice are shown in Figure 5. The eNOS expression was detected in both normal and ulcerated gastric tissues. In contrast, the iNOS expression was very high in the ulcerated tissues, but insignificant in normal gastric tissues. Quantification of the bands revealed that stomach ulceration increased the expressions of iNOS (62%, $p < 0.001$) and eNOS (44%, $p < 0.01$), compared to normal mice. Treatment with RM reduced the iNOS expression (21.6%, $p < 0.05$), along with dramatic increase in the eNOS expression (56.3%, $p < 0.001$), compared to the untreated mice. In contrast, Omez reduced the eNOS expression (23.6%, $p < 0.05$) significantly, but showed insignificant effect on the iNOS expression.

3.5. Regulation of the serum NO level

At peak ulceration, there was a significant increase (55.1%, $p < 0.01$) in the serum nitrite level compared to the normal mice. Treatment with RM and Omez reduced it by 49.8% ($p < 0.01$) and 29.3% ($p < 0.01$) respectively, the effect of RM being significantly better ($p < 0.05$) than that of Omez (Figure 6).

3.6. Modulation of the serum TGF- β 1 level

Compared to the normal value, ulceration reduced the level of serum TGF- β 1 (Figure 7) by 48% ($p < 0.001$). Treatment with RM and Omez, however, increased it by 79.6% ($p < 0.001$) and 26.3% ($p < 0.05$)

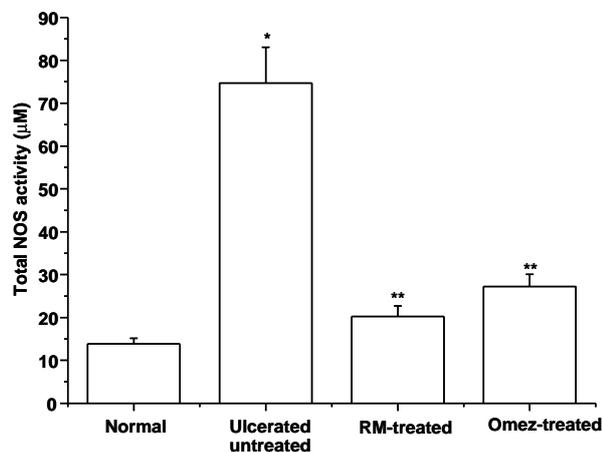


Figure 4. Effects of RM and Omez in regulating the serum total NOS activity in the indomethacin-induced ulcerated mice. The NOS level (mean \pm S.E.M., $n = 15$) was measured by ELISA. * $p < 0.001$ compared to normal mice; ** $p < 0.001$ compared to untreated mice.

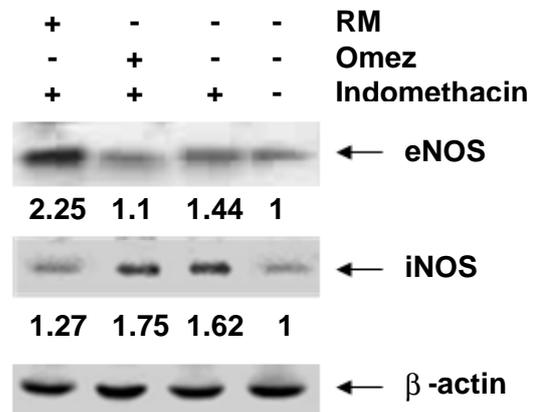


Figure 5. Western blots of eNOS and iNOS expressions of normal, ulcerated and drug-treated gastric tissues of mice. The numerical figures (arbitrary unit) reveals the density scanning results, considering that of normal mice as 1.

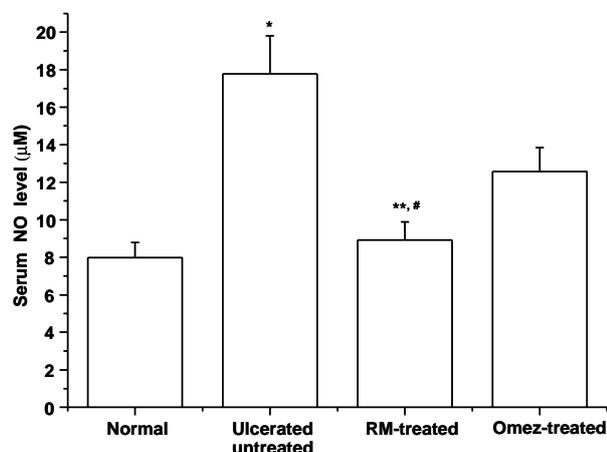


Figure 6. Effects of RM and Omez in regulating serum NO level in acute gastric ulcerated mice. The NO level (mean \pm S.E.M., $n = 15$) was measured by ELISA. * $p < 0.01$ compared to normal mice; ** $p < 0.01$ compared to the untreated mice; # $p < 0.05$ compared to Omez-treated mice.

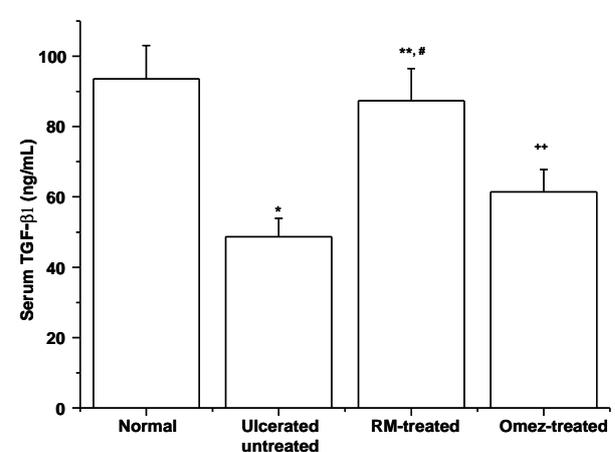


Figure 7. Effects of RM and Omez in regulating serum TGF β 1 level in acute gastric ulcerated mice. The serum TGF β 1 level (mean \pm S.E.M., $n = 15$) was measured by ELISA. * $p < 0.001$ compared to normal mice; ** $p < 0.001$, # $p < 0.05$ compared to untreated mice; * $p < 0.05$ compared to Omez-treated mice.

respectively, compared to the untreated mice. RM was significantly ($p < 0.05$) more potent than Omez.

3.7. Regulation of the serum Th1 (TNF- α and IL-6) and Th2 (IL-4) cytokines

Compared to the normal value, ulceration drastically increased the serum TNF- α and IL-6 ~ 4.4 and ~ 3.5 folds, respectively ($p < 0.001$). RM suppressed the levels of both TNF- α (66.9%, $p < 0.001$) and IL-6 56% ($p < 0.001$) significantly, compared to the untreated mice. Omez, however, reduced both the cytokines by $\sim 13.3\%$ ($p < 0.05$), which were much less than that by RM.

In contrast, the serum IL-4 level in the ulcerated mice was reduced by 28.3% ($p < 0.05$), compared to the normal mice. Treatment with RM improved it appreciably (95.4%, $p < 0.001$), while the effect of Omez (14.3% increase) was significantly less ($p < 0.001$) than that of RM. The results on the cytokine modulation are summarized in Figure 8.

4. Discussion

The non-steroidal anti-inflammatory drugs (NSAIDs) including indomethacin are most widely prescribed for the treatment of pain and inflammation. However,

they are also known to cause gastrointestinal (GI) damage, characterized by hyperemia, and increased vascular permeability, as well as delayed ulcer healing (14). Ulcer-healing is a complex process involving a combination of wound retraction and re-epithelization wherein several factors such as enzymes, cytokines, and soluble mediators, liberated during the inflammatory response play crucial roles. The impressive healing capacity of RM ($IC_{50} = 23.30 \pm 3.50$ mg/kg) against the indomethacin-induced gastric ulceration in mice (9) encouraged us to investigate its probable modulatory effect on arginase and NOS as well as the Th1/Th2 cytokines profiles since these are some of the established mediators of wound healing.

Earlier, we have established the healing action of mal B by histology (11). Quantification of the histological slides in terms of damage score (DS) provided a better assessment of the quality of healing. This was also substantiated from our results with the MPO assay. The MPO activity, a marker of neutrophil aggregation at the site of inflammation is frequently increased in ulcerated conditions, and reduced during wound healing (15). Our studies depicted that while indomethacin administration enhanced the gastric mucosal MPO activity, treatment with RM (40 mg/kg \times 3 d) and Omez (3 mg/kg \times 3 d) reduced it almost equally. These results are consistent with our previous

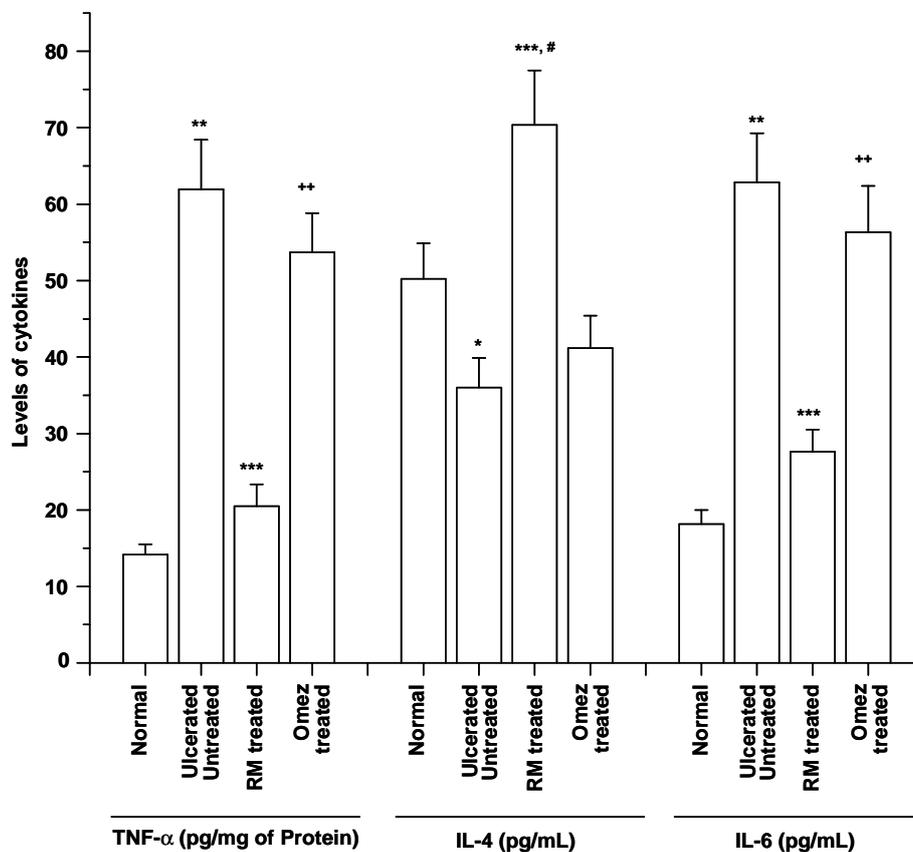


Figure 8. Effects of RM and Omez in regulating serum TNF- α , IL-4 and IL-6 levels in acute gastric ulcerated mice. The cytokine levels (mean \pm S.E.M., $n = 15$) were measured by ELISA. * $p < 0.05$, ** $p < 0.001$ compared to normal mice; + $p < 0.05$, *** $p < 0.001$ compared to untreated mice; # $p < 0.001$ compared to Omez-treated mice.

histological results, where both RM and Omez were found to produce similar ulcer healing at the designated doses (9). It is worth mentioning that extending the period of treatment led to slightly better healing, but a large part of this was due to natural healing. Since that would not provide much information on the drug action, this was not pursued. Likewise, the effect of increasing daily dose of RM was only marginal. The phytochemical analysis of RM revealed the presence of four diarylnonanoids, malabaricones A-D as its major constituents. All these compounds contain a resorcinol moiety, which is known to induce irreversible, hydrogen peroxide-dependent loss of activities of the heme-containing peroxidases such as MPO (16).

Metabolism of arginine that can be catalyzed by arginase, and NOS, plays a vital role in gastric ulceration and its healing. Upregulation of arginase increases the level of polyamines, which play a significant role in wound healing. The regulatory role of arginase in acute intestinal inflammation and tissue repair has been demonstrated (17,18). On the other hand, NOS catabolizes L-arginine by a different pathway to produce L-citrulline and NO. The signaling molecule, NO has long been the subject of extensive research with respect to its role in GI mucosal defense and the pathogenesis of mucosal injury. NOSs exist as constitutive (cNOS), and inducible isoforms (iNOS). Of these, the endothelial NOS (eNOS), belonging to the cNOS isoforms generates only nanomolar concentrations of NO that dilates the blood vessels and increases blood flow in the gastric mucosa (19) and also contributes to angiogenesis (20,21). The eNOS expression plays a major role in wound healing that gets inhibited if the eNOS expression is depleted (20,22). On the other hand, iNOS that can be largely induced under certain pathological conditions (23) acts as a killer molecule, and is involved in inflammation. The enhanced generation of NO in the inflamed gastric mucosa may be toxic in the digestive systems, and contribute to the pathogenesis of peptic ulcer disease (24). An increase in iNOS activity, and a decrease in eNOS activity in the gastric mucosa are closely related to the development of gastric mucosal lesions. Because the potential high-output source of NO in mammalian cells is iNOS, factors involved in the induction and expression of iNOS activity are key determinants of the NO-mediated toxicity. Sustained overproduction of NO by iNOS is detrimental and contributes to inflammation in various gastroduodenal disorders (15).

The intense reciprocal regulation of NOS and arginase activities *in vivo*, and the temporal switch between them decides ulceration and healing (8,25). Our results showed considerable down regulation of the mucosal arginase level along with an increased expression of the iNOS due to ulceration. This suggested a shift of the arginine metabolism towards the NO/iNOS pathway during ulceration. The elevated

expressions of both iNOS and eNOS accounted for the increased total NOS activity as well as serum nitrite level due to ulceration. The iNOS is probably derived from inflammatory neutrophils and macrophages that would contribute to stimulate inflammatory situation, explaining the ulcerogenic property of indomethacin. Simultaneous generation of superoxide and NO by the macrophages would produce peroxynitrite creating oxidative stress. Its stimulated generation, as observed in this study, may also delay proximal duodenal contractions, by exposing mucus to gastric acidity.

Treatment with RM and Omez, especially the former restored the arginase activity almost to normalcy. RM also raised the eNOS/iNOS ratio to a level favorable for efficient ulcer-healing. This would amount to generation of more polyamine at the expense of the iNOS-derived NO that may be a key contributing factor in the anti-ulcer effect of RM. The reduction of the total NOS activity and NO level by RM was primarily due to suppression of the iNOS expression. Even a moderate suppression of iNOS expression would reduce NO release substantially, since the enzyme is capable of generating NO many folds.

In contrast, despite showing less effect on modulating eNOS/iNOS expressions and NO production, Omez provided excellent healing. This may be due to other operative mechanism in its healing action as observed by us and others (10,26). The total NOS reduction by Omez was due to less eNOS expression. This was also evident from the fact that its effect on NO reduction was much less.

Although iNOS is very strongly induced in ulcerated tissue in the stomach, the NO-derived from it does not appear to play a role in modulating healing. Most of the previous studies of NO and inflammation focused on the role of iNOS, whereas the role of eNOS that can also be markedly enhanced by various stimuli or tissue injury has been underestimated. Using eNOS deficient mice, the importance of eNOS and eNOS-derived NO in regulating microvascular structure during acute inflammation has been demonstrated (27). Our results suggested that the eNOS-derived NO contributed maximum to the ulcer healing property of RM, although a role for neuronal NOS-derived NO cannot be excluded.

Stimulation of inflammatory cytokines is extremely important in mucosal defense. One of the most prominent modes of mediation of indomethacin-induced gastropathy is the increased expression of the pro-inflammatory cytokines (28,29), which also correlates with the extent of ulceration. Even the cross-talk amongst NOS/NO and arginase/polyamine is guided by the cytokine profile of the host (2,3). In view of this, the immune response due to ulceration, and its modulation by RM and Omez was monitored. This enabled us to associate the inflammatory response with a better prognosis.

Indomethacin administration raised the levels of pro-inflammatory cytokines (TNF- α and IL-6) while reducing the anti-inflammatory cytokines (IL-4 and TGF- β), thereby creating a cytokine imbalance. Increased TNF- α is known to increase iNOS activity by promoting binding of NF- κ B to the iNOS promoter (25,30). The increased levels of Th1 cytokines due to ulceration would augment the iNOS/NO pathway to produce excess NO, which is likely to promote oxidative stress and result in ulceration (31,32). Our result on the decreased IL-4 level due to ulceration was in tune with a previous report (33).

Treatment with RM, however, restored the imbalance by increasing the levels of IL-4 and TGF- β beyond the normal values. The upregulation of the anti-inflammatory cytokines by RM is likely to inhibit the stimulatory effect of indomethacin on the level of pro-inflammatory cytokine release in blood and gastric mucosa. The immunosuppressive Th2 cytokine, TGF- β has a direct role in stimulating epithelial restitution (34). Besides suppressing the IFN- γ -induced iNOS gene expression and thereby generation of excess NO, it also increases arginase activity during inflammatory processes (8,25,35). The altered arginase activity and iNOS expressions observed by us during ulceration, and RM treatment are consistent with their respective effects in modulating the mucosal TGF- β status. The enhanced IL-4 level by RM would trigger the TGF- β -SMAD-signaling pathway to stimulate the extracellular remodeling and subsequent tissue repair. In contrast, except for the TGF- β , the other cytokines were not affected significantly by Omez, as reported earlier (33). This was also reflected in its marginal effect in regulating the enzymes, arginase and NOS.

Overall, RM modulated the cytokine profile to shift the balance in favour of arginase/polyamine *vis-a-vis* iNOS/NO pathway, besides improving the eNOS expression. A combination of all these events might tilt the balance in favour of the repair mechanisms, explaining its ulcer-healing action. The bimodal nature of general immune responses is explained by the Th1/Th2 paradigm (36). The regulatory T cells and Th2 cytokines often collaborate to suppress the Th1 response. Perhaps even more importantly, they strongly promote the mechanism of wound healing. However, the role of cytokine imbalance in gastropathy has not been adequately emphasized. Our results highlighted that the balance of the pro- and anti-inflammatory, as well as regulatory cytokines could play a significant role in the NSAID-induced gastric mucosal injury.

References

1. Glavin GB, Szabo S. Experimental gastric mucosal injury: laboratory models reveal mechanisms of pathogenesis and new therapeutic strategies. *FASEB J* 1992; 6:825-831.
2. Satriano J. Arginine: At the crossroads of the arginine pathways. *Ann NY Acad Sci* 2003; 1009:34-43.
3. Jenkinson CP, Grody WW, Cederbaum SD. Comparative properties of arginases. *Comp Biochem Physiol* 1996; 114B:107-132.
4. Bogdan C. Nitric oxide and the immune response. *Nat Immunol* 2001; 2:907-916.
5. Caldwell MD, Mastrofrancesco B, Shearer J, Bereiter D. The temporal change in amino acid concentration within wound fluid-A putative rationale. *Prog Clin Biol Res* 1991; 365:205-222.
6. Witte MB, Barbul A. Role of nitric oxide in wound repair. *Am J Surg* 2002; 183:406-412.
7. Calatayud S, Barrachina MD, Esplugues JV. Nitric oxide: Relation to integrity, injury, and healing of the gastric mucosa. *Microscop Res Techniq* 2001; 53:325-335.
8. Modolell M, Corraliza IM, Link F, Soler G, Eichmann K. Reciprocal regulation of the nitric oxide synthase/arginase balance in mouse bone marrow-derived macrophages by TH1 and TH2 cytokines. *Eur J Immunol* 1995; 25:1101-1104.
9. Banerjee D, Maity B, Bandyopadhyay SK, Bauri AK, Chattopadhyay S. Gastroprotective properties of *Myristica malabarica* against indomethacin-induced stomach ulceration: a mechanistic exploration. *J Pharm Pharmacol* 2007; 59:1555-1565.
10. Banerjee D, Bauri AK, Guha R, Bandyopadhyay SK, Chattopadhyay S. Healing properties of malabaricone B and malabaricone C, against indomethacin-induced gastric ulceration and mechanism of action. *Eur J Pharmacol* 2008; 578:300-312.
11. Suzuki K, Ota H, Sasagawa S, Sakatani T, Fujikura T. Assay method for myeloperoxidase in human polymorphonuclear leukocytes. *Anal Biochem* 1983; 132:345-352.
12. del Ara Rangel M, Gozalez-Polo RA, Caro A, del-Amo E, Palomo L, Soler EGH, Fuentes JM. Diagnostic performance of arginase activity in colorectal cancer. *Clin Exp Med* 2002; 2:53-57.
13. Thakur VS, Shankar B, Chatterjee S, Premachandran S, Sainis KB. Role of tumor derived transforming growth factor-beta 1(TGF β 1) in site-dependant tumorigenicity of murine ascitic lymphosarcoma. *Cancer Immunol Immunother* 2005; 54:837-847.
14. Fiorucci S, Antonelli E, Morelli A. Mechanism of non-steroidal anti-inflammatory drug-gastropathy. *Dig Liv Dis* 2001; 31 (suppl 2):S35-S43.
15. Souza MHL, Lemos HP, Oliveira RB, Cunha FQ. Gastric damage and granulocyte infiltration induced by indomethacin in tumour necrosis factor receptor 1 (TNF-R1) or inducible nitric oxide synthase (iNOS) deficient mice. *Gut* 2004; 53:791-796.
16. Forbes E, Murase T, Yang M, Matthaehi KI, Lee JJ, Lee NA, Foster PS, Hogan SP. Immunopathogenesis of experimental ulcerative colitis is mediated by eosinophil peroxidase. *J Immunol* 2004; 172:5664-5675.
17. Bernard AC, Mistry SK, Morris Jr SM, O'Brien WE, Tsuei BJ, Maley ME, Shirley LA, Kearney PA, Boulanger BR, Ochoa JB. Alterations in arginine metabolic enzymes in trauma. *Shock* 2001; 15:215-219.
18. Satriano J. Interregulation of nitric oxide and polyamines. *Amino Acids* 2004; 26:321-329.
19. Whittle BJR. In: *The Physiology of the gastrointestinal tract* (Johnson LR, ed.), Raven Press, New York, USA, 1994; pp. 267-294.

20. Ma L, Wallace JL. Endothelial nitric oxide synthase modulates gastric ulcer healing in rats. *Am J Physiol Gastrointest Liver Physiol* 2000; 279:G341-G346.
21. Ziche M, Morbidelli L, Masini E, Amerini S, Granger HJ, Maggi CA. Nitric oxide mediates angiogenesis *in vivo* and endothelial cell growth and migration *in vitro* promoted by substance P. *J Clin Invest* 1994; 94:2036-2044.
22. Konturek SJ, Brzozowski T, Majka J, Pytko-Polonczyk J, Stachura J. Inhibition of nitric oxide synthase delays healing of chronic gastric ulcers. *Eur J Pharmacol* 1993; 239:215-217.
23. Anggård E. Nitric oxide: mediator, murderer, and medicine. *Lancet* 1994; 343:1199-1206.
24. Jaiswal M, LaRusso NF, Gores GJ. Nitric oxide in gastrointestinal epithelial cell carcinogenesis: linking inflammation to oncogenesis. *Am J Physiol Gastrointest Liver Physiol* 2001; 281:G626-G634.
25. Shearer JD, Richards JR, Mills CD, Caldwell MD. Differential regulation of macrophage arginine metabolism: a proposed role in wound healing. *Am J Physiol* 1997; 272:E181.
26. Ng KM, Cho CH, Chang FY, Luo JC, Lin HC, Lin HY, Chi CW, Lee SD. Omeprazole promotes gastric epithelial cell migration. *J Pharm Pharmacol* 2008; 60:655-660.
27. Luo JC, Shin VY, Liu ESL, So WHL, Ye YN, Chang FY, Cho CH. Non-ulcerogenic dose of dexamethasone delays gastric ulcer healing in rats. *J Pharm Exp Therap* 2003; 307:692-698.
28. Yoshikawa T, Naito Y, Kishi A, Tomii T, Kaneko T, Iinuma S, Ichikawa H, Yasuda M, Takahashi S, Kondo M. Role of active oxygen, lipid peroxidation, and antioxidants in the pathogenesis of gastric mucosal injury induced by indomethacin in rats. *Gut* 1993; 34:732-737.
29. Brzozowski T, Konturek PC, Konturek SJ, Sliwowski Z, Pajdo R, Drozdowicz D, Ptak A, Hahn EG. Classic NSAID and selective cyclooxygenase (COX)-1 and COX-2 inhibitors in healing of chronic gastric ulcers. *Microscop Res Tech* 2001; 43:343-353.
30. Titheradge MA. Nitric oxide in septic shock. *Biochim Biophys Acta* 1999; 1411:437-455.
31. Chatterjee S, Premachandran S, Bagewadikar RS, Bhattacharya S, Chattopadhyay S, Poduval TB. Arginine metabolic pathways determine its therapeutic benefit in experimental heat stroke: Role of Th₁/Th₂ cytokine balance. *Nitric Oxide* 2006; 15:408-416.
32. Murphy MP. Nitric oxide and cell death. *Biochim Biophys Acta* 1999; 1411:401-414.
33. Slomiany BL, Piotrowski J, Slomiany A. Role of endothelin-1 and constitutive nitric oxide synthase in gastric mucosal resistance to indomethacin injury: effect of antiulcer agents. *Scand J Gastroenterol* 1999; 34:459-464.
34. Kaviratne M, Hesse M, Leusink M, Cheever AW, Davies SJ, McKerrow JH, Wakefield LM, Letterio JJ, Wynn TA. IL-13 activates a mechanism of liver fibrosis that is completely TGF- β independent. *J Immunol* 2004; 173:4020-4029.
35. Mitani T, Terashima M, Yoshimura H, Nariai Y, Tanigawa Y. TGF- β 1 enhances degradation of IFN- γ induced iNOS protein *via* proteasomes in RAW 264.7 cells. *Nitric Oxide* 2005; 13:78-87.
36. Abbas AK, Murphy KM, Sher A. Functional diversity of helper T lymphocytes. *Nature* 1996; 383:787-793.

(Received November 14, 2008; Accepted November 22, 2008)

Drug Discoveries & Therapeutics

Guide for Authors

1. Scope of Articles

Drug Discoveries & Therapeutics mainly publishes articles related to basic and clinical pharmaceutical research such as pharmaceutical and therapeutical chemistry, pharmacology, pharmacy, pharmacokinetics, industrial pharmacy, pharmaceutical manufacturing, pharmaceutical technology, drug delivery, toxicology, and traditional herb medicine. Studies on drug-related fields such as biology, biochemistry, physiology, microbiology, and immunology are also within the scope of this journal.

2. Submission Types

Original Articles should be reports new, significant, innovative, and original findings. An Article should contain the following sections: Title page, Abstract, Introduction, Materials and Methods, Results, Discussion, Acknowledgments, References, Figure legends, and Tables. There are no specific length restrictions for the overall manuscript or individual sections. However, we expect authors to present and discuss their findings concisely.

Brief Reports should be short and clear reports on new original findings and not exceed 4000 words with no more than two display items. *Drug Discoveries & Therapeutics* encourages younger researchers and doctors to report their research findings. Case reports are included in this category. A Brief Report contains the same sections as an Original Article, but Results and Discussion sections must be combined.

Reviews should include educational overviews for general researchers and doctors, and review articles for more specialized readers.

Policy Forum presents issues in science policy, including public health, the medical care system, and social science. Policy Forum essays should not exceed 2,000 words.

News articles should not exceed 500 words including one display item. These articles should function as an international news source with regard to topics in the life and social sciences and medicine. Submissions are not restricted to journal staff - anyone can submit news articles on subjects that would be of interest to *Drug Discoveries & Therapeutics*' readers.

Letters discuss material published in *Drug Discoveries & Therapeutics* in the last 6 months or issues of general interest. Letters should not exceed 800 words and 6 references.

3. Manuscript Preparation

Preparation of text. Manuscripts should be written in correct American English and submitted as a Microsoft Word (.doc) file in a single-column format. Manuscripts must be paginated and double-spaced throughout. Use Symbol font for all Greek characters. Do not import the figures into the text file but indicate their approximate locations directly on the manuscript. The manuscript file should be smaller than 5 MB in size.

Title page. The title page must include 1) the title of the paper, 2) name(s) and affiliation(s) of the author(s), 3) a statement indicating to whom correspondence and proofs should be sent along with a complete mailing address, telephone/fax numbers, and e-mail address, and 4) up to five key words or phrases.

Abstract. A one-paragraph abstract consisting of no more than 250 words must be included. It should state the purpose of the study, basic procedures used, main findings, and conclusions.

Abbreviations. All nonstandard abbreviations must be listed in alphabetical order, giving each abbreviation followed by its spelled-out version. Spell out the term upon first mention and follow it with the abbreviated form in parentheses. Thereafter, use the abbreviated form.

Introduction. The introduction should be a concise statement of the basis for the study and its scientific context.

Materials and Methods. Subsections under this heading should include sufficient instruction to replicate experiments, but well-established protocols may be simply referenced. *Drug Discoveries & Therapeutics* endorses the principles of the Declaration of Helsinki and expects that all research involving humans will have been conducted in accordance with these principles. All laboratory animal studies must be approved by the authors' Institutional Review Board(s).

Results. The results section should provide details of all of the experiments that are required to support the conclusions of the paper. If necessary, subheadings may be used for an orderly presentation. All figures, tables, and photographs must be referred in the text.

Discussion. The discussion should include conclusions derived from the study and supported by the data. Consideration should be given to the impact that these conclusions have on the body of knowledge in which context the experiments were conducted. In Brief Reports, Results and Discussion sections must be combined.

Acknowledgments. All funding sources should be credited in the Acknowledgments section. In addition, people who contributed to the work but who do not fit the criteria for authors should be listed along with their contributions.

References. References should be numbered in the order in which they appear in the text. Cite references in text using a number in parentheses. Citing of unpublished results and personal communications in the reference list is not recommended but these sources may be mentioned in the text. For all references, list all authors, but if there are more than fifteen authors, list the first three authors and add "et al." Abbreviate journal names as they appear in PubMed. Web references can be included in the reference list.

Example 1:

Hamamoto H, Kamura K, Razanajatovo IM, Murakami K, Santa T, Sekimizu K. Effects of molecular mass and hydrophobicity on transport rates through non-specific pathways of the silkworm

larva midgut. *Int J Antimicrob Agents* 2005; 26:38-42.

Example 2:

Mizuochi T. Microscale sequencing of N-linked oligosaccharides of glycoproteins using hydrazinolysis, Bio-Gel P-4, and sequential exoglycosidase digestion. In: *Methods in Molecular Biology: Vol. 14 Glycoprotein analysis in biomedicine* (Hounsell T, ed.). Humana Press, Totowa, NJ, USA, 1993; pp. 55-68.

Example 3:

Drug Discoveries & Therapeutics. Hot topics & news: China-Japan Medical Workshop on Drug Discoveries and Therapeutics 2007. <http://www.ddtjournal.com/hotnews.php> (accessed July 1, 2007).

Figure legends. Include a short title and a short explanation. Methods described in detail in the Materials and methods section should not be repeated in the legend. Symbols used in the figure must be explained. The number of data points represented in a graph must be indicated.

Tables. All tables should have a concise title and be typed double-spaced on pages separate from the text. Do not use vertical rules. Tables should be numbered with Roman numerals consecutively in accordance with their appearance in the text. Place footnotes to tables below the table body and indicate them with lowercase superscript letters.

Language editing. Manuscripts submitted by authors whose primary language is not English should have their work proofread by a native English speaker before submission. The Editing Support Organization can provide English proofreading, Japanese-English translation, and Chinese-English translation services to authors who want to publish in *Drug Discoveries & Therapeutics* and need assistance before submitting an article. Authors can contact this organization directly at <http://www.iacmhr.com/iac-eso>.

IAC-ESO was established in order to facilitate manuscript preparation by researchers whose native language is not English and to help edit work intended for

international academic journals. Quality revision, translation, and editing services are offered by our staff, who are native speakers of particular languages and who are familiar with academic writing and journal editing in English.

4. Figure Preparation

All figures should be clear and cited in numerical order in the text. Figures must fit a one- or two-column format on the journal page: 8.3 cm (3.3 in.) wide for a single column; 17.3 cm (6.8 in.) wide for a double column; maximum height: 24.0 cm (9.5 in.). Only use the following fonts in the figure: Arial and Helvetica. Provide all figures as separate files. Acceptable file formats are JPEG and TIFF. Please note that files saved in JPEG or TIFF format in PowerPoint lack sufficient resolution for publication. Each Figure file should be smaller than 10 MB in size. Do not compress files. A fee is charged for a color illustration or photograph.

5. Online Submission

Manuscripts should be submitted to *Drug Discoveries & Therapeutics* online at <http://www.ddtjournal.com>. The manuscript file should be smaller than 10 MB in size. If for any reason you are unable to submit a file online, please contact the Editorial Office by e-mail: office@ddtjournal.com.

Editorial and Head Office

Wei TANG, MD PhD
Secretary-in-General
TSUIN-IKIZAKA 410
2-17-5 Hongo, Bunkyo-ku
Tokyo 113-0033
Japan
Tel: 03-5840-9697
Fax: 03-5840-9698
E-mail: office@ddtjournal.com

Cover letter. A cover letter from the corresponding author including the following information must accompany the submission: name, address, phone and fax numbers, and e-mail address of the corresponding author. This should include a statement affirming that all authors concur with the submission and that the material submitted for publication has not been previously published and is not under consideration for publication elsewhere and a

statement regarding conflicting financial interests.

Authors may recommend up to three qualified reviewers other than members of Editorial board. Authors may also request that certain (but not more than three) reviewers not be chosen.

The cover letter should be submitted as a Microsoft Word (.doc) file (smaller than 1 MB) at the same time the work is submitted online.

6. Accepted Manuscripts

Proofs. Rough galley proofs in PDF format are supplied to the corresponding author via e-mail. Corrections must be returned within 4 working days of receipt of the proofs. Subsequent corrections will not be possible, so please ensure all desired corrections are indicated. Note that we may proceed with publication of the article if no response is received.

Transfer of copyrights. Upon acceptance of an article, authors will be asked to agree to a transfer of copyright. This transfer will ensure the widest possible dissemination of information. A letter will be sent to the corresponding author confirming receipt of the manuscript. A form facilitating transfer of copyright will be provided. If excerpts from other copyrighted works are included, the author(s) must obtain written permission from the copyright owners and credit the source(s) in the article.

Cover submissions. Authors whose manuscripts are accepted for publication in *Drug Discoveries & Therapeutics* may submit cover images. Color submission is welcome. A brief cover legend should be submitted with the image.

Revised February 2008



Drug Discoveries & Therapeutics



Editorial Office

TSUIN-IKIZAKA 410
2-17-5 Hongo, Bunkyo-ku
Tokyo 113-0033, Japan

Tel: 03-5840-9697
Fax: 03-5840-9698
E-mail: office@ddtjournal.com
URL: www.ddtjournal.com

JOURNAL PUBLISHING AGREEMENT

Ms No:

Article entitled:

Corresponding author:

To be published in Drug Discoveries & Therapeutics

Assignment of publishing rights:

I hereby assign to International Advancement Center for Medicine & Health Research Co., Ltd. (IACMHR Co., Ltd.) publishing Drug Discoveries & Therapeutics the copyright in the manuscript identified above and any supplemental tables and illustrations (the articles) in all forms and media, throughout the world, in all languages, for the full term of copyright, effective when and if the article is accepted for publication. This transfer includes the rights to provide the article in electronic and online forms and systems.

I understand that I retain or am hereby granted (without the need to obtain further permission) rights to use certain versions of the article for certain scholarly purpose and that no rights in patent, trademarks or other intellectual property rights are transferred to the journal. Rights to use the articles for personal use, internal institutional use and scholarly posting are retained.

Author warranties:

I affirm the author warranties noted below.

- 1) The article I have submitted to the journal is original and has not been published elsewhere.
- 2) The article is not currently being considered for publication by any other journal. If accepted, it will not be submitted elsewhere.
- 3) The article contains no libelous or other unlawful statements and does not contain any materials that invade individual privacy or proprietary rights or any statutory copyright.
- 4) I have obtained written permission from copyright owners for any excerpts from copyrighted works that are included and have credited the sources in my article.
- 5) I confirm that all commercial affiliations, stock or equity interests, or patent-licensing arrangements that could be considered to pose a financial conflict of interest regarding the article have been disclosed.
- 6) If the article was prepared jointly with other authors, I have informed the co-authors(s) of the terms of this publishing agreement and that I am signing on their behalf as their agents.

Your Status:

- I am the sole author of the manuscript.
 I am one author signing on behalf of all co-authors of the manuscript.

Please tick one of the above boxes (as appropriate) and then sign and date the document in black ink.

Signature:

Date:

Name printed:

Please return the completed and signed original of this form by express mail or fax, or by e-mailing a scanned copy of the signed original to:

Drug Discoveries & Therapeutics office
TSUIN-IKIZAKA 410, 2-17-5 Hongo,
Bunkyo-ku, Tokyo 113-0033, Japan
E-mail: proof-editing@ddtjournal.com
Fax: +81-3-5840-9698

